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Integrated cardiovascular/respiratory control in type 1 diabetes evidences functional imbalance: Possible role of hypoxia

L. Bianchi^a, C. Porta^a, A. Rinaldi^a, C. Gazzaruso^b, P. Fratino^b, P. DeCata^b, P. Protti^a, R. Paltro^a, L. Bernardi^{a,c,*}

^a Department of Internal Medicine, University of Pavia, Pavia, Italy

^b Department of Internal Medicine and Endocrinology, IRCCS Fondazione S. Maugeri, Pavia, Italy

^c Folkhälsan Institute of Genetics, Folkhälsan Research Center, University of Helsinki, Helsinki, Finland

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ABSTRACT

Background: Cardiovascular (baroreflex) and respiratory (chemoreflex) control mechanisms were studied separately in diabetes, but their reciprocal interaction (well known for diseases like heart failure) had never been comprehensively assessed. We hypothesized that prevalent autonomic neuropathy would depress both reflexes, whereas prevalent autonomic imbalance through sympathetic activation would depress the baroreflex but enhance the chemoreflexes.

Methods: In 46 type-1 diabetic subjects (7.0 ± 0.9 year duration) and 103 age-matched controls we measured the baroreflex (average of 7 methods), and the chemoreflexes, (hypercapnic: ventilation/carbon dioxide slope during hyperoxic progressive hypercapnia; hypoxic: ventilation/oxygen saturation slope during normocapnic progressive hypoxia). Autonomic dysfunction was evaluated by cardiovascular reflex tests.

Results: Resting oxygen saturation and baroreflex sensitivity were reduced in the diabetic group, whereas the hypercapnic chemoreflex was significantly increased in the entire diabetic group. Despite lower oxygen saturation the hypoxic chemoreflex showed a trend toward a depression in the diabetic group.

Conclusion: Cardio-respiratory control imbalance is a common finding in early type 1 diabetes. A reduced sensitivity to hypoxia seems a primary factor leading to reflex sympathetic activation (enhanced hypercapnic chemoreflex and baroreflex depression), hence suggesting a functional origin of cardio-respiratory control imbalance in initial diabetes.

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1. Introduction

In diabetes, the impairment in autonomic function is associated with increased morbidity and mortality [1]. This abnormality could be observed at an early stage by a depressed baroreflex sensitivity [2,3], which is also a well-established marker of cardiovascular mortality for cardiovascular causes [4,5].

Abbreviations: BRS, baroreflex sensitivity; $\text{CO}_{2\text{et}}$, end-tidal carbon dioxide pressure; DLCO, diffusing capacity of the lung for carbon monoxide; FEV₁, forced expiratory volume (at the end of the first second of forced expiration); FVC, forced vital capacity; HIF-1, hypoxia-inducible factor-1; HVR, hypoxic chemoreflex (ventilatory response); HCVR, hypercapnic chemoreflex (ventilatory response); HRV, heart rate variability; N+ subgroup, presence of incipient autonomic dysfunction; N- subgroup, absence of autonomic dysfunction; SDNN, standard deviation of all RR intervals; T1D, type-1 diabetic subjects; VE, minute ventilation; Vt, tidal volume; VT-CO₂, ventilatory threshold for carbon dioxide; CXCR4, C-X-C chemokine receptor type 4; eNOS, endothelial nitric oxide synthase; EPC, endothelial progenitor cells; GLO-1, glyoxalase-1; GSH, glutathione; MGO, methylglyoxal; ROS, reactive oxygen species; SDF-1, CXCR4-ligand stromal cell-derived factor-1; VEGF, vascular endothelial growth factor.

* Corresponding author at: Folkhälsan Research Center, Biomedicum Helsinki (C318b), PO Box 63, FIN-00014, University of Helsinki, Finland.

E-mail address: lbern1ps@unipiv.it (L. Bernardi).

Recent data from our laboratory suggests that the baroreflex abnormality could be to some extent functional, and evidences point to similarities to the more general dysregulation occurring in congestive heart failure [6,7]. In chronic heart failure, a reduced baroreflex and a sympathetic overactivity, are linked to an overactivity of the reflexes that regulate ventilation (chemoreflexes), in a clear inverse relationship [8]. These functional abnormalities are implicated in the reduced exercise performance, and interventions aimed at restoring the cardio-respiratory control (e.g. physical exercise) improved both cardiovascular regulation and exercise tolerance, hence resulting in important clinical improvement in patients with heart failure [9].

