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The NDE1 genomic locus can affect treatment of psychiatric illness through gene expression changes related to microRNA-484

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cohort, in which DISC1 (disrupted in schizophrenia 1) was previously associated with schizophrenia [1,2], and which led to observation of association with four other genes (NDE1, NDELI, PDE4B and PDE4D) [3,4] that encode protein binding partners of the DISC1 protein [5–8]. The idea that such protein interaction partners of DISC1 are encoded for by genes which show genetic interaction in mental illness is termed the DISC1 network hypothesis. Specifically, multiple associations for interaction partners of DISC1 are encoded for by genes which partners of the DISC1 protein [5–8]. The idea that such protein


...phenotypes, including memory [12], cognitive and neuroimaging [13] phenotypes, have been reported for DISC1 in Finnish cohorts. By conditioning genome-wide linkage data for schizophrenia on DISC1, a peak of linkage at chromosome 16p was observed [3], near to NDE1 (nuclear distribution element 1). This was followed up through association analysis at the NDE1 locus, leading to the observation that a haplotype and its constituent SNPs associate with schizophrenia in this cohort, in a gender-dependent manner [3]. Genetic association for schizophrenia was, therefore, tested for other DISC1 binding partners in this family cohort [4]. Although SNPs and haplotypes from six other genes were initially observed to associate, only variants in NDE1L (NDE-like 1, a close paralogue of NDE1) and in the phosphodiesterases PDE4B and PDE4D replicated when tested in a second, distinct sample from the cohort [4]. Recently, through further investigation of the roles of these variants in the DISC1 network, the NDE1 locus has been identified to increase risk of schizophrenia in this Finnish family cohort through interaction with high birth weight, a promising proxy measure for multiple pre- and/or perinatal environmental factors [14].

The role of the DISC1 network as a source of genetic risk for neuropsychiatric disorders is controversial due to the absence, to date, of evidence for their involvement in population based genomic studies of common variation [15,16]. However, these genes have been implicated at least within specific populations through strong evidence emerging from family based approaches and the studies of rare variants. In addition to the evidence from the Finnish family cohort, the DISC1 and PDE4B genes are disrupted by chromosomal aberrations in Scottish families with major mental illness [8,17,18]. Furthermore, NDE1 is independently implicated in major mental illness through its presence at 16p13.11, which is subject to duplications in schizophrenia [19–22], as well as being directly implicated through rare SNPs in patients [23]. The importance of the NDE1 protein for neurodevelopment more generally has been dramatically demonstrated in individuals with biallelic loss of the functional NDE1 gene, leading to severe microcephaly phenotypes, sometimes described in conjunction with lissencephaly or hydrocephaly [24–27]. Deletion of only one copy of the 16p13.11 locus, meanwhile, has been associated with neurological conditions including autism and epilepsy [28]. Recently it was shown that expression of mature miR-484, a microRNA that is encoded on an untranslated exon of NDE1, led to alterations in neural progenitor proliferation and differentiation, as well as behavioural changes in mice, thus implicating the microRNA in the phenotypes associated with 16p13.11 duplication [29].

We have previously studied the effect of DISC1 network genetic variation on gene expression in a small (n = 30) publicly available population cohort of the CEU (Utah residents with North and Western European ancestry) individuals. Using gene expression measured on the same individuals, by different laboratories, we were able to observe 528 genes as being replicable in their differential expression across 24 variants studied, of which 35 genes had pre-existing supporting evidence for a role in psychosis [30]. Intriguingly, seven of these affected genes were noted to be targets for drugs prescribed for psychiatric illness, leading to the hypothesis that these DISC1 network variants, through their action on gene expression, may alter treatment outcome for medications designed to target these genes [30].

Here, in order to advance our understanding of the role these genes play in the aetiology of schizophrenia in Finland, we take this approach further. This is accomplished by using data on gene expression levels in case families in which these DISC1 network genetic variants have been previously demonstrated to associate with schizophrenia [1–4], as well as by using data collected on how different psychotropic medications are used by the affected individuals within these families.

