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Yun, Young Ju

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Graphical Abstract

Triplet-Triplet Annihilation Photon-Upconversion in Hydrophilic Media with Biorelevant Cholesteryl Triplet Energy Acceptors

Young Ju Yun^{‡,#}, Jussi Isokuortti^{◇,#}, Timo Laaksonen^{◇,†}, Nikita Durandin[◇] and A. Jean-Luc Ayitou^{‡,*}

[‡] Department of Chemistry, Illinois Institute of Technology, Chicago, IL 60616, United States.

[◇] Engineering and Natural Sciences, Tampere University, Korkeakoulunkatu 8, 33720 Tampere, Finland.

[†] Division of Pharmaceutical Biosciences, Faculty of Pharmacy, University of Helsinki, Viikinkaari 5 E, 00014 Helsinki, Finland.

These authors have contributed equally to this work

*Corresponding author E-mail

aayitou@iit.edu

ABSTRACT: We report two new biorelevant cholesteryl-based triplet energy acceptors, derivatives of **DPA** or 9,10-diphenylanthracene (**C–DPA** and **C₂–DPA**). Using two different triplet sensitizers: **QDN** ($E_T \approx 1.77$ eV) and **PdTPP** ($E_T \approx 1.78$ eV), we were able to achieve both endothermic (with **QDN**) and exothermic (with **PdTPP**) triplet sensitization of **DPA**, **C–DPA** and **C₂–DPA** in hydrophilic PEG200 media. While the maximum rate of triplet energy transfer (TET) was achieved with **PdTPP** and **DPA** ($k_{TET} = 4.7 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$), for the cholesteryl-based acceptors, we found that the kinetic of the TET process was dependent upon the concentration of the acceptor. For **PdTPP/C–DPA** pair, the rate of the dynamic triplet energy quenching was $k_{TET} = 1.9 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$; however, at higher concentrations of the quencher, the system reached a stationary state due to formation of self-assembled sub-domains of **C–DPA** that likely slowed the TET process. It was also found that this aggregation of **C–DPA** in PEG200 led to a 3.5 folds increase in the I_{th} compared to 133 mW cm^{-2} for **DPA**. Subsequently, we estimated the Φ_{UC} for these donor/acceptor pairs: **QDN/DPA**, **PdTPP/DPA**, and **PdTPP/C–DPA**. With respect to the estimated I_{th} , we found that the uncorrected (for inner filter effect) quantum yields of TTA-UC were $2 \leq \Phi_{UC} \leq 12\%$.

INTRODUCTION:

Triplet-triplet annihilation photon-upconversion (TTA-UC) is an elegant photophysical process that allows to amplify photon energy for applications at the interface of solar energy conversion, optoelectronic science, and bio-imaging.¹⁻⁷ It has been well-documented that TTA-UC is a bi-molecular process, where the triplet energy donor/sensitizer absorbs low-energy light and the triplet energy acceptor emits higher-energy photons.^{8,9} Under this condition, TTA-UC has been shown to be a promising process vis-à-vis bio-imaging applications, where low-energy light is amplified and the resulting emission could be tailored for *in vivo* photo-excitation of biological markers¹⁰ or even achieve photochemical/biochemical transformations.^{11,12} In this scenario, TTA-UC chromophoric systems can be systematically functionalized with biocompatible/biorelevant groups such as cholesteryl in order to facilitate the integration of the modified donor/acceptor dyes into biological environments. In addition, the derivatized dyes could be used to study the underlying photo-kinetics and efficiency of the processes involved in TTA-UC in lipids and membranes. To study TTA-UC in lipids and membranes, it's also important to mimic the relevant conditions *viz.* viscosity, hydro-philicity/phobicity, and polarity of biological environments. A recent strategy in this regards proposed to achieve the TTA-UC processes in PEG200, which is a hydrophilic polymeric liquid that mimics cell membrane structures and intracellular conditions.¹³⁻¹⁶ With this in mind, we are hereby reporting two new cholesteryl-based 9,10-diphenylanthracene (**DPA**) derivatives (**C-DPA** and **C₂-DPA**) (Scheme 1) which we used as triplet energy acceptors for TTA-UC in PEG200. The present study also aimed at deciphering the effect of the biorelevant cholesteryl group on donor•••acceptor interactions and the resulting TTA-UC process.