On the other hand, we previously found that severe autonomic damage (as it occurs in familial dysautonomia) is associated with a reduction in both chemo- and baroreflexes [10].

In diabetes the cardio-respiratory integrated control remains to a large extent to be investigated. Although several studies have tested the hypothesis of altered respiratory or cardiovascular control, the respiratory and cardiovascular reflexes were tested only in small number of subjects and only separately on chemoreflex (hypercapnic only [11], hypoxic only [12], both chemoreflexes [13]) or baroreflex

[2,3]. Results were interpreted simply as yet another additional evidence of neuropathy and thus of little novelty.

Therefore, a comprehensive evaluation of cardiovascular, hypoxic and hypercapnic reflexes is missing so far, thus preventing us to understand how these reflexes might interact in diabetes.

This is particularly relevant in view of the previous hypothesis of an initially functional (hence reversible) autonomic abnormality in type 1 diabetes. In that case the different reflexes should be linked in a mutual reciprocal interaction, like in heart failure (e.g. ventilatory reflexes enhanced and cardiovascular reflexes depressed). Conversely, a generalized reduction in reflexes could be the result of an established neuropathy, as shown in familial dysautonomia [10].

In the present study we then evaluated the baroreflex sensitivity and the central and peripheral chemoreflex sensitivities in subjects with absent or minimal autonomic dysfunction (as evaluated by a standard battery of cardiovascular reflex tests). The type of the link should give us an indication of whether the extent of the possible abnormality should be interpreted. We reasoned that a generalized depression in both cardiovascular and respiratory reflexes could be an indication of autonomic neural damage (e.g. similar to familial dysautonomia), whereas an inverse relationship between cardiovascular and respiratory reflexes (depressed baroreflex and enhanced chemoreflex gain) would be an indication of functional disorder, eg similar to heart failure.

2. Methods

We studied 46 type-1 diabetic subjects (T1D) (33 ± 1 year, 22 female) and 103 age-matched controls. Clinical characteristics of study participants are shown in Table 1.

Type 1 diabetes was defined as C-peptide deficiency (<0.03 nmol/l) and initiation of permanent insulin treatment within one year after the diagnosis of diabetes. None of the patients showed clinical signs of cardiovascular disease. Only two patients had antihypertensive medication (ACE-inhibitors). The healthy control subjects were recruited by email advertisements among university students and staff. Only individuals with normal fasting glucose and without 1st degree relatives with diabetes mellitus were included. Before participation, all subjects gave their written informed consent. The study protocols were approved by the Ethics Committee of University of Pavia.

2.1. Experimental set-up

All subjects were investigated in a quiet room, at a temperature between 19 and 23 °C, between 8 a.m. and 2 p.m. Before the examination, the subjects received instructions to refrain from alcohol for 36 h, caffeinated beverages and cigarettes for 12 h. A light meal was permitted 2 h before testing. Electrocardiogram was recorded using a bipolar precordial lead. Continuous blood pressure was monitored with validated [14] applanation tonometry (Colin CBM 7000, San Antonio, USA) from the wrist of the left arm held at

heart level. A respiratory signal (by inductive respiratory belts positioned around the chest), pulse oximetry (Ohmeda Pulse Oximeter, Englewood, Colorado, USA) and carbon dioxide were also recorded (COSMOplus, Novametrix Wallingford, Connecticut, USA). All signals were acquired continuously on a personal computer (Apple Macintosh), at 300 samples/s/signal.

2.2. Autonomic testing

The subjects underwent five cardiovascular autonomic function tests using previously described methods [15]: 1) the difference between expiratory and inspiratory heart rate during slow deep breathing (6 cycles per minute), 2) the maximum/minimum 30/15 ratio of RR interval during active standing, 3) the systolic blood pressure response to standing, and 4) the maximum/minimum ratio of RR interval during a Valsalva manoeuvre, 5) the diastolic blood pressure increase during handgrip, obtained by compressing a sphygmomanometer cuff partially inflated and keeping a pressure of 30% of maximal strength for 2 min.