2. Material and methods

2.1. Study samples

The principal samples used here are part of a larger study of familial schizophrenia. These are Finnish patients born between 1940 and 1976, who were identified through the hospital discharge, disability pension and free medication registers [1,31]. The cohort totals 458 families (498 nuclear families) that contain 2756 individuals, of whom 2099 have been previously genotyped for the DISC1 network genes [1–4]. Of these genotyped individuals, 931 are classified as affected with major mental illnesses using criteria from the Diagnostic and Statistical Manual of Mental Disorders, fourth edition (DSM-IV) [32]. These include 635 diagnosed with schizophrenia, 125 with schizoaffective disorder, 95 with schizophrenia spectrum diagnoses and 76 with other mental illnesses, including bipolar disorder and major depression. Here, two sub-sets of this larger familial sample were used as discovery cohorts, firstly a sample of 18 families (64 individuals, 23 broadly affected: 15 with schizophrenia, three with schizoaffective disorder, three with schizophrenia spectrum diagnoses, and two with other mental illnesses; 41 unaffected family members) for whom RNA was available to study gene expression level changes, and secondly all 931 broadly affected individuals were used to study medicare use.

In order to replicate the gene expression results obtained from this family data, two independent cohorts were used. The first of these replication cohorts was a Finnish discordant twin pair sample ascertained for schizophrenia (N = 73), for which information about recruitment and clinical evaluation has been described previously [33]. Briefly, the participants are 18 schizophrenia patients, their 18 unaffected co-twins and 37 control twins who have provided blood samples for gene expression analysis (N = 73). The second replication cohort was the Genotype-Tissue Expression (GTEX) database (N = 338), a publicly available resource for exploring the correlation between genotypes and gene expression across multiple tissues and in a genome-wide manner (accessed on 2 September 2016, www.gtexportal.org/home) [34]. To best match the source of the RNA used in the discovery cohort studies, data from whole blood were used for the GTEX tests.

2.2. Gene expression data

Total RNA was extracted from fresh blood samples from 82 individuals, with 18 individuals excluded from further analysis as their samples’ RNA integrity number was lower than 8.
These individuals are from 18 families who were re-approached to provide RNA for gene expression analysis based on prior genetic observations in these families including DISC1 [1], RELN [35] and TOP3B [36]. Genome-wide gene expression measures were assayed for this discovery cohort using the Illumina HumanHT-12 v4.0 Expression BeadChip. Of the 48,212 probes on the chip, 11,976 were significantly detectable at a threshold of $p \leq 0.01$ in more than 90% of individuals. The expression data for these probes were processed using quantile normalization followed by log transformation. Raw anonymous data regarding this family cohort can be accessed at the Gene Expression Omnibus (GEO) database (GSE48072). For the replication twin cohort ($N = 73$), genome-wide gene expression data have been measured using the Illumina HumanWG6 v3.0 chip, as reported previously in detail [37]. After quality control and data processing, identical to that used on the family data, 18,559 probes from this chip were significantly detectable.

### 2.4. Association between genome-wide expression levels and genotypes

For the discovery cohort used to study gene expression, a mixed effect linear regression model was fitted for each probe and genotype using R (RStudio version 0.99.489) lme4 package [38], after correcting for gender, age, diagnostic status and family or twin status effects as a covariate. This analysis was performed separately but identically for the discovery cohort and the replication twin cohort. We used the false discovery rate (FDR) method in place of a family-wise error rate (FWER). FDR is widely applied for microarray analyses because it allows more genes to be extracted for further exploration, and was performed using the qvalue package in R [39] to estimate the FDRs of $q \leq 0.05$. The post hoc power of our small familial discovery cohort to detect gene expression changes was estimated using R package ssize.fdr [40]. The GTEx database was mined using its own in-built test procedure, entering in a list of gene IDs to be tested against our SNPs of interest. Data from whole blood was used in order to replicate only those genes identified as significantly altered in their gene expression levels at a cut-off of $p \leq 0.05$. When testing for replication, probes significant ($p < 0.05$) in the discovery cohort prior to application of FDR were studied. See figure 1 for a flow chart of analysis.

### 2.5. MicroRNA target prediction and enrichment analysis

A comprehensive list of predicted 3’UTR targets for miR-484 were obtained from the miRWalk database [41], considering only genes predicted by at least six of the 12 programs. Ingenuity pathway analysis (QIAGEN Redwood City, www.qiagen.com/ingenuity) was used to analyse potential enrichment of these genes among those significantly altered in their expression levels by NDE1 SNPs in the discovery and replication cohorts.

