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Graphical Abstract

Synthesis of novel 1,2-bis-quinolinyl-1,4-naphthoquinones: Targeting by ERK2, cytotoxicity and molecular docking

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X-ray structural analysis of **4c** (displacement parameters are drawn at 50% probability level)

Cartoon backbone representation of crystal structure binding mode (in cyan) and predicted docking pose (in pink) of 2SH inhibitor with the ERK2 receptor.

Synthesis of novel 1,2-bis-quinolinyl-1,4-naphthoquinones: Targeting by ERK2, cytotoxicity and molecular docking

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Highlights

- Novel series of bis-quinolinylquinones **3a-d** and **4a-c** were obtained and identified by different spectroscopic techniques.
- Targeting ERK2, compounds **3a**, **3b** and **4c** inhibited ETS-1 phosphorylation by ERK2 in a dose dependent manner.
- Molecular docking studies were carried out to explore the binding pattern of some of the newly synthesized compounds towards ATP-binding site of ERK2 receptor.
- Compound **4c** showed selectivity for ERK2 inhibition with ATP-competitive inhibition mechanism and further structure improvement for enhancing its pharmacokinetic profile is recommended.

- *In vitro* anticancer activity against the NCI-60 panel of tumor cell lines revealed that the tested compounds demonstrated from mild to strong cytotoxic activity.
- The cytotoxic effect of compound **3d** was apparent across all tumor cell lines tested, except leukemia, with 47 out of the 52 non-leukemic cell lines showing negative growth percentages.
- NCI *in vitro* five dose testing revealed that compounds **3b** and **3d** represent promising lead anticancer compounds with GI₅₀ values ranging between (0.31-3.04 μ M), and (0.26 – 0.93 μ M), respectively.

Abstract

In an effort to establish new candidates targeting ERK2 with improved antineoplastic activity, two novel series of *N*-2,3-bis(6-substituted-4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)naphthalene-1,4-diones **3a-d** and substituted *N*-(methyl/ethyl)bisquinolinone triethylammonium salts **4b,c** were successfully synthesized. The synthesis involved the reactions of 2 equivalents of 4-hydroxy-2(1*H*)-quinolinones **1a-f** and one equivalent of 1,4-naphthoquinone (**2**) in a mixture of ethanol/dimethylformamide (1:1) as a solvent and 0.5 ml Et₃N. In case of the reaction of 6-methyl-4-hydroxyquinolone **1b** with **2**, a side product **4a** of the second series was obtained. In general, the presence of free NH-quinolone gave a single compound of the first series, whereas reaction of *N*-methyl/ethyl-quinolones **1e,f** with **2** enhanced the formation of compounds of the second series. The structures of the new compounds were proved by different spectroscopic techniques such as IR, NMR (2D-NMR) and mass spectra, elemental analysis, and X-ray crystallography. In order to further elucidate the mechanism of action of these newly synthesized compounds, compounds **3a**, **3b**, **4b** and

4c were investigated for their MAP Kinase pathway inhibition together with molecular docking using ATP-binding site of ERK2 receptor. The results revealed that compounds **3a**, **3b** and **4c** inhibited ETS-1 phosphorylation by ERK2 in a dose dependent manner. Also, compound **4c** showed highest potency and selectivity for ERK2 inhibition with ATP-competitive inhibition mechanism which was confirmed by the formation of three hydrogen bond in the molecular docking studies. Interestingly, the synthesized compounds were then tested for their *in vitro* anticancer activity against the NCI-60 panel of tumor cell lines. The results revealed that the selected compounds displayed from modest to strong cytotoxic activities. Compound **3b** demonstrated broad spectrum anti-tumor activity against the nine tumor sub-panels tested, while compound **3d** proved to be lethal to the majority of the cancer lines which was approved by their promising GI₅₀ and TGI values in NCI *in vitro* five dose testing. These results revealed that the synthesized compounds can potentially serve as leads for the development of novel chemotherapeutic agents and structure improvement will be necessary for some derivatives for enhancing its cellular activities and pharmacokinetic profile.

Keywords: *1,2-Bis-quinolinyl-1,4-naphthoquinones; 1,4-Naphthoquinone; Cytotoxicity; Targeting by ERK2; Molecular docking.*

1. Introduction

Mitogen activated protein kinase (MAPK) is probably considered the major signaling transduction pathway in cell biology which plays a pivotal role in the regulation of multiple cell processes [1]. MAPK pathway is implicated in the regulation of gene expression, apoptosis, metabolism, cell proliferation and differentiation [2]. Indeed, MAPK appears to induce tumor growth, angiogenesis and metastasis by dysregulation of three major pathways which are ERK, JNK and p38 [3]. ERK2 inhibition attracted a great interest within the ERK kinase subgroup since

this protein can have cell pro-proliferative effects. In this light, a number of studies have been performed for the development of ERK inhibitors for human cancer treatment [4]. The quinone scaffold is present in many clinically used cancer chemotherapeutic drugs [1,2]. Furthermore, 1,4-naphthoquinones are used widely in the pharmaceutical and agrochemical industries. They possess a variety of pharmacological properties such as anticancer [3-6], antimicrobial [7,8], antiviral [9-11], antifungal [12-15], and antiplatelet activities [16-18]. Folkers et al. [19] reported that 1,4-benzoquinone derivatives - the analogs of coenzyme Q10 - are potent compounds in an antitumor test system in vivo, in addition to their effective antitumor activity in other tumor systems. 2-Phenyl-3-hydroxyquinolin-4(1H)-ones exhibit pronounced biological activities and have been identified as strong anticancer agents with significant in vitro cytotoxicity against various cancer cell lines [20-23]. Bis-quinolones of type I (Figure 1) occur naturally, and have fluorescent properties and biological activity [24,25]. In a recent communication [26], the synthesis of the zwitter-ionic 4-hydroxycoumarin derivatives **II** (Figure 1) was reported, through a unique reaction of 4-hydroxycoumarins with *p*-benzoquinone and pyridine in aqueous acetone. A recent report shows the utility of 4-hydroxy-1-phenylquinolin-2(1H)-one in a one-pot synthesis of pyrimidoquinolines [27]. We reacted 4-hydroxy-1-phenylquinolin-2(1H)-one with diethyl acetylenedicarboxylate, to give ethyl 5,6-dihydro-2,5-dioxo-6,9-disubstituted-2H-pyrano[3,2-c]quinoline-4-carboxylates in good yields [28]. We also synthesized 2'-amino-2,5'-dioxo-5',6'-dihydro-spiro(indoline-3,4'-pyrano[3,2-c]-quinoline)-3'-carbonitriles in good to excellent yields from the reactions of 4-hydroxy-1-phenylquinolin-2(1H)-one with 2-(2-oxo-1,2-dihydroindol-3-ylidene)-malononitrile [29]. In view of the aforesaid and the expected anticancer activity of the above pharmacophores, we now report the design of the novel title compounds as a combination of the most active quinolinone and naphthoquinone entities. ERK2 inhibition mechanism towards some of the synthesized compounds was studied. Meanwhile, molecular docking study for the synthesized compounds with ATP-binding site of ERK2 receptor was carried out to reveal the inhibitor-receptor binding mode. Also, the cytotoxicity of the synthesized compounds was

evaluated against NCI 60 cell lines of different nine tumor subpanels. Moreover, compounds **3b** and **3d** were further selected towards NCI *in vitro* five dose testing.