2.3. Baroreflex testing

Electrocardiogram and continuous blood pressure were recorded in the supine position at rest during 5 min of spontaneous breathing, to measure baroreflex sensitivity (BRS).

2.4. Chemoreflex testing

During chemoreflex testing, the participants were seated, and connected to a rebreathing circuit through a mouthpiece, in a fashion similar to that previously described and validated [16,17]. Rebreathing into a closed circuit causes a progressive reduction in inspired oxygen and increase in carbon dioxide concentration, both of which stimulate ventilation. When the response to varying oxygen was to be assessed (hypoxic chemoreflex), end-tidal carbon dioxide pressure ($\text{CO}_{2\text{et}}$) was kept constant at 40 ± 1 mm Hg by passing a portion of the expired air into a scrubbing circuit (containing soda lime) before returning it to the rebreathing bag.

Conversely, when the response to carbon dioxide was to be tested (hypercapnic chemoreflex), oxygen was continuously supplied at a very low flow (1 l/min) to the rebreathing circuit in order to maintain the percentage of arterial oxygen saturation (SaO_2) at baseline values ($>95\%$). The amount of air in the rebreathing circuit was set at 5 l, in order to maintain the duration of each test (and hence the individual's compliance) at about 7 min. Before each rebreathing test, the subjects breathed room air through the same mouthpiece as during the rebreathing, in order to collect baseline data. The rebreathing tests terminated when SaO_2 reached 80% (response to hypoxia) and when $\text{CO}_{2\text{et}}$ reached 55 mm Hg (response to hypercapnia) (Fig. 1). In each condition, we continuously measured the carbon dioxide via the COSMOplus connected to the mouthpiece and SaO_2 . The airway flow was measured continuously by a heated Fleish pneumotachograph (Metabo, Epalinges, Switzerland) connected to a differential pressure transducer (RS part N395-257, Corby, UK) connected in series in the expiratory arm of the rebreathing circuit.

2.5. Assessment of baroreflex sensitivity and heart rate variability

Previous studies have shown a poor correlation between different indices of BRS, while, on the other hand, no method has shown clear superior performance over the other [18]. Accordingly, we have used all the most common methods, and analysed the BRS as their average, as previously validated [19]. BRS was determined from the time series of RR interval and systolic blood pressure using the sequence method for 1) positive and 2) negative sequences, or spectral analysis for the 3) low frequency, 4) high frequency, and 5) for the average of the low- and high-frequency components, 6) the transfer function technique and 7) by the standard deviation method (BRS-SD), following the technical details previously explained [19]. Additionally, the standard deviation of all RR intervals (SDNN) was considered an index of global RR interval variability.

2.6. Analysis of respiration/chemoreflex sensitivity

The ventilatory flow signal was integrated by software and each breath was identified. Breathing rate, tidal volume (V_t) and minute ventilation (V_E) relative to each breath were recognized, with their corresponding values of SaO_2 and $\text{CO}_{2\text{et}}$.

The chemoreflex sensitivity to hypoxia (hypoxic ventilatory response) or hypercapnia (hypercapnic ventilatory response) was obtained from the slope of the linear regression line of minute ventilation plotted against SaO_2 or $\text{CO}_{2\text{et}}$, respectively (Fig. 1). The change in ventilation due to hypercapnia is interpreted as the central chemoreflex sensitivity, whereas the change in ventilation due to hypoxia is interpreted as peripheral chemoreflex sensitivity. The point at which the ventilation started to increase during the progressive HCVr (called ventilatory threshold (VT-CO_2 , Fig. 1) was identified by interpolating the ventilation/ $\text{CO}_{2\text{et}}$ plot using a fourth-order polynomial function.

2.7. Statistical analysis

Data are presented as mean $\pm$ standard error. Differences were analysed by one-way ANOVA factorial design. If overall significant changes were observed ($p < 0.05$), then significance was tested by Sheffe's test. Correlation between different variables was evaluated by linear regression analysis.

Table 1

Clinical characteristics of study participants (mean $\pm$ standard error).