### 2.6. Medication data

Prescription medication data from the 931 affected individuals of the schizophrenia family cohort for the period 1 January 1996 to 31 December 2005 was obtained from the Finnish National Prescription Register of the National Social Insurance Institution (SII) [42], thus forming the discovery cohort for medication use. In this cohort ‘medication use’ is based on purchases of prescribed psychoactive drugs for which the SII have paid a reimbursement. Data from this register contain information on date of purchase and the dose, stated as the international standard daily defined dose. Thus, using this data, medication periods were defined according to method 4 proposed by Mantel-Teeuwisse et al. [43], multiplying defined daily dose by a factor of 1.1 and filling 15-day gaps between medication periods. This medication period data was used to determine the probability of cessation of each drug by genotype and converted into a binary variable using three months as a cut-off. This three months cut-off reflects that an individual either purchased more of the same medication, or purchased a different medication within a three-month period after the original purchase date. Only psychoactive medications with at least 15 instances of use for three months or less were taken for association analysis of the individual drugs, meaning that, of the possible 101 psychoactive drugs for which data was available, only 24 were analysed. When data from multiple drugs were combined in order to study classes of medication, all medications for which data are available were used, regardless of individual frequency. In order to account for the fact that
some medication periods may come from the same individual, analysis of the medication usage used logistic regression with GEE-estimation (generalized estimating equations), as used by the geepack package for R [44,45]. Bonferroni correction was used to correct for the multiple tests in the analysis of the individual medications (24 tests) and the groups of medications (five tests). While \( p \)-values are presented unadjusted, only those that would surpass the Bonferroni correction-based thresholds (\( p / C = 0.0021 \) and \( p / C = 0.01 \), respectively) are highlighted. See figure 1 for a flow chart of analysis.

### 2.7. Cell culture and western blotting

To determine whether miR-484 had the potential to affect the expression of selected proteins in a human cell-based system, NLF neuroblastoma cells (Children’s Hospital of Philadelphia) were grown in RPMI 1640/10% fetal calf serum/2 mM L-glutamine (all from Thermo Fischer Scientific) and transfected with 50 nM of either a mimic of mature miR-484 (QIAGEN, Sy-hsa-miR-484) or a negative control microRNA (QIAGEN, AllStars Negative Control microRNA) using Lipofectamine 2000 (Thermo Fischer Scientific) according to the manufacturer’s instructions. After 48 h, cells were lysed using PBS/1% Triton X-100/20 mM MgCl\(_2\) containing protease inhibitor cocktails and DNaseI. Lysates were western blotted and proteins detected using the following antibodies: anti-\( \alpha \)-actin (Sigma, A2066), anti-CYP2C19 (Novus Biologicals, NBP1-19698), anti-CYP2D6 (Abnova, H00001565-B01P), anti-NDE1 (Proteintech, 10233-1-AP), anti-TRIOBP (Sigma, HPA019769) and anti-\( \alpha \)-tubulin (Sigma, T9026). IRDye secondary antibodies were used (LI-COR) and the signal visualized using an Odyssey CLx infrared imaging system (LI-COR). Protein bands of interest were then isolated and quantified using IMAGE STUDIO software (LI-COR). In the case of TRIOBP and CYP2D6, which are each encoded for by multiple splice variants [46–48], the species at 72 and 56 kDa, respectively, were selected for analysis as they are known to be biologically active [49,50]. In the case of NDE1, all of the bands between 40 and 45 kDa, which are consistent with its large number of known phosphorylation events [51], were measured and pooled. All membranes were probed
with secondary antibodies alone first to ensure specificity of signal. Antibody signals were normalized to actin as a loading control. Mean fold-changes between the control and miR-484-treated samples were calculated from seven to eight technical replicates, performed from the same batch of cells. Three independent experiments of this type were then performed for biological replication, and the results compared by two-tailed paired Student’s t-test.

3. Results

3.1. Replication of gene expression changes from previous studies

In our previous analysis of gene expression in association with DISC1 network variants using publicly available data on the CEU (Utah residents with North and Western European ancestry) individuals, 86 genes were found to be differentially expressed across the three DISC1 network variants that could be directly replicated here (DISC1 rs821616, 18 genes; NDE1 rs4781678, 39 genes; NDE1 rs2242549, 29 genes) [30]. To verify these results, made in a cohort of 30 individuals using technical replications, these three variants were tested again, this time using the Finnish family cohort. In total, six of the gene expression changes previously reported could be replicated, for the genes TRIOBP, ZNF500, KIAA1468, FCER1G, SULT1A1 and two probes for the ST3GAL5 gene. These were all in association with the status of the NDE1 gene locus (rs2242549). In the previous analysis, NDE1 rs2242549 associated with changes in gene expression levels of 29 genes, thus these six genes represent a 21% overlap.