Figure 1. Structures of bis-quinolones **I** and Zwitterionic salt of quinolone **II**

2. Experimental Section

2.1. Biological investigation

Targeting ERK2 inhibition:

In order to study the mechanism of action of the newly synthesized compounds, compounds **3a**, **3b**, **4b** and **4c** were investigated for MAP Kinase pathway inhibitory activity. The results revealed that treatment of *BRAF*- mutant A375 melanoma cell lines with the tested compounds **3a**, **3b** and **4c** inhibited Ets-1 phosphorylation by ERK2 in a dose dependent manner (**Table 1** and **Fig. 4**). In addition, the results recorded in **Table 1** indicated that compound **4c** potently inhibited ERK2 activity with IC₅₀ value of 0.71 μM. While, compounds **3a** and **3b** inhibited ERK2 activity with IC₅₀ values of 10.68 μM and 8.65 μM, respectively. Moreover, compound **4c** inhibited phosphorylation by JNK and P38α at IC₅₀ values of 10.5 μM and 10.2 μM, respectively. This in turn, indicated the selectivity of compound **4c** towards ERK2 inhibitory activity which may be attributed to the formation of three hydrogen bonds with Asp111, Ser153 and Cys166 in the ATP-binding site of ERK2 receptor (as illustrated in molecular docking calculations). Also, **Fig. 4 (E)** demonstrated that the ability of compound **4c** to inhibit Ets-1 phosphorylation by ERK2 is decreased by increasing concentrations of ATP which suggesting ATP-competitive inhibition mechanism.

Table 1. The ability of the tested compounds to inhibit MAP kinases activity

	3a	3b	4b	4c
	IC ₅₀ ^a (μM)	IC ₅₀ ^a (μM)	IC ₅₀ ^a (μM)	IC ₅₀ ^a (μM)
ERK2	10.68±1.2	8.65±1.4	>100	0.71±0.1
JNK2	>100	94±10.2	>100	10.5±1.4
p38α	>100	89±9.1	>100	10.2±1.3

^aIC₅₀ Determined as described in Methods section, Data were fitted to equation 1.

Figure 1. The effect of compounds **3a**, **3b** and **4c** on the MAP kinases activity *in vitro*. In an *in vitro* kinase assays, **3a** (A) **3b** (B) and **4c** (C) inhibited Ets-1 phosphorylation by ERK2 in a dose dependent manner. (D) Compound **4c** showed less potency in inhibiting GST-c-Jun (1-221) phosphorylation by JNK2 and GST-ATF2 (1-115) phosphorylation by P38 α if compared to its ability to inhibit ERK2 kinase activity. Data were fitted to Eq. 1 (E) Higher concentrations of ATP decreased the ability of compound **4c** to inhibit Ets-1 phosphorylation by ERK2 suggesting ATP-competitive inhibition mechanism.

2.3. Molecular docking calculations

To reveal the binding features of the synthesized compounds with ERK2 receptor, molecular docking calculations were performed. Docking calculations were carried out on the top potent compounds towards the ATP-active site of ERK2 receptor (PDB code: 4O6E) using Autodock4.2 software. Self-docking of the original ligand (i.e. 2SH) with ERK2 receptor was first carried out to assess the performance of Autodock in predicting the ligand-receptor binding mode. Self-docking calculation indicated excellent performance of Autodock software with RMSD value of 0.25 Å (Figures 5 and 6).

Figure 5. 2D Ligplus representation of (i) **3a**, (ii) **3b**, (iii) **3d**, and (iv) **4c** with important amino acid residues of ATP-binding site of ERK2 receptor.

Figure 6. Cartoon backbone representation of crystal structure binding mode (in cyan) and predicted docking pose (in pink) of 2SH inhibitor with the ERK2 receptor.

Molecular docking calculations were then performed for the synthesized compounds **3a**, **3b**, **3d** and **4c** towards ATP-active site of ERK2 receptor. The binding features of these compounds were then investigated. 2D representations of the binding modes of the examined compounds inside the ATP-binding site of ERK2 receptor are depicted in Figure 5. According to the docking results, compounds **3a**, **3b** and **3d** showed similar binding modes and affinities with ERK2 receptor. The calculated binding energies were -8.65, -8.62 and -8.84 kcal/mol for compounds **3a**, **3b** and **3d**, respectively, with ERK2 receptor. The analysis of binding mode showed that compound **3a** forms two essential hydrogen bonds with lengths of 2.68 and 2.67 Å with carboxylate group of Asp167 amino acid. Similarly, compounds **3b** and **3d** form an essential hydrogen bond with Asp167 with bond length of 2.67 and 2.65 Å, respectively. While, substitution of R² with a methyl group and bromine atom in compounds **3b** and **3d**, respectively, resulted in formation of a hydrogen bond between the *N*-quinolin atom and carboxylate group of Asn154 with bond length of 3.11 and 3.00 Å, respectively. Compared to compound **3** derivatives, compound **4c** showed better affinity towards ERK2 with binding energy of -9.63 kcal/mol. The better affinity of **4c** may be attributed to the formation of three hydrogen bonds with Asp111, Ser153 and Cys166 with bond lengths of 2.62, 2.95 and 3.20 Å, compared to two hydrogen bonds for compound **3** derivatives. The molecular docking results could support the postulation that our synthesized compounds may act as potent ERK2 inhibitors.

2.1.1. *In vitro* anticancer screening at NCI

A series of new fused dihydroxyisoquinolin-naphthalene-1,4-diones were synthesized and submitted to Development Therapeutic Program, National Cancer Institute (NCI), Chemotherapeutic Research division, USA, for evaluation of their *in vitro* antitumor activities.

Compounds **3a-d** and **4b,c** were selected and evaluated against 60 cell lines which included nine tumor subpanels (leukemia, melanoma, lung, colon, CNS, ovarian, renal, prostate and breast cancers). Potential anticancer effect of the synthesized compounds at single concentration of 10^{-5} M has been reported as the graph of mean growth percentage (G%) of all the treated sixty cell lines compared to untreated control cells which give both inhibition values (between 0 and 100) and cytotoxicity values (less than 0).

Table 1. Growth percent (G %) of the NCI-60 panel tumor cell lines at 10^{-5} M concentration of compounds **3a-d**, and **4b,c**

Subpanel tumor cell lines	% Growth Percent (G %)					
	3a	3b	3c	3d	4b	4c
Leukemia						
CCRF-CEM	157.34	16.64	123.82	25.33	154.72	115.36
HL-60(TB)	126.94	8.93	115.05	45.57	173.59	139.37
K-562	107.83	10.70	101.76	14.32	127.16	112.87
MOLT-4	100.16	-13.27	119.44	30.95	123.74	109.37
RPMI-8226	111.83	32.81	107.40	6.40	112.10	106.00
SR	102.03	23.12	125.95	20.78	123.68	99.16
Non-Small Cell Lung Cancer						
A549/ATCC	106.51	21.84	106.57	-32.20	102.98	102.69
EKVX	ND	ND	ND	ND	ND	ND
HOP-62	90.58	8.69	97.16	-52.85	92.57	98.12
HOP-92	104.40	38.67	101.81	-13.17	115.72	107.30
NCI-H226	94.53	-1.84	111.49	-30.41	96.43	103.94
NCI-H23	80.95	15.52	92.32	-43.00	90.46	97.55
NCI-H322M	86.05	38.80	95.01	-3.29	89.13	86.60
NCI-H460	93.39	-39.82	110.78	-80.88	109.24	109.68]
NCI-H522	104.91	7.58	88.89	-31.36	96.60	101.92
Colon Cancer						
COLO 205	98.03	69.95	111.13	73.06	104.66	105.28
HCC-2998	95.32	62.03	92.50	-44.34	95.91	103.12
HCT-116	94.46	12.93	104.95	-21.67	112.33	102.96
HCT-15	94.52	13.71	104.45	-36.33	94.96	92.28
HT29	110.94	21.67	108.77	25.75	98.61	107.52
KM 12	93.44	18.46	100.50	-53.20	96.45	96.96
SW-620	93.64	7.11	105.55	-70.66	107.45	99.87
CNS Cancer						
SF-268	100.04	9.40	102.51	-41.39	102.31	92.12
SF-295	102.40	49.86	96.29	17.19	102.81	100.28
SF-539	100.24	61.11	104.25	-57.73	102.09	99.73
SNB-19	96.45	26.35	104.11	4.13	98.32	103.48
SNB-75	94.29	14.38	91.68	-35.78	111.60	100.25
U251	101.93	-16.17	109.35	-57.86	106.73	98.32
Melanoma						
LOX IMVI	94.03	12.07	95.52	-67.49	94.81	93.52
MALME-3M	85.19	-39.07	106.59	-74.34	100.38	87.85
M14	98.32	35.68	108.82	-31.93	102.68	96.61
MDA-MB-435	99.81	30.71	106.13	-63.36	108.62	102.39
SK-MEL-2	107.78	11.80	101.33	-14.67	98.85	113.69
SK-MEL-28	108.14	34.91	108.43	-47.84	103.70	104.43
SK-MEL-5	96.30	25.84	108.92	-9.06	95.99	100.92
UACC-257	110.22	63.05	117.16	-51.58	105.10	104.98
UACC-62	95.06	29.49	92.34	-48.85	90.84	99.15
Ovarian Cancer						
IGROV1	66.52	6.46	100.30	-55.93	87.58	92.79