	T1D	Controls	p value
N	46	103	
M/F	24/22	52/51	ns
Age (years)	33.2 ± 1.1	36.4 ± 1.4	ns
Weight (Kg)	63.1 ± 1.5	67.2 ± 1.1	0.025
Height (cm)	167.1 ± 1.4	171.3 ± 0.9	0.008
Body mass index (Kg/m^2)	22.5 ± 0.3	22.9 ± 0.3	ns
Years from diagnosis	7.0 ± 0.9	–	
HbA1c (%) (mmol/mol)	8.19 ± 0.24	–	
	66		
Fasting glucose (mg/dL)	184.4 ± 12.7	–	
Diabetes complications	Diabetic retinopathy:	–	
	Proliferative: 2		
	Non-proliferative: 6		
	Microalbuminuria: 2		
Other pathologies	Hypertension: 2	–	
	Depressive sd.: 1		
	H. Pylori gastritis: 1		
	Hepatitis B: 2		
	Hepatitis C: 1		
	Hiatal hernia: 1		
Medications	ACE-inhibitors: 2 pz	–	
Autonomic score	0.8 ± 0.1	0.0 ± 0.0	–

Fig. 1. Schematic measure of chemoreflex sensitivity using a rebreathing circuit to increase CO_2 at a constant FiO_2 or to decrease FiO_2 at a constant CO_2 .

3. Results

3.1. Autonomic score

The results obtained in the diabetic subjects are presented in Table 1. Mild signs of autonomic involvement were present in 26/46 subjects (average score 1.42 ± 0.14), and absent in 20/46 patients. Five subjects presented 2 abnormal cardiovascular tests, and 3 subjects presented 3 abnormal cardiovascular tests [15].

3.2. Cardiovascular data

The results during 5 min of spontaneous breathing are shown in Table 2. RR interval (inter beat interval) was shorter in T1D compared to controls (T1D: 800 ± 19 vs controls: 865 ± 13 msec, $p < 0.01$). Also SDNN was lower in T1D compared to controls (40.0 ± 3.7 vs 56.8 ± 2.5 msec, $p < 0.005$). No significant difference was seen in blood pressures between groups.

3.3. Respiratory data

The average results at rest during 5 min of spontaneous breathing are shown in Table 2. SaO_2 was lower in T1D compared to controls ($96.7 \pm 0.1\%$ vs $97.4 \pm 0.1\%$, $p < 0.005$). Conversely $\text{CO}_{2\text{et}}$ was higher in T1D (40.3 ± 0.3 mm Hg vs controls: 38.4 ± 0.4 mm Hg, $p < 0.01$). Ventilatory threshold was similar in each group, but the difference between resting $\text{CO}_{2\text{et}}$ and the ventilatory threshold was lower in T1D compared to controls (2.4 ± 0.3 vs 4.3 ± 0.5 , $p < 0.01$). There was a

tendency toward higher ventilation: VE, Vt and breathing rate were all slightly increased in T1D compared to controls.

3.4. Baroreflex sensitivity

Baroreflex sensitivity was reduced in the entire diabetic group (9.8 ± 0.9 vs controls: 13.4 ± 0.8 ms/mm Hg, $p < 0.01$) (Table 3).

3.5. Chemoreflex sensitivity

The hypoxic chemoreflex showed a trend toward a depression (0.73 ± 0.9 vs controls: 0.91 ± 0.07 L/min/% SaO_2 , p:ns) (Table 3).

Conversely, the hypercapnic chemoreflex was significantly increased in the entire diabetic group (1.86 ± 0.19 vs 1.41 ± 0.13 L/min/mm Hg CO_2 in controls, $p < 0.05$) (Table 3).

4. Discussion

4.1. Main findings

Our study shows for the first time to our knowledge that there exist a link between the abnormality in cardiovascular control and the abnormality in respiratory control in type 1 diabetes. We found in our T1D:

1. Low resting SaO_2 and high $\text{CO}_{2\text{et}}$ with a tendency toward higher ventilation compared to controls.
2. Depression in baroreflex and heart rate variability but enhanced “central” (hypercapnic) chemoreflex and not enhanced “peripheral” (hypoxic) chemoreflex.