3.2. Differentially expressed genes associated with DISC1 pathway genotype

To further investigate the effect of DISC1 network variants on gene expression, we used this Finnish family cohort as a discovery sample to investigate the association of previously positive DISC1 and NDE1 variants with genome-wide gene expression of 11,976 probes. Notably, the NDE1 SNP rs2242549 was significantly associated with gene expression levels of a large number of the probes (table 1 and electronic supplementary material, S1, figure S1). Specifically, 3,824 probes representing 3,314 distinct genes showed uncorrected association with the NDE1 SNP rs2242549 (p ≤ 0.05), of which 2,908 probes, representing 2,542 distinct genes, were associated at FDR ≤5%. We also verified that, despite the size of the discovery cohort, it had sufficient power to detect the differentially expressed genes associated with the rs2242549 variant (electronic supplementary material, figure S2). By contrast, no probes were significantly altered in expression levels in association at FDR ≤5% with either the DISC1 variants tested (SNP rs821616 of the ‘HEP3’ haplotype comprising rs751229 and rs3738401) or with the other NDE1 SNPs tested (rs4781678 or rs1050162).

3.3. Replication of gene expression changes associated with NDE1 genotypes

In order to replicate these observations, we pursued three lines of supporting evidence. Firstly, we noted that although all variants tested are in high D' linkage disequilibrium (LD) and thus

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Table 1. Number of probes and genes significantly altered by variants in the DISC1 network, how they replicate across different cohorts, and how they overlap with the predicted targets of miR-484. n.a., value was not applicable as it was not tested; n.s., value was not significant nor returned by IPA.

<table>
<thead>
<tr>
<th>Variant</th>
<th>Gene variant</th>
<th>Gene expression change</th>
<th>Discovery cohort</th>
<th>Follow-up cohort</th>
<th>Enrichment for predicted targets of miR-484</th>
<th>q ≤ 0.05 probes</th>
<th>q ≤ 0.05 genes</th>
</tr>
</thead>
<tbody>
<tr>
<td>DISC1 haplotype HEP3</td>
<td>rs821616</td>
<td>482</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>NDE1 rs4781678</td>
<td>513</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>NDE1 rs2242549</td>
<td>507</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>NDE1 rs1050162</td>
<td>561</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>enrichment for predicted targets of miR-484</td>
<td>2.2 × 10⁻⁴</td>
<td>5.5 × 10⁻²</td>
<td>0.05</td>
<td>0.05</td>
<td>0.05</td>
<td>0.05</td>
<td>0.05</td>
</tr>
</tbody>
</table>
tag the underlying haplotype structure, only one SNP was in high $R^2$ LD with rs2242549 (rs1050162, $R^2 = 0.88$, $n = 1891$ individuals genotyped at both loci; electronic supplementary material, figure S3), and could, therefore, be assumed to act as an internal replication of observations. Thus, within the discovery cohort, of the genes whose expression levels were associated with rs2242549, 752 probes representing 695 genes were also significantly associated with rs1050162 prior to application of the FDR ($p \leq 0.05$). Secondly, we used existing data from a twin cohort for schizophrenia from Finland [33,37] as a replication cohort, identifying 56 probes, each representing a different gene, with replicable significant alteration in their expression levels associated with rs2242549 (table 1 and electronic supplementary material, S1). Finally, we used the publicly available database GTEx as an additional independent replication cohort, from which we were able to directly test 2651 of the 3314 genes identified (those probes with an official gene name), confirming that 180 genes also display significant gene expression changes related to the NDE1 SNP rs2242549 in this database (table 1 and electronic supplementary material, S1).

In total, 794 probes representing 719 genes had supporting evidence from at least one additional source for their changes in expression related to the NDE1 variant rs2242549, of which 76 probes from 73 genes had supporting evidence from more than one source, and four probes from independent genes (ITGB5, OVGPI, PGRMC1, TST) had supporting evidence from all three sources, that is SNPs in high LD in the discovery cohort, as well as independent replication cohorts of Finnish twins and the GTEx database.

### 3.4. Enrichment of miR-484 target genes among genes whose expression is associated with NDE1 genotype

The finding that expression levels of such a large number of genes could be altered by a single genetic locus was surprising, especially given that the principal functions of the NDE1 protein are not known to be in gene regulation. The NDE1 locus also encodes for a microRNA (miR-484), which is located on a non-coding 5’ exon of the longest splice variant of the NDE1 gene (figure 2). Since the major function of microRNAs is in the regulation of expression of other genes it is the most likely explanation for the sheer number of expression changes observed to associate with these SNPs. We therefore investigated whether the set of genes whose expression is altered by these NDE1 SNPs overlapped with those genes predicted to be targets of miR-484.