OVCAR-3	95.64	38.73	103.69	-81.47	100.85	105.06
OVCAR-4	109.09	25.25	96.41	-59.70	118.91	108.12
OVCAR-5	98.36	66.42	109.93	10.35	95.98	98.17
OVCAR-8	98.74	14.44	108.14	-23.54	101.28	104.82
NCI/ADR-RES	94.30	14.91	98.53	-28.90	97.97	96.44
SK-OV-3	94.09	43.99	102.85	-32.58	102.10	109.02
Renal Cancer						
786-0	105.89	12.73	106.72	-63.21	115.76	101.79
ACHN	92.74	6.58	106.51	-54.02	96.81	97.34
CAKI-1	94.26	-2.66	94.15	-71.98	96.09	100.02
RXF 393	101.14	-10.80	124.22	-76.09	109.09	110.90
SN12C	89.83	-4.86	102.59	-75.38	89.90	99.10
TK-10	129.41	46.95	97.55	-38.99	109.69	97.50
UO-31	82.98	-11.02	85.09	-65.01	88.43	83.69
Prostate Cancer						
PC-3	99.39	2.45	108.80	-60.51	104.23	105.00
DU-145	102.77	-58.73	108.34	-38.24	102.72	101.53
Breast Cancer						
MCF7	86.18	12.27	97.03	-35.19	101.74	93.26
MDA-MB-231/ATCC	98.38	-6.31	99.23	-8.73	92.92	90.37
HS 578T	93.67	32.36	113.15	-21.93	110.10	107.27
BT-549	102.88	32.74	110.39	-25.44	109.55	105.52
T-47D	97.46	11.86	77.24	-34.25	95.71	95.97
MDA-MB-468	94.12	-23.13	126.84	-11.52	98.25	99.39

ND: Not determined

In the present investigation, compounds **3a**, **3c**, **4b** and **4c** (Table 1) displayed modest potency against the tested tumor cell lines and were considered to be the least effective members. By contrast, compound **3d** proved to be lethal to the majority of the cancer lines. The cytotoxic effect of compound **3d** was apparent across all tumor cell lines tested, except leukemia, with 47 out of the 52 non-leukemic cell lines showing negative growth percentages. Also, strong lethality was induced by this compound against leukemia cancer cell lines RPMI-8226 and K-562 where the G% were 6.40% and 14.32%, respectively. Similarly, a complete cell death is achieved by compound **3b** against leukemia cancer cell line MOLT-4, non-small cell lung cancer cell lines NCI-H226, NCI-H460, CNS cancer cell line U251, melanoma cancer cell line MALME-3M, ovarian cancer cell line OVCAR-3, renal cancer cell lines CAKI-1, RXF-393, SN12C, UO-31, prostate cancer cell line DU-145, breast cancer cell lines MDA-MB-231/ATCC and MDA-MB-468. However, this compound elicited considerably greater growth inhibition activity against many cancer cell lines such as leukemia cancer cell line HL-

60(TB), K-562, non-small cell lung cancer cell line HOP-62, NCI-H522, colon cancer cell line SW-620, CNS cancer cell line SF-268, melanoma cancer cell line SK-MEL-2, ovarian cancer cell line IGROV1, renal cancer cell line ACHN, prostate cancer cell line PC-3 and breast cancer cell line T-47D, where the G% were 8.93%, 10.70%, 8.69%, 7.58%, 7.11%, 9.40%, 11.80%, 6.46%, 6.58%, 2.45% and 11.86%, respectively.

From the above mentioned results, it is obvious that compound **4c** is considered to be the least effective against the tested tumor cell lines. However, it achieved potent selectivity and inhibition of Ets-1 phosphorylation by ERK2. The matter that may be attributed to, when *BRAF*- mutant A375 melanoma cell lines been treated with compound **4c**, the most potent derivative, it did not exhibit any effect on the MAP kinase pathway, which can be the reason behind its null effect on the NCI cytotoxicity studies. Given its potency towards ERK activity *in-vitro*. So, further structure improvement is recommended to enhance its ability to get into the cells for improving its cellular activities and pharmacokinetic profile.

2.1.2. In vitro five-dose NCI 60 cell panel assay

Interestingly, compounds **3b** and **3d** exhibited the most pronounced inhibitory activity in single dose preliminary tests, so they were progressed to further advanced five dose testing. The cytotoxic and/or growth inhibitory effects of the compounds were quantified *in vitro* against the full NCI-60 panel of human tumor cell lines using a series of five 10-fold dilutions, ranging from 10^{-4} M to 10^{-8} M. Three dose response parameters were calculated for each cell line, GI₅₀ (molar concentration required for 50% growth inhibition), TGI (molar concentration leading to total growth inhibition), and LC₅₀ (molar concentration resulting in 50% cell death) (Tables 2 and 3).

Figure 2. Growth inhibitory activity of compound **3d** at different dose levels against the NCI-60 cell lines, grouped into the nine different cancer types included in the panel.

Figure 3. Growth inhibitory activity of compound **3b** at different dose levels against the NCI-60 cell lines, grouped into the nine different cancer types included in the panel

Table 2. NCI cytotoxic parameters (GI₅₀, TGI and LC₅₀) of compound **3d** in μM .

Subpanel tumor cell lines	GI ₅₀	TGI	LC ₅₀
Leukemia			
CCRF-CEM	0.93	>16.7	16.7
HL-60 (TB)	ND	ND	ND
K-562	0.59	>16.7	16.7
MOLT-4	0.82	16.7	16.7
RPMI-8226	0.71	16.7	16.7
SR	0.93	16.7	16.7
Non-Small Cell Lung Cancer			
A549/ATCC	0.58	16.7	16.7
EKVX	0.35	1.53	16.7
HOP-62	0.29	0.66	1.50
HOP-92	0.38	ND	16.7
NCI-H226	0.34	0.94	16.7
NCI-H23	0.30	0.79	16.7
NCI-H322M	0.53	16.7	16.7
NCI-H460	0.42	1.02	16.7
NCI-H522	0.36	1.11	16.7
Colon Cancer			
COLO 205	16.7	16.7	16.7
HCC-2998	0.35	0.83	4.14
HCT-116	0.38	1.00	16.7
HCT-15	0.31	0.67	1.47
HT29	3.87	16.7	16.7
KM 12	0.57	1.97	16.7
SW-620	0.35	0.76	ND
CNS Cancer			
SF-268	0.47	ND	16.7
SF-295	0.27	0.57	1.18
SF-539	0.31	0.64	ND
SNB-19	0.61	16.7	16.7
SNB-75	0.26	0.54	1.11
U251	0.37	0.80	16.7
Melanoma			
LOX IMVI	0.28	0.55	1.09
MALME-3M	0.37	0.77	1.62
M14	0.52	1.60	16.7
MDA-MB-435	0.57	2.11	7.07
SK-MEL-2	0.47	1.46	16.7
SK-MEL-28	0.38	0.93	16.7
SK-MEL-5	0.48	1.47	12.8
UACC-257	0.48	1.11	16.7
UACC-62	0.55	16.7	16.7
Ovarian Cancer			
IGROV1	0.49	1.41	16.7
OVCAR-3	0.39	0.93	8.68
OVCAR-4	ND	ND	ND
OVCAR-5	4.19	16.7	16.7
OVCAR-8	0.53	16.7	16.7
NCI/ADR-RES	0.36	1.08	16.7
SK-OV-3	0.39	1.22	16.7
Renal Cancer			
786-0	0.36	0.81	16.7
A498	ND	16.7	16.7
ACHN	0.33	0.91	16.7
CAKI-1	0.38	1.03	16.7
RXF 393	0.30	0.65	1.41
SN12C	0.36	0.79	16.7
TK-10	0.48	1.18	16.7
UO-31	0.29	0.64	1.41
Prostate Cancer			
PC-3	0.30	0.69	ND
DU-145	0.43	1.11	16.7
Breast Cancer			
MCF7	0.33	1.21	16.7
MDA-MB-231/ATCC	0.54	16.7	16.7

HS 578T	0.44	ND	16.7
BT-549	0.42	1.35	16.7
T-47D	0.31	ND	16.7
MDA-MB-468	0.41	1.49	16.7

ND: Not determined

Table 3. NCI cytotoxic parameters (GI₅₀, TGI and LC₅₀) of compound **3b** in μ M.