A) Donors / Sensitizers

QDN

PdTPP

B) Acceptors / Annihilators

DPA

C-DPA

C₂-DPA

Scheme 1. Chemical structures of sensitizers (**QDN** and **PdTPP**) and annihilators (**DPA**, **C-DPA**, and **C₂-DPA**).

Recently, a new family of triplet sensitizers: quinoidal naphthyl thioamides (**QDN**) (Scheme 1) were reported by our group.¹⁷ **QDN** are structurally compatible with over-the-counter acceptors/fluorescent polyaromatic hydrocarbons such as perylene^{18,19} and 9,10-diphenylanthracene (**DPA**) which are routinely employed for TTA-UC processes. We had previously established that the triplet energy of **QDN** ($E_T \approx 1.77$ eV, in 77K EtOH:DCM glassy matrix) is suitable for an exothermic energy transfer to perylene ($E_T \approx 1.53$ eV); however, it was unexpected that **QDN** could be used to sensitize **DPA** ($E_T \approx 1.78$ eV)²⁰ in deoxygenated PEG200, since its measured E_T value at room temperature in that medium was estimated at 1.67 eV, far below the triplet energy of **DPA**. It's worth noting that, in order to maintain the deoxygenated state

of all of our photo-samples, 30 mM of oleic acid was added to the PEG200 material (oleic acid was used as oxygen scavenger). Triplet sensitization study of **DPA** using **QDN** and/or another conventional sensitizer 5,10,15,20-TetraPhenylPorphyrinato Palladium(II) (**PdTTP**) with $E_T \approx 1.78 \text{ eV}^{20}$ revealed that both endothermic **QDN**→**DPA** and exothermic **PdTTP**→**DPATET** could be achieved in biorelevant PEG200 media. Armed with these exciting findings, we furthered the TTA-UC studies with the new cholesteryl-based triplet energy acceptors **C-DPA** and **C₂-DPA**. Expectedly, the cholesteryl unit would not only enhance interactions of the hydrophobic donor•••acceptor in the hydrophilic PEG200 medium, but also induce the formation of self-assembled sub-domains at higher concentrations of the donor/acceptor chromophoric systems.

RESULTS & DISCUSSION:

The synthetic procedures for all precursors of **C-DPA** and **C₂-DPA** are extensively described in the Supporting Information (Scheme S1). **C-DPA** and **C₂-DPA** were synthesized following Suzuki coupling reaction conditions, where cholesteryl 4-bromobenzyl ether (**Br-PhCH₂-O-Chol**) was reacted with 10-phenyl-9-anthracene boronic acid for **C-DPA** and 9,10-Anthracenediboronic acid bis(pinacol) ester for **C₂-DPA** (Supporting Information, Scheme S1-S4). **C-DPA** and **C₂-DPA** were successfully characterized by NMR (Supporting Information, Figure S1-S8), thermogravimetry (Supporting Information, Figure S12), and single-crystal XRD (Supporting Information, Figure S13).

Figure 1. UV-vis profiles of (A) **QDN** and **PdTPP** and (B) **DPA**, **C-DPA**, and **C₂-DPA** in PEG200. (C) Emission profiles of **QDN** and **PdTPP** and (D) **DPA**, **C-DPA**, and **C₂-DPA** recorded in PEG200 with O.D. = 0.2; at the excitation wavelength 532 nm for **QDN** and **PdTPP**, 357 nm for **DPA**, 380 nm for **C-DPA** and **C₂-DPA**.

UV-vis absorption profiles of **C-DPA** and **C₂-DPA**, recorded in PEG200, revealed the same transitions features that are seen in the spectrum of parent **DPA** (Figure 1B). The absorption features of **C-DPA** was precisely the same as parent **DPA**, which is not surprising. However, the spectrum of **C₂-DPA** exhibits a slight hypsochromic-shift compared to **C-PDA** or **DPA**. The positive offset in the spectrum of **C₂-DPA** is suggestive of self- or H-aggregation, which could be problematic for an efficient donor•••acceptor interaction in PEG200. Importantly the positive spectral shift was not observed in the absorption spectrum of **C₂-DPA** in DCM solution (Supporting information, Figure S11). Similarly, the steady-state emission profiles of **C-DPA** and

C₂–DPA depict the typical vibronic bands of **DPA** with no apparent/trivial spectral shift. With this result, we ascertain that the cholesteryl unit has little impact on the optoelectronic transitions of the new acceptors (Figure 1D). Nevertheless, the emission spectra recorded in PEG200 are slightly red-shifted presumably due to aggregation or in the hydrophilic PEG200 medium. (Supporting information, Figure S9–S11).