Using the miRWalk database [41], 16,027 gene targets are predicted for miR-484, of which 2588 are predicted by at least six or more of the 12 independent prediction programs used in the database (data collated June 2016). Upon examining the list of genes whose expression is altered by SNPs in the NDE1 locus at FDR ≤5% in our discovery cohort, these probes were indeed seen to be enriched for predicted targets of miR-484 ($p = 3 \times 10^{-8}$). Employing the same tests to the three replication studies described above, the enrichment in miR-484 targets was also present for the set of these genes whose expression level is significantly associated with the genotype of both NDE1 SNPs rs2242549 and rs1050162 (which were in high LD), and among those genes which could also be observed in the replication twin cohort. In contrast, they were not enriched among those genes replicated by data from the GTEx database (table 1).

### 3.5. Medication cessation associated with NDE1 genotype

In our previously published analysis we found NDE1 genotypes that significantly associated with expression levels of targets for particular medications relevant to mental health [30]. We therefore tested whether NDE1 genotypes were also associated with cessation of specific medications in a discovery cohort consisting of all affected individuals from the Finnish family cohort. Screening all of the medications frequently used within this discovery cohort, we observed an association between NDE1 rs4781678 genotype and early cessation of use of the antipsychotic levomepromazine (OR = 4.13 per C allele; 95% CI = 1.72–9.91; $p = 0.0090$). When analysed in interaction with gender, association was further observed across NDE1 genotypes with early cessation of the use of diazepam and citalopram (electronic supplementary material, table S2).
two drugs that share a common principal metabolizing enzyme, CYP2C19 (cytochrome P450 2C19, electronic supplementary material, table S3) [54–57]. We therefore asked whether NDE1 genotype was associated with the subset of medications metabolized by the CYP2C19 enzyme. Since four out of seven of the drugs metabolized by CYP2C19 were selective serotonin reuptake inhibitors (SSRIs), we studied these as a separate group as well as further separated based on CYP2C19 metabolism. No significant interaction was observed for the grouping of all drugs metabolized by CYP2C19, however a genotype by gender interaction was noted when all SSRIs were grouped together (rs2075512, OR = 0.37; 95% CI = 0.17–0.79; p = 0.010). When all SSRIs metabolized by CYP2C19 were tested, this genotype by gender interaction became significant (ranging from: OR = 0.27 to 0.31; 95% CI = 0.11 to 0.64 to 0.71; p = 0.0030 to 0.0060) for four out of the five NDE1 markers tested, while no interaction was noted for SSRIs not metabolized by CYP2C19 (table 2 and electronic supplementary material, figure S4). We analysed the remaining drugs metabolized by CYP2C19 together as another grouping (non-SSRIs metabolized by CYP2C19). Interestingly, a significant interaction was observed (ranging from: OR = 3.33 to 5.82; 95% CI = 1.44 to 2.25–7.25 to 15.0; p = 0.0013 to 0.00030) for all five NDE1 markers (table 2 and electronic supplementary material, figure S4). In this case, however, the gender effect is reversed, with SNPs being associated with cessation among females, in contrast to SSRIs metabolized by CYP2C19, for which SNPs were associated with cessation among males.

3.6. The effect of miR-484 on CYP2C19 in cultured cells

Given that a major effect of the NDE1 locus variants examined here seems to be to alter the expression of genes targeted by miR-484, presumably due to altered expression of this miR-484, we hypothesized that the pharmacological consequences of these variants were also likely to occur through miR-484. For this to be the case, miR-484 would need to affect the metabolic activity effects on psychoactive medication.

We therefore conducted a proof-of-principle experiment in NLF human neuroblastoma cells, into which we transfected a mimic of the mature form of human miR-484. Protein levels of CYP2C19 were significantly upregulated following miR-484 mimic of the mature form of human miR-484. Protein levels of NLF human neuroblastoma cells, into which we transfected a control microRNA (figure 3). In contrast, no effect on the transfection, when compared with transfection with a negative control microRNA. NLF human neuroblastoma cells, into which we transfected a control microRNA (figure 3). In contrast, no effect on the transfection, when compared with transfection with a negative control microRNA.

Table 2. Results for the association of NDE1 variants with groups of medications based on their metabolism by the CYP2C19 enzyme and/or selective serotonin reuptake inhibitor (SSRI) class status, showing the p-values and odds ratios (and 95% CIs) for the interaction model. p-values ≤ 0.01 are below the Bonferroni correction threshold for the five groups tested. p-Values and their respective ORs that are below the Bonferroni threshold are in bold. Medications included in each group analysis can be found in the electronic supplementary material, figure S4 and table S3.