Subpanel tumor cell lines	GI ₅₀	TGI	LC ₅₀
Leukemia			
CCRF-CEM	0.83	8.03	>100
HL-60 (TB)	ND	ND	ND
K-562	0.38	3.44	>100
MOLT-4	0.53	3.02	>100
RPMI-8226	2.32	6.68	>100
SR	1.49	ND	>100
Non-Small Cell Lung Cancer			
A549/ATCC	2.10	4.26	8.64
EKVX	1.72	5.66	>100
HOP-62	1.81	3.97	ND
HOP-92	1.66	5.10	>100
NCI-H226	1.79	3.73	ND
NCI-H23	1.63	4.04	10.4
NCI-H322M	2.24	5.85	>100
NCI-H460	1.78	3.39	6.44
NCI-H522	1.88	4.25	ND
Colon Cancer			
COLO 205	2.85	12.8	>100
HCC-2998	2.30	4.97	>100
HCT-116	0.42	1.53	4.60
HCT-15	1.08	3.05	>100
HT29	1.83	3.80	>100
KM 12	1.38	3.07	>100
SW-620	1.56	3.44	7.58
CNS Cancer			
SF-268	2.24	5.47	>100
SF-295	1.67	3.55	7.58
SF-539	1.67	3.31	6.57
SNB-19	2.31	7.30	>100
SNB-75	1.42	2.84	5.68
U251	1.24	2.70	5.88
Melanoma			
LOX IMVI	1.28	2.71	5.76
MALME-3M	1.76	3.54	7.11
M14	1.73	3.70	ND
MDA-MB-435	1.69	3.29	6.39
SK-MEL-2	2.12	4.40	ND
SK-MEL-28	1.87	4.00	ND
SK-MEL-5	1.87	3.81	7.73
UACC-257	3.04	7.98	>100
UACC-62	1.76	3.79	8.19
Ovarian Cancer			
IGROV1	1.88	4.25	ND
OVCAR-3	0.83	2.40	6.24
OVCAR-4	1.17	2.43	5.06
OVCAR-5	2.87	>100	>100
OVCAR-8	2.38	ND	>100
NCI/ADR-RES	2.11	7.56	>100
SK-OV-3	2.15	5.36	>100
Renal Cancer			
786-0	1.78	3.84	ND
A498	1.52	ND	>100
ACHN	1.72	4.36	>100
CAKI-1	1.54	2.95	ND
RXF 393	2.04	4.17	ND
SN12C	1.80	3.63	7.29
TK-10	2.91	5.97	>100

UO-31	1.48	2.95	ND
Prostate Cancer			
PC-3	1.11	2.55	ND
DU-145	1.18	2.90	7.11
Breast Cancer			
MCF7	1.59	4.58	>100
MDA-MB-231/ATCC	0.46	3.86	>100
HS 578T	2.34	6.33	>100
BT-549	1.33	ND	>100
T-47D	1.48	5.26	>100
MDA-MB-468	0.31	0.99	>100

ND: Not determined

2.4. Chemistry

2.4.1. Material and Methods

Melting points were taken in open capillaries on a Gallenkamp melting point apparatus (Weiss-Gallenkamp, Loughborough, UK) and are uncorrected. The IR spectra were recorded from potassium bromide disks with a FT device, Minia University. Elemental analyses were carried out at the Microanalytical Center, Cairo University, Egypt. The NMR spectra were measured in DMSO- d_6 on a Bruker AV-400 spectrometer (400 MHz for ^1H , 100 MHz for ^{13}C , and 40.55 MHz for ^{15}N); and the chemical shifts are expressed in δ (ppm), versus internal tetramethylsilane (TMS) = 0 for ^1H and ^{13}C , and external liquid ammonia = 0 for ^{15}N . Coupling constants are stated in Hz. Correlations were established using ^1H - ^1H COSY, and ^1H - ^{13}C and ^1H - ^{15}N HSQC and HMBC experiments. Mass spectra were recorded on a Finnigan Fab 70 eV, Institute of Organic Chemistry, Karlsruhe University, Karlsruhe, Germany. TLC was performed on analytical Merck 9385 silica aluminum sheets (Kieselgel 60) with Pf_{254} indicator; TLC's were viewed at $\lambda_{\text{max}} = 254 \text{ nm}$.

2.4.2. Starting materials

1,6-Disubstituted-quinoline-2,4-(1*H*,3*H*)-diones **1a-f** were prepared according to the literature [32,33]. 1,4-Naphthoquinone (**2**) (Aldrich) was used as received.

2.4.3. Products

Reaction of 1a-f with 1,4-naphthoquinone (2)

A mixture of 4-hydroxy-2(1*H*)-quinolinones **1a-f** (1 mmol) and 1,4-naphthoquinone (**2**, 0.158 g, 1 mmol) in ethanol/DMF (1:1) containing Et₃N (0.5 mL) was stirred at reflux temperature for 6-10 h. The reaction was monitored by TLC (until completion of the reaction). On cooling to room temperature, the precipitate was filtered off then washed with DMF (2 x 50 mL). The filtrate containing **4a-c**, was extracted by CH₂Cl₂, washed by cold water and the solvents were concentrated under vacuum. Compounds **3a-d** and **4a-c** were recrystallized from suitable solvents.

2,3-Bis(4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)naphthalene-1,4-dione (3a). This compound was obtained as yellow crystals (DMF), (0.315 g, 66%), mp. 350 °C. IR (KBr) ν = 3330 (NH), 1725, 1640 (CO), 1600, 1585 (Ar-C=C) cm⁻¹; NMR (Table 4); MS (FAB) *m/z* (%) 477 (M⁺, 20), 307 (35), 289 (18), 155 (30), 154 (100), 137 (67), 107 (18); *Anal.* Calcd for C₂₈H₁₆N₂O₆ (476.10) Calcd: C, 70.59; H, 3.38; N, 5.88. Found, C, 70.77; H, 3.35; N, 6.01.

2,3-Bis(4-hydroxy-6-methyl-2-oxo-1,2-dihydroquinolin-3-yl)naphthalene-1,4-dione (3b). This compound was obtained as yellow crystals (DMF/EtOH), (0.343 g, 68%), mp. 355 °C. IR (KBr) ν = 3340 (NH), 1735, 1625 (CO), 1605, 1590 (Ar-C=C) cm⁻¹; NMR (Table 5; MS (FAB) *m/z* (%) 505 (M⁺, 30), 335 (40), 154 (100), 107 (25). *Anal.* Calcd for C₃₀H₂₀N₂O₆ (504.13) Calcd: C, 71.42; H, 4.00; N, 5.55. Found, C, 71.28; H, 3.95; N, 5.41.

2,3-Bis(6-chloro-4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)naphthalene-1,4-dione (3c). This compound was obtained as violet crystals (DMF/EtOH), (0.349 g, 64%), mp.