Table 2Cardiovascular and respiratory data (mean $\pm$ standard error).

Test	Controls (N = 103)	T1D (N = 46)
RR (interbeat interval) msec	865 $\pm$ 13	800 $\pm$ 19**
RR stand deviation (SDNN) msec	56.8 $\pm$ 2.5	40.0 $\pm$ 3.7***
SBP (systolic blood pressure) mm Hg	113.8 $\pm$ 1.8	114.6 $\pm$ 2.2
DBP (diastolic blood pressure) mm Hg	63.1 $\pm$ 1.5	62.8 $\pm$ 2.2*
VE (minute ventilation) (L/min)	10.9 $\pm$ 0.5	12.5 $\pm$ 0.8
Vt (tidal volume) (ml)	985 $\pm$ 57	1014 $\pm$ 89
Breathing rate (/min)	12.6 $\pm$ 0.4	13.9 $\pm$ 0.6
SaO ₂ (%)	97.4 $\pm$ 0.1	96.7 $\pm$ 0.1***
CO ₂ et (end-tidal) (mm Hg)	38.4 $\pm$ 0.4	40.3 $\pm$ 0.3**
CO ₂ et at ventilatory threshold point (mm Hg)	42.0 $\pm$ 0.5	42.7 $\pm$ 0.39
Diff. (resting average CO ₂ et – Vent threshold)	4.3 $\pm$ 0.5	2.4 $\pm$ 0.3**

* p < 0.05 vs controls.

** p < 0.01 vs controls.

*** p < 0.005 vs controls.

These findings suggest that hypoxia due to insufficient hypoxic ventilatory stimulus leads to compensatory enhancement of central chemoreflex through sympathetic activation, which reduces the BRS and the heart rate variability. As a consequence, in our diabetic patients the cardio-respiratory control abnormalities (baro-chemoreflex) seem more likely the result of a functional alteration rather than anatomic neural lesions.

4.2. Low SaO₂ and high CO₂et with a tendency toward higher ventilation

Diabetic patients showed lower resting SaO₂ compared to controls (Table 2) despite a tendency toward higher ventilation (VE, Vt and breathing rate were all slightly increased). This apparently slight difference in SaO₂ was never emphasized before, but it should be noted that due to the dissociation curve of hemoglobin, in the normoxic range even a small difference in SaO₂ implies a large difference in PaO₂. Furthermore, elevated blood HbA_{1c} levels (as it was the case in our T1D patients) lead to an overestimation of SaO₂ by SpO₂ monitored with a pulse oximetry [21]. Thus, we can infer that diabetic patients have indeed a lower PaO₂ compared to controls. The PaO₂ is the input to the chemoreceptors, but despite this should represent a higher ventilatory stimulus, the hypoxic chemoreflex was not increased (or rather showed a trend toward a reduction) in our T1D patients. Our results therefore indicate a condition of relative hypoxemia in T1D.

Although many studies identified several possible mechanisms leading to hypoxia, so far this condition in diabetes has never been formally established. Different possible mechanisms could be involved.

Firstly, a reduced lung function (evaluated as low forced vital capacity, low forced expiratory volume in one second and abnormalities in diffusion capacity (DLCO) due to glycosylation of basal membranes) had been shown in diabetes. The reduced lung function was inversely related to blood glucose levels, duration of diabetes and its severity is independent of smoking or obesity [22,23]. Some possible causes are: micro-angiopathy of the alveolar capillaries and pulmonary arterioles [24,25], chronic low-grade tissue inflammation [26], autonomic neuropathy involving the respiratory muscles [27], loss of elastic recoil secondary to collagen glycosylation of lung parenchyma [28], hypoxia-