<table>
<thead>
<tr>
<th>NDE1 variant</th>
<th>SNP</th>
<th>SNP × gender</th>
<th>OR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs4781678</td>
<td>0.79</td>
<td>0.90</td>
<td>0.97 (0.55–1.69)</td>
</tr>
<tr>
<td>rs2242549</td>
<td>0.56</td>
<td>0.74</td>
<td>1.10 (0.64–1.86)</td>
</tr>
<tr>
<td>rs881803</td>
<td>0.29</td>
<td>0.15</td>
<td>1.53 (0.86–2.73)</td>
</tr>
<tr>
<td>rs2075512</td>
<td>0.64</td>
<td>0.79</td>
<td>0.93 (0.53–1.63)</td>
</tr>
<tr>
<td>haplotype</td>
<td>0.35</td>
<td>0.73</td>
<td>1.10 (0.63–1.93)</td>
</tr>
<tr>
<td>SSRIs (N = 357–402, no. of instances: 496–563)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs4781678</td>
<td>0.31</td>
<td>0.033</td>
<td>0.43 (0.20–0.93)</td>
</tr>
<tr>
<td>rs2242549</td>
<td>0.92</td>
<td>0.031</td>
<td>0.46 (0.23–0.93)</td>
</tr>
<tr>
<td>rs881803</td>
<td>0.41</td>
<td>0.29</td>
<td>0.67 (0.32–1.40)</td>
</tr>
<tr>
<td>rs2075512</td>
<td>0.97</td>
<td>0.010</td>
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</tr>
<tr>
<td>haplotype</td>
<td>0.87</td>
<td>0.029</td>
<td>0.42 (0.19–0.91)</td>
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<td>SSRIs not metabolized by CYP2C19 (N = 282–318, no. of instances: 357–395)</td>
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</tr>
<tr>
<td>rs4781678</td>
<td>0.42</td>
<td>0.003</td>
<td>0.27 (0.11–0.64)</td>
</tr>
<tr>
<td>rs2242549</td>
<td>0.47</td>
<td>0.006</td>
<td>0.30 (0.13–0.70)</td>
</tr>
<tr>
<td>rs881803</td>
<td>0.51</td>
<td>0.064</td>
<td>0.44 (0.19–1.05)</td>
</tr>
<tr>
<td>rs2075512</td>
<td>0.39</td>
<td>0.005</td>
<td>0.31 (0.13–0.70)</td>
</tr>
<tr>
<td>haplotype</td>
<td>0.95</td>
<td>0.006</td>
<td>0.30 (0.13–0.71)</td>
</tr>
<tr>
<td>non-SSRIs not metabolized by CYP2C19 (N = 123–145, no. of instances: 142–169)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs4781678</td>
<td>0.48</td>
<td>0.71</td>
<td>1.36 (0.27–6.87)</td>
</tr>
<tr>
<td>rs2242549</td>
<td>0.30</td>
<td>0.58</td>
<td>1.37 (0.45–4.23)</td>
</tr>
<tr>
<td>rs881803</td>
<td>0.58</td>
<td>0.23</td>
<td>2.24 (0.60–8.38)</td>
</tr>
<tr>
<td>rs2075512</td>
<td>0.10</td>
<td>0.88</td>
<td>0.88 (0.19–4.13)</td>
</tr>
<tr>
<td>haplotype</td>
<td>0.66</td>
<td>0.77</td>
<td>1.25 (0.27–5.76)</td>
</tr>
<tr>
<td>non-SSRIs not metabolized by CYP2C19 (N = 318–355, no. of instances: 411–459)</td>
<td></td>
<td></td>
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<tr>
<td>rs4781678</td>
<td>0.24</td>
<td>0.0051</td>
<td>3.33 (1.44–7.73)</td>
</tr>
<tr>
<td>rs2242549</td>
<td>0.62</td>
<td>0.0013</td>
<td>3.42 (1.61–7.25)</td>
</tr>
<tr>
<td>rs881803</td>
<td>0.63</td>
<td>0.00030</td>
<td>5.82 (2.25–15.0)</td>
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<tr>
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<td>0.0064</td>
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<tr>
<td>haplotype</td>
<td>0.22</td>
<td>0.00030</td>
<td>4.93 (2.07–11.76)</td>
</tr>
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</table>

Finally, we also used the same system to investigate two proteins that are predicted targets of miR-484, each of which illustrates a potential false negative in this study, possibly as a result of power limitations. The first of these, TRIOBP, was observed to be significant in our discovery cohort but not in the replication cohorts, while the other is NDE1 itself, which was highly significant in the larger GTEx database (β = 0.24; s.e. = 0.037; t-statistic = 6.6; p = 2.3 × 10–10), but not originally observed in our discovery cohort. TRIOBP was further selected as it is also an example of the six genes whose expression was associated with NDE1 variation both here and in our previous analysis [30]. In both cases, protein levels were subtly, but statistically significantly, altered following treatment with the miR-484 mimic (figure 3), in comparison to the negative control microRNA.
4. Discussion