345 °C. IR (KBr) ν = 3320 (NH), 1725, 1620 (CO), 1600, 1585 (Ar-C=C) cm^{-1} ; ^1H NMR δ_{H} 11.49 (bs; 2H, NH), 11.16 (b; 2H, OH), 8.13 (AA'XX', $J_{\text{AX}} = 7.6$, $J_{\text{AX}'} = 1.4$, $J_{\text{XX}} = 0$; 2H, H-4'), 7.96 (AA'XX, $J_{\text{AA}} = 7.0$, $J_{\text{AX}} = 7.6$, $J_{\text{AX}'} = 1.4$; 2H, H-5'), 7.77 (d, $J = 1.8$; 2H, H-5), 7.55 (dd, $J = 8.8$, 1.8; 2H, H-7), 7.18 (d, $J = 8.8$; 2H, H-8); ^{13}C NMR δ_{C} 182.24 (C-2'), 161.26 (C-2), 158.16 (C-4), 143.54 (C-1'), 136.82 (C-8a), 133.45 (C-5'), 132.47 (C-3'), 130.81 (C-7), 125.92 (C-4'), 125.28 (C-6), 122.31 (C-5), 116.85 (C-8), 116.36 (C-4^a), 106.08 (C-3); MS (FAB) m/z (%) 543/545/546 (M^+ , 18-22), 460 (15), 318 (27), 307 (27), 289 (47), 274 (42), 154 (100), 137 (68), 107 (20). *Anal.* Calcd for $\text{C}_{28}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}_6$ (545.02) Calcd: C, 61.67; H, 2.59; N, 5.14. Found, C, 61.80; H, 2.41; N, 5.09.

2,3-Bis(6-bromo-4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)naphthalene-1,4-dione

(3d). This compound was obtained as orange crystals (DMF), (0.444 g, 70%), mp. 348 – 350 °C. IR (KBr) ν = 3325 (NH), 1715, 1630 (CO), 1600, 1595 (Ar-C=C) cm^{-1} ; ^1H NMR δ_{H} 11.43 (s; 2H, NH), 11.14 (bs; 2H, OH), 8.12 (AA'XX', $J_{\text{AA}} = 7.1$, $J_{\text{AX}} = 7.4$, $J_{\text{AX}'} = 1.4$; 2H, H-4'), 7.88 (AAXX', $J_{\text{AX}} = 7.4$, $J_{\text{AX}'} = 1.4$, $J_{\text{XX}'} = 0$; 2H, H-5'), 7.77 (d, $J = 1.8$; 2H, H-5), 7.56 (dd, $J = 8.7$, 1.8; 2H, H-7), 7.17 (d, $J = 8.7$; 2H, H-8); ^{13}C NMR δ_{C} 182.55 (C-2'), 161.09 (C-2), 159.11 (C-4), 143.76 (C-1'), 136.09 (C-8a), 133.48 (C-5'), 132.33 (C-3'), 130.80 (C-7), 126.04 (C-4'), 125.97 (C-6), 121.98 (C-5), 116.88 (C-8), 116.40 (C-4a), 105.87 (C-3); MS (FAB) m/z (%) 633/634/636 (M^+ , 10-15), 307 (30), 289 (17), 154 (100), 137 (73), 136 (67), 107 (22). *Anal.* Calcd for $\text{C}_{28}\text{H}_{14}\text{Br}_2\text{N}_2\text{O}_6$ (631.92) Calcd: C, 53.02; H, 2.22; N, 4.42. Found, C, 53.17; H, 2.10; N, 4.58.

[Triethyl ammonium][2,3-bis(4-hydroxy-6-methyl-2-oxo-1,2-dihydroquinolin-3-yl)-naphthalene-1,4-dione] (4a). This compound was obtained as red crystals

(DMF/MeOH), (0.061 g, 10%), mp. >360 °C. IR (KBr) ν = 3210 (NH), 1679, 1625 (CO),

1605, 1590 (Ar-C=C) cm^{-1} , NMR (Table 6); MS (Fb) m/z (%) 606 (M^+ , 22), 504 (30), 307 (21), 154 (40), 136 (21), 102 (100); *Anal.* Calcd for $\text{C}_{36}\text{H}_{35}\text{N}_3\text{O}_6$ (605.25) Calcd: C, 71.39; H, 5.82; N, 6.94. Found, C, 71.54; H, 5.77; N, 7.12.

[Triethyl ammonium][2,3-bis(4-hydroxy-1-methyl-2-oxo-1,2-dihydroquinolin-3-yl)-naphthalene-1,4-dione] (4b). This compound was obtained as brown crystals (DMF), (0.472 g, 78%), mp. $>360^\circ\text{C}$. IR (KBr) $\nu = 3190$ (NH), 1676, 1635 (CO), 1600, 1585 (Ar-C=C) cm^{-1} ; ^1H NMR δ_{H} 8.9 (b; 2H, OH), 8.03 (AA'XX', $J_{\text{AX}} = 7.7$, $J_{\text{AX}'} = 1.3$, $J_{\text{XX}'} = 0$; 2H, H-4'), 8.01 (dd, $J = 7.2$, 1.4; 2H, H-5), 7.89 (AA'XX', $J_{\text{AA}} = 7.0$, $J_{\text{AX}} = 7.7$, $J_{\text{AX}'} = 1.3$; 2H, H-5'), 7.49 (dt, $J_d = 1.5$, $J_t = 7.7$; 2H, H-7), 7.31 (d, $J = 8.4$; 2H, H-8), 7.13 (t, $J = 7.4$; 2H, H-6), 3.42 (s; 6H, H-1a), 3.06 (q, $J = 7.3$; 6H, Et_3N), 1.14 (t, $J = 7.3$; 9H, Et_3N); ^{13}C NMR δ_{C} 184.07 (C-2'), 165.16 (C-4), 161.69 (C-2), 145.76 (C-1'), 139.43 (C-8a), 133.51 (C-5'), 132.66 (C-3'), 130.22 (C-7), 125.82 (C-4'), 124.75 (C-5), 120.41 (C-4a), 120.32 (C-6), 113.56 (C-8), 105.44 (C-3), 45.71 (Et_3N), 28.15 (C-1a), 8.73 (Et_3N); ^{15}N NMR δ_{N} 130.8 (N-1), 56.0 (Et_3N); MS (FAB) m/z (%) 606 (M^+ , 15), 504 (19), 307 (15), 154 (35), 136 (27), 102 (100). *Anal.* Calcd for $\text{C}_{36}\text{H}_{35}\text{N}_3\text{O}_6$ (605.25) Calcd: C, 71.39; H, 5.82; N, 6.94. Found, C, 71.53; H, 5.79; N, 7.11.

[Triethyl ammonium][2,3-bis(1-ethyl-4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)-naphthalene-1,4-dione] (4c). This compound was obtained as red crystals (DMF/EtOH), (0.520 g, 82%), mp. $320-321^\circ\text{C}$. IR (KBr) $\nu = 3320$ (NH), 1675, 1620 (CO), 1610, 1590 (Ar-C=C) cm^{-1} ; ^1H NMR ($\text{DMSO}-d_6$) δ_{H} = 1.04 (ABX₃, $J_{\text{AX}} = J_{\text{BX}} = 7.0$; 3H, H-1b), 1.09 (t, $J = 7.3$; 9H, Et_3N), 2.99 (q, $J = 7.2$; 6H, Et_3N), 3.99 (ABX₃, $J_{\text{AB}} = 14.3$, $J_{\text{BX}} = 7.1$; 2H, H-1a), 4.03 (ABX₃, $J_{\text{AB}} = 14.3$, $J_{\text{AX}} = 7.1$; 2H, H-1a), 7.04 (t, $J = 7.4$; 2H, H-6), 7.28 (d, $J = 8.4$; 2H, H-8), 7.42 (dt, $J_d = 1.5$, $J_t = 7.7$; 2H, H-7), 7.81 (AA'XX', $J_{\text{AA}'} = 7.2$, $J_{\text{AX}} = 7.5$,

$J_{AX'} = 1.5$; 2H, H-5'), 7.95 (dd, $J = 7.7, 1.3$; 2H, H-5), 7.96 (AA'XX', $J_{AX} = 7.5, J_{AX'} = 1.5, J_{XX'} = 0$; 2H, H-4'), 9.0 ppm (b; 2H, OH); ^{13}C NMR (DMSO- d_6) $\delta_{\text{C}} = 8.79$ (Et₃N), 13.01 (C-1b), 35.36 (C-1a), 45.72 (Et₃N), 105.38 (C-3), 113.34 (C-8), 120.09 (C-6), 120.51 (C-4a), 125.05 (C-5), 125.86 (C-4'), 130.89 (C-7), 132.64 (C-3'), 133.45 (C-5'), 138.39 (C-8a), 145.79 (C-1'), 161.33 (C-2), 165.01 (C-4), 184.02 ppm (C-2'); ^{15}N NMR (DMSO- d_6) $\delta_{\text{N}} = 55.4$ (Et₃N), 144.6 ppm (N-1); Ms (Fab) m/z (%) 634 (M⁺, 18), 533 (20), 307 (18), 154 (10), 102 (100). Anal. Calcd for C₃₈H₃₉N₃O₆ (633.28): Calcd: C, 72.02; H, 6.20; N, 6.63. Found, C, 72.18; H, 6.34; N, 6.55.