Figure 2. Stern–Volmer plots of (A) **QDN** and **DPA** (B) **PdTPP** and **DPA** (C) **PdTPP** and **C–DPA** in PEG200 with the resulting Stern–Volmer constants (K_{SV}) and TET rate constants (k_{TET}). (D) Intensity-dependent delayed emission of **C–DPA** in deoxygenated PEG200 using **PdTPP** as

the sensitizer and excitation wavelength of 532 nm. *Note: The phosphorescence samples were recorded in deoxygenated PEG200 in the presence of 30 mM of oleic acid.*

Next, we performed Stern-Volmer (S-V) quenching experiments, using conventional triplet energy acceptors e.g. **DPA**, to establish the best sensitizer (**QDN** vs. **PdTPP**) for efficient TET with both **C-DPA** and **C₂-DPA**. The quenching experiment was monitored by recording the phosphorescence of the two sensitizers in the presence of the acceptor/annihilator(s) with varying concentrations. The quenching data was analyzed following equations (1) and (2).²¹

$$K_{SV} = K_q \tau_0. \quad (1)$$

Where K_q is the quenching rate constant, τ_0 is the intrinsic phosphorescence lifetime of the sensitizer, and K_{SV} is the Stern-Volmer constant.

For dynamic quenching mechanism,²² we presumed that

$$\frac{\tau_0}{\tau} - 1 = K_q \tau_0 [Q] \quad (2)$$

Where τ is the phosphorescence lifetime of the sensitizer in the presence of the acceptor/annihilator(s), and $[Q]$ is the concentration of acceptor/annihilator(s).

Figure 2 shows the S-V plots with respect to the concentration of annihilator/quencher. The slope of the S-V plot is determined as $K_{SV} = K_q \tau_0$ (Eq. 1). Next, the rate of triplet energy transfer (k_{TET}) was computed from the following equation:

$$k_{TET} = \frac{K_{SV}}{\tau_0} \quad (3)$$

τ_0 for **QDN** was 160 μs , and that of **PdTPP** was 720 μs (both values were measured in in PEG200). We then established that for **DPA**, the TET rate constant $k_{\text{TET}} = 8.1 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ for **QDN** and $k_{\text{TET}} = 4.7 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ for **PdTPP** (Figures 2A & 2B). As one can see, the k_{TET} for **QDN**→**DPA** revealed an endothermic energy transfer with $\Delta E_{\text{T}} = -0.046 \text{ eV}$, whereas **PdTPP**→**DPA** TET with $\Delta E_{\text{T}} = -0.01 \text{ eV}$ is an exothermic process (assuming a rate of diffusion $k_{\text{diff}} = 5 \times 10^7$ in PEG200).^{13,14} These values indicate that for an efficient TET, the later sensitizer is the preferred triplet energy donor for both **C-DPA** and **C₂-DPA**. The S-V quenching experiments are summarized in Figure 2. While the phosphorescence quenching with **DPA** depicts the expected dynamic quenching behavior, the quenching study with **C-DPA** reached a stationary state after certain threshold concentration of the quencher. For example, as shown in Figure 2C, at low/medium concentration of **C-DPA**, the $k_{\text{TET}} = 1.9 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ for **PdTPP**→**C-DPA** in the dynamic regime was three times smaller than that of **PdTPP**→**DPA** suggesting hindered intermolecular interactions/collisions due to the presence of the cholesteryl moieties that may be shielding the **DPA** core. In this picture, we also hypothesized that at micellar concentration of 39 μM , there is a possibility of back **DPA**→**PdTPP** energy transfer via the FRET mechanism.

Figure 3. A cartoon depicting the energy flow and mechanism of TTA-UC. Proposed aggregation features of (A) **C-DPA** with low & medium concentration, (C) **C-DPA** with high concentration, and (D) **C₂-DPA** in PEG200. (B) Proposed TTA-UC of **C-DPA**. The sensitizer represents **PdTPP** or **QDN**.