Here we have demonstrated that variations within the NDE1 locus, encoding a protein of the DISC1 network of protein interaction partners, can affect both gene expression levels and medication usage of psychoactive drugs used to treat major mental illnesses. Specifically, two SNPs in high LD are associated with replicable expression changes in a large number of genes, and with early cessation of psychoactive medications metabolized by CYP2C19 in a gender-dependent manner. We propose that these observations are linked through the involvement of miR-484. This microRNA is encoded for within the 5′-most intron of NDE1, and the one which is most abundantly expressed, at least in cell culture [58]. Notably, the list of genes whose expression changes are associated with these NDE1 locus variants is significantly enriched for predicted targets of the microRNA, while expression of the CYP2C19 protein has been demonstrated in vitro to be significantly increased following addition of a mimic of mature human miR-484. This microRNA is encoded for within the 5′-untranslated exon of the longest splice variant of NDE1, and the one which is most abundantly expressed, at least in cell culture [58].

![Figure 3](image)

**Figure 3.** The effect of a miR-484 mimic on protein expression in human NLF neuroblastoma cells. (a) Sample western blots showing levels of six proteins in the lysates of cells which had been transfected with either a mimic of miR-484 or with a negative control microRNA. (b) Quantification of three independent experiments, each comprising 7–8 internal replicates. All proteins were normalized to actin.

It can be speculated that the mechanism implicated here through modulation of NDE1 splicing events.

The 16p13.11 locus, in which the NDE1 gene is found, is prone to copy number variations (CNVs), with these 16p13.11 CNVs having been repeatedly associated with psychiatric and neurological disorders [19–22,28]. This locus contains multiple genes, however not all NDE1 splice variants. It is therefore very likely that any event which affects either the expression or alternate splicing of NDE1 would also impact upon miR-484 expression. That many of the SNPs of the NDE1 haplotypes are found in the 3′-most intron of NDE1 (figure 2), which contains many motifs that regulate NDE1 splicing [61], therefore allows speculation that the SNPs could affect miR-484 expression indirectly through modulation of NDE1 splicing events.
changes, could also be a consequence of the duplication of this locus.

This study initially sought to replicate our previous work on the effect of DISC1 network variants on gene expression changes in the general population, using publicly available data on the CEU (Utah residents with North and Western European ancestry) individuals [30]. Of these previously identified 86 genes, we were able to replicate the observed changes in expression of six genes, including expression changes of two probes for ST3GAL5 and a probe for TRIOBP, the only one of these genes to be a predicted target for miR-484. All of these were in association with the NDE1 SNP rs2242549, and represented a 21% overlap with the findings from the prior study [30]. When we tested for gene expression alterations across the genome we identified a large number of probes (n = 2908) representing 2542 genes whose expression levels associated with variants at the NDE1 locus, specifically with the SNP rs2242549. A large proportion of these (752 out of 3824 probes, 695 out of 3314 genes) were significantly altered by another NDE1 SNP (rs1050162), which is in high LD with rs2242549. Yet replication attempts in independent cohorts, although providing validation for some genes (56 in an independent Finnish schizophrenia cohort and 180 using the GTEx database), did not provide unilateral confirmatory evidence, with the exception of four genes (ITGB5, OGVPI, PGRMC1, TST) identified across all three datasets tested. This lack of replication, combined with our new observations and their biological relevance through miR-484 to the NDE1 locus, suggests that, although the variants studied here are common to many populations, their relationship to potential functional mutations at this locus, and their specific biological consequences associated with schizophrenia and gene expression changes, may be unique to this Finnish family cohort [63]. This population difference may account for the lack of replication of most of the previously observed genes in the CEU population [30], and the lack of enrichment for miR-484 targets in the GTEx database. This is consistent with DISC1 variation playing a genetically heterogeneous role in the general incidence of schizophrenia, lacking common illness-associated variations which could be detected by genome-wide association studies [15,16] of global populations, but providing strong evidence for a role in the condition within specific populations and family studies [63].