X-ray Crystal Structure Determinations of 4c (include from the SI)

The single-crystal X-ray diffraction study was carried out on a Bruker D8 Venture diffractometer with Photon100 detector at 123(2) K using Cu-K α radiation ($\lambda = 1.54178$ Å). Direct Methods (SHELXS-97) [34] were used for structure solution and refinement was carried out using SHELXL-2014 (full-matrix least-squares on F^2) [35]. Hydrogen atoms were localized by difference electron density determination and refined using a riding model (H(O, N) free). A semi-empirical absorption correction was applied. The refinement with the listed atoms show residual electron density in two voids due to a heavily disordered water which could not be refined with split atoms. Therefore, the option "SQUEEZE" of the program package PLATON [36] was used to create a hkl file taking into account the residual electron density in the void areas. Therefore the atoms list and unit card do not agree (see cif-file for details).

4c: red crystals, C₃₂H₂₃N₂O₆ · C₆H₁₀N · 0.5 H₂O, $M_r = 642.73$, crystal size $0.18 \times 0.06 \times 0.03$ mm, monoclinic, space group $P2_1/n$ (No. 14), $a = 15.8124(5)$ Å, $b = 12.3817(4)$ Å, $c = 18.4717(6)$ Å, $\beta = 107.089(2)^\circ$, $V = 3456.8(2)$ Å³, $Z = 4$, $\rho = 1.235$

Mg/m³, $\mu(\text{Cu-K}\alpha) = 0.687 \text{ mm}^{-1}$, $F(000) = 1364$, $2\theta_{\text{max}} = 144.6^\circ$, 28835 reflections, of which 6783 were independent ($R_{\text{int}} = 0.045$), 430 parameters, $R_1 = 0.062$ (for 5318 $I > 2\sigma(I)$), $wR_2 = 0.169$ (all data), $S = 1.05$, largest diff. peak / hole = 0.815 (in the ammonium cation)/ -0.413 e Å⁻³. CCDC 1568166 (**4c**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif. Due to the bad quality of the data of 2a-Br (sb874) the data were not deposited with The Cambridge Crystallographic Data Centre.

Computational Methodology

Molecular docking into ERK2 receptor was carried out for the synthesized compounds as potential ERK2 inhibitors. All docking calculations were performed using Autodock4.2 software [37]. Receptor coordinates were taken from the crystal structure of ERK2 in complex with *N*-[(1*S*)-1-(3-Chloro-4-fluorophenyl)-2-hydroxyethyl]-2-(tetrahydro-2*H*-pyran-4-yl-amino)-5,8-dihydropyrido[3,4-*d*]pyrimidine-7(6*H*)-carboxamide (2SH) (PDB code: 4O6E [38]). The receptor was cleaned of water molecules, ions and the ligand 2SH; as well the missing residues were constructed with MODELLER software [39]. The protonation state of the receptor was investigated using H++ server [40]. AutoDock protocol [41] was then followed to prepare pdbqt file for ERK2 receptor. The related Autodock parameters were set as follows: 250 genetic algorithm (GA) runs, each with population size of 300, 25000000 energy evaluations, and a maximum of 27,000 generations per GA run. The docking grid was of dimension 70 Å × 70 Å × 70 Å with the spacing of 0.375 Å and located at 16.20, 7.50, 12.40 (XYZ

coordinates). All examined compounds were energetically minimized using MMFF94S force field with the help of SZYBKI software [42]. Gasteiger atomic charges were assigned for the minimized compounds [43].

3. Results and Discussion

3.1. Chemistry

When a mixture of 2 equivalent of 4-hydroxy-2-(1*H*)-quinolinones **1a-f** and one equivalent of 1,4-naphthoquinone (**2**) in ethanol/dimethylformamide (1:1) as a solvent and 0.5 ml Et₃N was stirred at reflux temperature for 6-10 h, the bis-quinolinylquinones **3a-d** and **4a-c** were obtained (Scheme 1).

Scheme 1. Reaction of 4-hydroxy-2-quinolinones **1a-f** with 1,4-naphthoquinone (**2**)

For example, compound **3a** was obtained in 66% yield after recrystallization (see the experimental section and Table 4). The assignment of the structure of compounds **3a-d** was confirmed by NMR analysis of the expected chemical shifts and 2D NMR spectra. In the ¹H NMR spectrum of **3a**, eight 2H signals were distinguished: two exchangeable singlets, a four-spin coupled system for the quinoline system, and an AA'XX' system for the naphthoquinone. The ¹³C spectrum has 14 lines, consistent with either **3a** or **3a'**

(Figure 7). The nitrogen atom resonates at $\delta_N = 142.1$ ppm; it gives HSQC correlation with a proton at $\delta_H = 11.32$ ppm, which therefore is assigned as the NH. This correlation leads to the assignment of quinolone tautomer **3a** rather than **3a'**. The nitrogen gives HMBC correlation to the doublet at $\delta_H = 7.15$ ppm, which is assigned as H-8; its attached carbon resonates at $\delta_C = 115.23$ ppm. The NH gives strong HMBC correlation with C-8 and with a carbon at $\delta_C = 105.42$ ppm, assigned as C-3; these are both three-bond correlations.

Table 4. Spectroscopic data of compound **3a**

¹H NMR:		¹H-¹H COSY:	Assignment:	
11.38 (s; 2H)			NH	
10.86 (bs; 2H)			OH	
8.10 (AA'XX', $J_{AX} = 7.5$, $J_{AX'} = 1.6$, $J_{XX'} = 0$; 2H)		7.92	H-4'	
7.92 (AA'XX', $J_{AA'} = 7.4$, $J_{AX} = 7.5$, $J_{AX'} = 1.6$; 2H)		8.10	H-5'	
7.80 (d, $J = 7.9$; 2H)		7.47, 7.15, 7.14	H-5	
7.47 (dt, $J_d = 1.2$, $J_t = 7.7$; 2H)		7.80, 7.15, 7.14	H-7	
7.15 (d, $J = 8.4$; 2H)		7.80, 7.47, 7.14	H-8	
7.14 (t, $J = 8.3$; 2H)		7.80, 7.47, 7.15	H-6	
¹³C NMR:	HSQC:	HMBC:	Assignment:	Calc:
182.44		8.10	C-2'	187.0
161.74		11.38	C-2	163.8
159.24		7.80, 7.15	C-4	162.3
144.01			C-1'	147.9
138.09		11.38, 7.80, 7.47, 7.14	C-8a	135.9
133.48	7.92	8.10	C-5'	134.8
132.51		7.92	C-3'	137.9
130.89	7.47	7.80	C-7	127.9
125.94	8.10	7.92	C-4'	130.2
123.30	7.80	7.47	C-5	126.4
121.23	7.14	7.15	C-6	124.0
115.23	7.15	11.38, 7.47, 7.14	C-8	120.3
114.89			C-4a	126.8
105.42		11.38	C-3	98.1
¹⁵N NMR:	HSQC:	HMBC:	Assignment:	
143.1	11.38	7.15	N-1	