induced insulin resistance [29]. Iozzo et al. found a significant decrease in the arterial partial pressure of O₂ and in DLCO, with a significant increase in the alveolo-arterial gradient of O₂ in patients with hypertriglyceridemia, a common condition in type 2 diabetes [30]. Among pathologies causing a “reduced lung function” obstructive sleep apnea syndrome might be particularly relevant. This condition is frequent in type 2 diabetic patients and the two conditions seem to occur in a bidirectional association: obstructive sleep apnoea, through the effects of intermittent hypoxaemia and sleep fragmentation, could stimulate insulin resistance, glucose intolerance, and type 2 diabetes. In reverse, type 2 diabetes might increase the predisposition to, or accelerate the progression of obstructive and central sleep apnoea, possibly through the development of peripheral neuropathy and abnormalities of ventilatory and upper airway neural control [31,32,33]. Additionally there is a decreased oxygen transport and transfer to the tissues, due not only to macro/microangiopathy but also to hemoglobin glycosylation that plays a role in regulating both erythrocyte metabolism and Hb-oxygen affinity. HbA_{1c} shifts the dissociation curve of the hemoglobin to the left and decreases the p50 (partial pressure of oxygen at which 50% of hemoglobin is saturated with oxygen). A lower p50 values indicate increased Hb-oxygen affinity and lower oxygen transfer to peripheral tissues [34,35].

In diabetes there seems to be not only a hypoxic status but also an inadequate response to it, which also seems to start independently from neuropathy. Catrina et al. [36] demonstrated that both hyperglycemia and hypoxia play essential pathophysiological roles in the complications of diabetes, which may result from a defective response of the tissues to low oxygen tension. The hyperglycemia seems to blunt the function of hypoxia-inducible factor-1 (HIF-1), a transcription factor that is essential for adaptive responses of the cells to hypoxia. These findings were confirmed by Bento and Pereira [37] that suggested a molecular mechanism whereby methylglyoxal (MGO), which accumulates in high-glucose conditions, leads to a rapid proteasome-dependent degradation of HIF-1 α (a labile subunit of HIF-1) under hypoxia. This impaired HIF-1 α regulation has several deleterious consequences, including decreased production of vascular endothelial growth factor (VEGF) and its receptor, C-X-C chemokine receptor type 4 (CXCR4) and CXCR4-ligand stromal cell-derived factor-1 (SDF-1) and decreased production and activity of endothelial nitric oxide synthase (eNOS). Other target genes include erythropoietin, glucose transporter 1–3 and lactate dehydrogenase-A. All of these genes and proteins are required for an appropriate response to hypoxia, being the driving forces of neovascularization, myocardial collateralization, wound healing, renal and neural function and endothelial progenitor cells (EPC) mobilization to injured areas [38]. Thus, a decrease in their levels has detrimental consequences for cell and tissue adaptation and survival at low oxygen levels, leading to complications such as diabetic retinopathy, poor wound healing, neuropathies, cardiovascular and renal diseases [38].

Thus, these findings together with those of the present study, suggest that in T1D there is a reduction in arterial and tissue oxygenation, possibly due to a pulmonary dysfunction not compensated by an enhanced hypoxic peripheral chemoreflex, possibly as a consequence of multiple cellular abnormalities (hyperglycemia, low-grade inflammation, oxidative stress), blunting the normal response to hypoxia.

4.3. Enhanced “central” (hypercapnic) chemoreflex, but reduced baroreflex and heart rate variability

Our results suggest for the first time an integrated view in the pathophysiologic mechanisms present in diabetes that links the hypoxia, as a “primum movens”, with baro- and chemoreflexes dysfunction. In fact, the consequence of the reduced sensitivity to hypoxia would be a compensatory sympathetic activation leading to a stimulation of the hypercapnic central chemoreflex, but also to a depression of baroreflex, heart rate variability and autonomic function tests [39]. It is of note that, despite a high resting and-tidal CO₂ levels (likely resulting from the

Table 3Baro- and chemo- reflexes (mean $\pm$ standard error).

Test	Controls (N = 103)	T1D (N = 46)
Baroreflex sensitivity (ms/mm Hg)	13.4 $\pm$ 0.8	9.8 $\pm$ 0.9**
Hypercapnic chemoreflex (central) (L/min/mm Hg)	1.41 $\pm$ 0.13	1.86 $\pm$ 0.19*
Hypoxic chemoreflex (peripheral) (L/min/%SaO ₂)	0.91 $\pm$ 0.07	0.73 $\pm$ 0.08

* p < 0.05.