Another potential explanation for our inability to replicate our observations across cohorts is the fact that our power to detect these effects is reduced due to the small sample sizes used here. Although we have demonstrated that we have 80% power in our discovery cohort to detect large changes in gene expression (Δ = 0.52), this is for our observed 90th percentile of the standard deviation for all genes from our data (σ = 0.53) (electronic supplementary material, figure S2); probes with smaller standard deviations would not be detectable, either for these probes in replication cohorts or for other probes in the discovery cohort. Thus, we verified our findings in a neuroblastoma cell culture model for two genes, which were predicted to be targets of miR-484, but provided inconsistent observations. The first, TRIOBP, was observed in our previously published study of the publicly available data on the CEU (Utah residents with North and Western European ancestry) individuals [30] and was replicated in our discovery family cohort, but not in either the twin or GTEx replication cohorts. In contrast, the second, NDE1, was not observed in any of the Finnish cohorts but was strongly implicated in the larger GTEx data where NDE1 expression levels were strongly associated with the NDE1 SNP rs2242549 genotype (β = 0.24; s.e. = 0.037; t-statistic = 6.6; p = 2.3 × 10⁻9). The proteins encoded for by these genes were each found to be significantly increased by the presence of the mimic of miR-484. Such a verification analysis would be required for all genes implicated in this study. However, with such a large number of genes identified this was not feasible with the cell culture model used here.

When the DISC1 network was studied with respect to treatment, the NDE1 locus again demonstrated association, specifically in interaction with gender for drugs metabolized by CYP2C19. The degree of expression of cytochrome P450 enzymes in lymphocytes was too low to allow us to investigate potential changes in expression level in our family data. Instead we demonstrated in a cell culture model that miR-484 is capable of increasing the expression of CYP2C19, but not that of another major metabolizing enzyme for psychoactive drugs, CYP2D6. The miRWalk database does not robustly predict either enzyme to be a target for miR-484, thus the mechanism of this action needs to be further investigated. The results from the cell culture assays clearly indicate, however, that miR-484 is capable of affecting the expression level of CYP2C19, suggesting that the mechanism through which the locus confers risk and alters medication usage could be the same. In spite of the lack of confident prediction, a direct effect of miR-484 on the CYP2C19 locus remains the simplest explanation for these observations, although indirect effects remain possible.

In the case of medication use, we employed a dichotomous variable based on a cut-off of ceasing to use the prescribed medication after three months or less. This was designed to indicate that a treatment was either not considered to be working or else was having side effects which were too severe and its use was, therefore, stopped. Since the cell culture experiment showed that CYP2C19 protein expression is increased by miR-484, it can be hypothesized that the medications are more rapidly metabolized in individuals carrying these variants, leading to a reduced efficacy of those treatments. Interestingly, the genetic effects on medication differ depending on both class of drug and gender. Males carrying the risk alleles had a higher probability of cessation of use for SSRIIs metabolized by CYP2C19, while females carrying the risk alleles had an increased probability of cessation for non-SSRI drugs that are metabolized by CYP2C19. Although the mechanism for these effects remains unclear at this time, it is noteworthy that the original association between the NDE1 locus and schizophrenia in these families was significant only in females [3]. Taken together this implies that one or more gender-specific effects act as modifying factors in conjunction with the NDE1/miR-484 locus, although this cannot be easily modelled in our cell culture system.

Here, through the identification of altered gene expression patterns that led to the functional implication of miR-484, which is coded on an untranslated exon of the primary isoform of NDE1, we identified a means by which genetic variation in the DISC1 network can not only increase risk of major mental illnesses, but also how those same variants can alter treatment response to specific psychoactive medications through the regulation of their metabolizing enzyme. This study has, therefore, provided new biological insight into psychiatric disorders for which novel medications could be designed, as well as suggesting that
knowledge of an individual’s genotype within the NDE1/miR-484 locus may have potential value in the targeting of current therapies.

Ethics. In all studies, the principles recommended in the Declaration of Helsinki and its amendments were followed. The study has been approved by the Coordinating Ethics committee of the Hospital District of Helsinki and Uusimaa. Informed consent was obtained from all participants.

Data accessibility. The discovery cohort used here is ascertained based on schizophrenia, a disorder that remains highly stigmatizing to the patients and their families. Thus, anonymized gene expression data for schizophrenia, a disorder that remains highly stigmatizing to the patients and their families. Thus, anonymized gene expression data

Authors’ contributions. N.J.B., L.U.V. and W.H. wrote the manuscript text; N.J.B., M.P., L.U.V. and W.H. prepared the manuscript figures; N.J.B., J.S., J.L., J.H. and W.H. designed the study; T.P., J.S., J.L., T.D.C. and J.H. provided access to samples and data; N.J.B. performed laboratory experiments; N.J.B., L.U.V., M.P., A.B.Z., A.O.A., M.T.H. V.S. and W.H. performed the analysis; all authors reviewed the manuscript.

Competing interests. We have no competing interests.

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