The **NH** also gives *weak* HMBC correlation with carbons at $\delta_C = 161.74$ and 138.09 ppm, assigned as C-2 and C-8a respectively; these are both two-bond correlations. Also, C-8a gives HMBC correlation with all protons in the four-spin system *except* H-8. In addition to other correlation with the triplet at $\delta_H = 7.14$ ppm, which assigned as H-6; this is a three-bond correlation. The attached carbon appears at $\delta_C = 121.23$ ppm, and gives HMBC correlation back to H-8. The other doublet in the four-spin system (at $\delta_H = 7.80$ ppm) must be H-5 and the other triplet (at $\delta_H = 7.47$ ppm) must be H-7; their attached carbons appear $\delta_C = 123.30$ (C-5) and 130.89 ppm (C-7), respectively. C-5 gives HMBC correlation with H-7, and C-7 gives HMBC correlation with H-5; these are three-bond correlations. H-5 and H-8 give HMBC correlation with a carbon at $\delta_C = 159.24$ ppm, assigned based on chemical shift as C-4 rather than C-4a; the carbon at $\delta_C = 114.89$ ppm, is assigned as C-4a. Moreover, the other aromatic protons are the AA'XX' system of the naphthoquinone substructure. The downfield signal at $\delta_H = 8.10$ gives HMBC correlation with the quinone carbonyl carbon at $\delta_C = 182.44$ ppm; accordingly, these protons are assigned as H-4' and the upfield signal at $\delta_H = 7.92$ ppm, is assigned as H-5'. The attached carbons appear at $\delta_C = 125.94$ (C-4') and 133.48 ppm (C-5'). A carbon at $\delta_C = 132.51$ gives HMBC correlation with H-5', and is assigned as H-3'. The remaining carbon at $\delta_C = 144.01$ ppm, is assigned as C-1'. The NMR spectroscopic data of **3a** are shown in Table 4.

Figure 7. Alternative structure of **3a**, assigned as **3a'**

In case of the reaction between **1b** with **2**, products **3b** and **4a** were separated (Scheme 1). Compound **3b** was obtained as the major product (68%), whereas salt **4a** was in 10% yield percentage. Compound **3b** (the distinctive carbons were assigned as shown in Figure 8) is the 1:2 adduct of 1,4-naphthoquinone (**2**) and 6-methylquinoline-2,4-dione (**1b**). The ^1H spectrum consists of two 6H methyl signals and seven 2H signals: two exchangeable singlets, a three-spin coupled system for the quinoline system, and an AA'XX' system for the naphthoquinone. The ^{13}C spectrum has 15 lines, consistent with **3b**. The assignment of a quinolone tautomer, rather than a quinolinol, rests on the observation of ^1H - ^{15}N HSQC correlation.

Figure 8. Distinctive carbons and nitrogen of compound **3b**

Table 5. Spectroscopic data of compound **3b**

^1H NMR:	^1H - ^1H COSY:	Assignment:
11.32 (bs; 2H)	7.60	NH
10.71 (b; 2H)		OH
8.10 (AA'XX', $J_{AA'} = 7.1$, $J_{AX} = 7.7$, $J_{AX'} = 1.4$; 2H)	7.93	H-4'
7.93 (AA'XX', $J_{AX} = 7.7$, $J_{AX'} = 1.4$, $J_{XX'} = 0$; 2H)	8.10	H-5'

7.60 ("s"; 2H)	7.30	H-5
7.30 (dd, $J = 8.3, 1.3$; 2H)	7.60, 7.07	H-7
7.07 (d, $J = 8.4$; 2H)	7.30	H-8
2.29 (s; 6H)	7.60, 7.30	H-6a

¹³ C NMR:	HSQC:	HMBC:	Assignment:	Calc:
182.47			C-2'	187.0
161.76			C-2	163.8
159.04		7.60, 7.07	C-4	162.8
144.14			C-1'	147.9
136.05		7.60, 7.30	C-8a	132.9
133.52	7.93	8.10	C-5'	134.8
132.48	7.30		C-7	128.6
130.34		7.07, 2.29	C-6	133.2
125.96	8.10		C-4'	130.2
122.72	7.60	2.29	C-5	127.1
115.11			C-4a	126.7
114.89	7.07	11.32	C-8	120.2
105.50		11.32	C-3	98.1
20.49	2.29		C-6a	21.3

¹⁵ N NMR:	HSQC:	HMBC:	Assignment:
142.1	11.32		N-1

The methyl protons and the three-spin system of the 6-substituted quinoline substructure are distinctive at δ_H 2.29 (H-6a), 7.60 (apparent singlet, actually a doublet with a small J ; H-5), 7.30 (dd; H-7), and 7.07 (doublet, large J ; H-8) respectively; they give HSQC correlation with their respective attached carbons at δ_C 20.49 (C-6a), 122.77 (C-5), 132.48 (C-7), and 114.89 (C-8). A carbon at δ_C 130.34 gives HMBC correlation with H-6a and H-8, and is assigned as C-6; a carbon at δ_C 136.05 gives HMBC correlation with H-5 and H-7, and is assigned as C-8a, which is three bonds from each of these protons. A carbon at δ_C 159.04 gives HMBC correlation with H-5 and H-8; these correlations would be consistent with either C-4a (two and three bonds) or C-4 (three and four), but the carbon is assigned based on chemical shift as C-4. The nitrogen atom resonates at δ_N 142.1; it gives HSQC correlation with a proton at δ_H 11.32, which therefore is assigned as

the NH. This proton gives HMBC correlation with one or both of C-8 and the carbon at δ_C 115.11; based partly on this correlation and partly on chemical shift, this carbon is assigned as C-4a. The NH also gives HMBC correlation with a carbon at δ_C 105.50, assigned as C-3. The other aromatic protons are the AA'XX' system of the naphthoquinone substructure. On chemical-shift grounds, the downfield signal at δ_H 8.10 is assigned as H-4' and the upfield signal at δ_H 7.93 is assigned as H-5'; the attached carbons appear at δ_C 125.96 (C-4') and 133.52 (C-5'). A carbon at δ_C 132.12 gives HMBC correlation with H-5', and is assigned as H-3'. The remaining carbons at δ_C 182.47, 161.76, and 144.14 are assigned, on chemical-shift grounds, as C-2', C-2, and C-1' respectively.

Based on NMR spectroscopic data (Table 6), the structure of **4a** was defined as triethylammonium salt of 2,3-bis(quinolon-3-yl)naphthoquinone (Scheme 1). The triethyl amine salt was formed between triethyl amine molecule and the hydroxyl groups (Figure 9).

Figure 9. Structure of side product **4a** obtained by the reaction of **1b** with **2**

The ^1H spectrum consists of one 6H methyl signal and seven 2H signals: two exchangeable singlets, a three-spin coupled system for the quinoline system, and an AA'XX' system for the naphthoquinone. An equivalent of triethylamine is present. The ^{13}C spectrum has 17 lines, consistent with a triethylammonium salt of **4a**. The ^1H - ^{15}N

HSQC correlation observed in sample **4a** is absent, but the nitrogen has the same hybridization, so the compound is assigned as a quinolone not a quinolinol. We believe the triethylammonium ion to be associated with the hydroxyl groups not the nitrogens, because C-4 resonates over 7 ppm downfield of its position in **4a**, while the chemical shift of N-1 changes by less than 1 ppm. The methyl protons and the three-spin system of the 6-substituted quinoline substructure are distinctive at δ_{H} 2.30 (H-6a), 7.65 (apparent singlet, a doublet with a small J ; H-5), 7.17 (dd; H-7), and 7.02 (doublet, large J ; H-8) respectively; they give HSQC correlation with their respective attached carbons at δ_{C} 20.55 (C-6a), 123.93 (C-5), 132.68 (C-7), and 114.34 (C-8). A carbon at δ_{C} 129.71 gives HMBC correlation with H-6a and H-8, and is assigned as C-6; a carbon at δ_{C} 136.59 gives HMBC correlation with H-5 and H-7 and H-8, and is assigned as C-8a. A carbon at δ_{C} 166.24 gives HMBC correlation with H-5 and H-8, and is assigned as C-4; this is over 7 ppm downfield of its shift in **4a**. The only other carbon more than 2 ppm from its position in **4a** is C-4a at δ_{C} 119.14, shifted by 4 ppm. The nitrogen atom resonates at δ_{N} 142.9, only 0.8 ppm downfield of its shift in **4a**; it does not give HSQC correlation, but gives HMBC correlation with H-8. Also, H-8 gives HMBC correlation with C-4a. The carbon at δ_{C} 105.90 is assigned, based on chemical shift, as C-3. The other aromatic protons are the AA'XX' system of the naphthoquinone substructure. On chemical-shift grounds, the downfield signal at δ_{H} 8.01 is assigned as H-4', and the upfield signal at δ_{H} 7.87 is assigned as H-5'; the attached carbons appear at δ_{C} 125.78 (C-4') and 133.39 (C-5'). A carbon at δ_{C} 132.54 gives HMBC correlation with H-5', and is assigned as H-3'. The remaining carbons at δ_{C} 183.98, 162.66, and 146.13 are assigned, on chemical-shift

grounds, as C-2', C-2, and C-1' respectively; C-2' also gives HMBC correlation with H-4'.