** p < 0.01.

mechanisms discussed above) the diabetic patients showed values very close to the ventilatory threshold for CO₂, a finding that matched the increased central chemoreflex sensitivity. The fact that the threshold was similar as in the control group further confirmed that the sensitivity to CO₂ was indeed not compromised. Thus, it seems that these alterations, rather than the *consequence* of neuropathy, likely suggest a more complex derangement of cardiorespiratory reflexes, which may eventually lead to neuropathy with time.

Our previous study indeed demonstrated that oxygen administration improves heart rate variability and BRS more in T1D than in controls [40]. The increased response to hyperoxia suggested a pre-existing condition of tissue hypoxia that functionally restrained parasympathetic activity in patients with type 1 diabetes [40]. Our present findings are also supported by most studies performed in diabetes, showing either abnormal ventilatory response to hypoxia with a preserved response to hypercapnia in type 2 diabetes [13] or a depressed baroreflex sensitivity [2,3], although in these previous studies in diabetes the possible link between these two essential reflexes was not considered and as a consequence the combination of cardiorespiratory control tests was not performed. A reciprocal interaction of baroreflex and peripheral chemoreflex, was seen in healthy volunteers [41] or in patients with cardiovascular disease [8,42]. Many relevant cardiovascular/respiratory diseases (e.g. heart failure, hypertension, chronic obstructive pulmonary diseases) exhibit potential vicious circles of reflex derangements [8]: the reduced cardiovascular control (baroreflex attenuation) interplays with increased breathing stimulation (chemoreflex augmentation) through a sustained sympathetic activation and a reduced vagal activity. The same reciprocal baro- and chemo-reflex interaction seen in T1D in the present study allowed us to suggest that the abnormalities observed could be the result of a functional imbalance rather than neuropathy, similar to what is already established in heart failure [8,43]. This is clinically important because a functional imbalance is in principle potentially reversible.

4.4. Clinical implications

The respiratory control alterations observed in the present study might result from reduced response to hypoxia, and lead to compensatory activation of hypercapnic chemoreflex (which in turn limits in part the extent of hypoxia) and sympathetic activity (which reduces heart rate variability, BRS, and the autonomic cardiovascular tests).

Respiratory control alterations could be important to worsen both the autonomic dysfunction and its consequences, e.g. by worsening exercise tolerance in diabetes, similarly to chronic congestive heart failure [44]. In turn, resting hypoxemia and sympathetic activation are important factors leading to low-grade systemic inflammation, reactive oxygen species accumulation (ROS) and oxidative stress [31,45,46], which are well known to be involved in the pathophysiology of diabetes and most of its systemic complications [47,48]. The functional nature of this alteration, at least in early stages, suggest possible correcting factors aimed at adjusting respiratory control abnormalities, like physical activity, breathing control exercises, positive airway pressure treatment. Other potential interventions include drugs like Desferrioxamine and dimethylxylglycine, which both stabilize and activate HIF-1 α , or gene transfer-based stabilization of HIF-1 α (adenovirus encoding a constitutively active form of HIF-1 α) [49]. Our results warrant further researches that could identify new therapies for preventing diabetic complications.

Conclusion

We have shown that the autonomic dysfunction of diabetes is part of a complex reflex derangement, relating the cardiovascular (low BRS) to the respiratory control (inadequate, not enhanced hypoxic chemoreflex and increase in hypercapnic chemoreflex).

T1D showed to be not only “hypoxic” at rest, but also with a reduced sensitivity to hypoxia which was partially compensated by a stimulation of central chemoreflex, through sympathetic activation that reduced BRS and autonomic function tests as well (normally interpreted as

“neuropathy”). We show in the present study that what is normally called “autonomic neuropathy” could be in many cases a functional condition of sympathetic activation, driven by many factors, one of which seems to be resting hypoxia. Our data provide support to the hypothesis that techniques or drugs capable of increasing the sensitivity to or relieve hypoxia could be potentially helpful in improving the autonomic dysfunction and ultimately prevent diabetic complications.

Conflict of interest

No potential conflicts of interest relevant to this article were reported.

Author contributions

L.B. (1st author), C.P., A.R., C.G., P.F., P.DC., P.P., R.P, drafted the manuscript and researched data. L.B. (senior author) developed the study concept and design, contributed to interpretation and analysis, edited and revised the manuscript; he had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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