Table 6. Spectroscopic data of compound **4a**.

¹ H NMR:	¹ H- ¹ H COSY:	Assignment:
10.56 (bs; 2H)		NH
8.01 (AA'XX', $J_{AX} = 7.7$, $J_{AX'} = 1.4$, $J_{XX'} = 0$; 2H)	7.87	H-4'
7.87 (AA'XX', $J_{AA'} = 7.0$, $J_{AX} = 7.7$, $J_{AX'} = 1.4$; 2H)	8.01	H-5'
7.65 ("s"; 2H)	7.17, 7.02, 2.30	H-5
7.17 (dd, $J = 8.3$, 1.8; 2H)	7.65, 7.02, 2.30	H-7
7.02 (d, $J = 8.2$; 2H)	7.65, 7.17, 2.30	H-8
5.1 (b; 2H)		OH
3.01 (q, $J = 7.3$; 6H)	1.11	Et ₃ N
2.30 (s; 6H)	7.65, 7.17, 7.02	H-6a
1.11 (t, $J = 7.3$; 9H)	3.01	Et ₃ N

¹⁵ N NMR:	HSQC:	HMBC:	Assignment:
142.9		7.02	N-1
60.7		1.11	Et ₃ N

¹³ C NMR:	HSQC:	HMBC:	Assignment:
183.98		8.01	C-2'
166.24		7.65, 7.02	C-4
162.66			C-2
146.13			C-1'
136.59		7.65, 7.17, 7.02	C-8a
133.39	7.87	8.01, 7.87	C-5'
132.68	7.17		C-7
131.54		7.87	C-3'
129.71		7.02, 2.30	C-6
125.78	8.01	7.87	C-4'
123.93	7.65	2.30	C-5
119.14		7.02	C-4a
114.34	7.02		C-8
105.90			C-3
45.70	3.01	3.01, 1.11	Et ₃ N
20.55	2.30	7.65	C-6a
8.88	1.11	3.01	Et ₃ N

Compound **4c** was formed from the reaction of **1f** with **2** and by the same way formed as in case of **4a** and **4b**. Et₃N molecule is engaged in the salt with hydroxyl groups.

Spectroscopic parameters appear in the Experimental Section; X-ray structural analysis proved the structure of **4c** as shown in Figure 10.

Figure 10. X-ray structural analysis of **4c** (displacement parameters are drawn at 50% probability level)

3.2. Biology

As shown in Table 2, it is obvious that compound **3d** exhibited marvelous cytotoxic activity against all tested human cancer cell lines with GI_{50} values ranging between (0.26 – 0.93 μM) except for colon cancer cell line HT29 (GI_{50} = 3.87 μM), ovarian cancer cell line OVCAR-5 (GI_{50} = 4.19 μM) and colon cancer cell line COLO 205 (GI_{50} = 16.7 μM). Moreover, this compound is proved to be sensitive towards almost the tested cell lines, particularly non-small cell lung, CNS, melanoma and renal cancer cell lines where they showed the highest growth inhibitory activity. In the case of non-small cell lung cancer, CNS and renal cancer cell lines the average GI_{50} value was around 0.38 μM , while the GI_{50} value of melanoma cancer cell lines were around 0.45 μM . Meanwhile, the range of TGI value was (0.45 – 16.7 μM). The LC_{50} values ranged between 1.09 and 16.7 μM . The cytotoxic effects were strongest in the non-small cell lung cancer line HOP-62 (1.50

μM), the colon cancer cell line HCT-15 ($1.47\ \mu\text{M}$), the CNS cancer lines SNB-75 ($1.11\ \mu\text{M}$) and SF-295 ($1.18\ \mu\text{M}$), the melanoma cancer cell line LOX IMVI ($1.09\ \mu\text{M}$) and the renal cancer lines RXF 393 and UO-31 ($1.41\ \mu\text{M}$).

The results recorded in Table 3 indicated that compound **3b** demonstrated broad spectrum anti-tumor activity against the nine tumor sub-panels tested. It exhibited strong growth inhibitory activity against all the tested human cancer cell lines, with GI_{50} values ranged between ($0.31\text{--}3.04\ \mu\text{M}$) and the TGI values ranged between ($0.99\text{--}12.8\ \mu\text{M}$). A noticeable sensitivity profile was observed for leukemia cancer cell lines (GI_{50} values ranging from $0.38\text{--}2.32\ \mu\text{M}$). Furthermore, this compound achieved a highly active growth inhibitory activity against non-small cell lung, colon, CNS, melanoma, ovarian and renal cancers, where an almost similar average GI_{50} ranging from ($1.63\text{--}1.92\ \mu\text{M}$) were recorded. Similarly, the tested compound experienced remarkable sensitivity towards prostate and breast cancer cell lines, where the average GI_{50} was around $1.2\ \mu\text{M}$. Notably, the highest growth inhibitory activity was observed against the breast cancer cell line MDA-MB-468 with GI_{50} value $0.31\ \mu\text{M}$ and the strongest cytotoxic effects were observed in the non-small cell lung cancer lines NCI-H460 ($\text{LC}_{50} = 6.44\ \mu\text{M}$), A549/ATCC ($\text{LC}_{50} = 8.46\ \mu\text{M}$) and NCI-H23 ($\text{LC}_{50} = 10.4\ \mu\text{M}$), the colon cancer cell lines HCT-116 ($\text{LC}_{50} = 4.60\ \mu\text{M}$), SW-620 ($\text{LC}_{50} = 7.58\ \mu\text{M}$), all CNS cancer lines except SF-268, SNB-19, the melanoma lines except UACC-257, the ovarian cancer lines OVCAR-4 ($\text{LC}_{50} = 5.06\ \mu\text{M}$) and OVCAR-3 ($\text{LC}_{50} = 6.24\ \mu\text{M}$), the renal cancer line SN12C ($\text{LC}_{50} = 7.29\ \mu\text{M}$) and the prostate cancer line DU-145 ($\text{LC}_{50} = 7.11\ \mu\text{M}$).

4. Conclusion

In this paper, we shed the biologist towards the importance of the biological activity of 2-quinolones. Many interesting classes of this type of compounds would be valuable for drug future discovery. Herein we have a success to synthesize and in good yields two novel series of 2,3-bis-quinolinyl naphthalene-1,4-diones and bis-quinolinone triethylammonium salts. Compounds **3a**, **3b** and **4c** inhibited ETS-1 phosphorylation by ERK2 in a dose dependent manner. Moreover, Compound **4c** showed selectivity for ERK2 inhibition together with ATP-competitive inhibition which was revealed by its binding pattern in the ATP-binding site of ERK2 receptor. However, an additional structure optimization is required for enhancing its pharmacokinetic profile. Compound **3d** exhibited marvelous cytotoxicity towards approximately all NCI tumor cell lines tested showing negative growth percentages. The cytotoxic effect of compound **3d** was apparent towards approximately all NCI tumor cell lines tested showing negative growth percentages. Compound **3b** and **3d** demonstrated broad spectrum anti-tumor activity against the nine tumor sub-panels tested in NCI *in vitro* five dose testing with GI₅₀ values ranging between (0.31-3.04 μ M), and (0.26 – 0.93 μ M), respectively and TGI values ranging between (0.99- 12.8 μ M) and (0.45 – 16.7 μ M), respectively. Finally, the newly synthesized compounds represent good starting point for the development of new cytotoxic lead compounds.

5. References

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this profile
is encouraging, development as therapeutic agents will require
additional optimization efforts.