From the S-V quenching studies, we deduced that the maximum TET and possibly upconversion process could be achieved at $[\text{C-DPA}] = 39 \mu\text{M}$ with 34% quenching efficiency rate (without any aggregation of the acceptor). We attributed the plateau of the S-V curved beyond $[\text{C-DPA}] = 39 \mu\text{M}$ to the aggregation of **C-DPA** units to form self-assembled sub-domains or phase-separation of **C-DPA**, which was visible to the naked eye, indicating that the critical micellar concentration of the cholesteryl-based **DPA** is around $39 \mu\text{M}$. Accordingly, we postulated that the

cholesteryl moieties of **C-DPA** might form a barrier preventing effective interactions of the sensitizer with the core **DPA** of the acceptor (Figures 3A, 3B, and 3C). Results for the S-V quenching studies with **C₂-DPA** are described in the Supporting information (Figure S12); with this acceptor, it was deemed unnecessary to carry out subsequent investigation with **C₂-DPA**, as no TTA-UC was detected using both **QDM** and **PdTPP** (Figures 3D).

Figure 4. Double logarithmic plot of the upconverted emission for (A) **QDN** and **DPA** (B) **PdTPP** and **DPA** (C) **PdTPP** and **C-DPA** as a function of power density of the 532 nm Nd:YAG laser.

Figure 4 shows the double logarithmic plots of the upconverted emission intensity with respect to the 532 nm excitation power density. All three donor-acceptor pairs display the typical low-to-high power density regimes transition: from quadratic (slope = 2) to linear (slope = 1)

intensity dependence, which is a clear indication of the expected TTA process.¹³ The power density threshold (I_{th})²³, where the intensity of the TTA process becomes the dominant deactivation channel for an annihilator triplet state, and the quantum yield of the upconversion process is equal to half of its maximum. From Figure 4, we found that I_{th} are 1790, 133, and 490 mW/cm² for **QDN/DPA**, **PdTPP/DPA**, and **PdTPP/C-DPA**, respectively. We also hypothesize that the intrinsic radiative and non-radiative decays of the sensitizer could be predominant at low power densities. Under this consideration, at power densities above the I_{th} , the TTA process was prevalently approaching a maximum delayed fluorescence driven by the TTA-UC process. Also, the high I_{th} for **QDN/DPA** suggests a less efficient (endothermic) energy transfer¹⁴ as revealed earlier from the S-V quenching results. For the **PdTPP/DPA** pair, the threshold intensity is substantially lower than that of **QDN/DPA**. This observation/result is also in good agreement with the relatively high k_{TET} from the S-V quenching study (Figure 2A and 2B).

Furthermore, we measured the quantum yield of TTA-UC (Φ_{UC}) of the three pairs of donor-acceptor using equation 4 and rhodamine 6G²⁴ as photon counter standard.

$$QY_{UC} = QY_{std} \times \left(\frac{A_{std}}{A_{UC}} \right) \times \left(\frac{I_{UC}}{I_{std}} \right) \times \left(\frac{\eta_{UC}}{\eta_{std}} \right)^2 \quad (4)$$

Where QY represents the quantum yield, A is absorbance (O.D.) at the excitation wavelength.

I denotes the integrated photoluminescence, and η is the refractive index of the solvent.

$_{std}$ stands for the standard (here, rhodamine 6G) sample, and $_{UC}$ denotes the upconversion sample.

We then estimated that the Φ_{UC} for (**QDN/DPA**) is 2% (4% after correcting for inner filter effect), 6% (12%) for (**PdTPP/DPA**), and 1% (2%) for (**PdTPP/C-DPA**).

CONCLUSIONS:

In summary, this study aims at providing additional molecular tools to the upconversion community wherein over-the-counter annihilators, such as **DPA**, have been functionalized with cholesteryl tag(s) to afford **C-DPA** and **C₂-DPA**. In the present work, we successfully employed both **DPA** and **C-DPA** to achieve TTA-UC in biorelevant hydrophilic and viscous polymeric liquid PEG200 media. Moreover, this study offers insight on the behavior of these modified annihilators in terms of donor•••acceptor interactions and the resulting effect on the upconversion process. Using Stern-Volmer quenching experiments, we found that the rate of **PdTPP**→**C-DPA** TET is three times lower than that of **PdTPP**→**DPA**. This reduce k_{TET} value was hypothesized to originate from the shielding effect of the cholesteryl unit. Beyond a certain concentration threshold, **C-DPA** formed a self-assembled sub-domains in the PEG200 medium, which was observed by a plateau in the TET efficiency. This finding was further consolidated by investigating the upconversion efficiency of **C-DPA** upon increasing its concentration. While the cholesteryl units may be beneficial in terms of incorporating dye molecules into lipid bilayers, micelles, and hydrogels and also provide some shielding to molecular oxygen to perform TTA-UC in biological media, we discovered that this substituent may hamper effective donor•••acceptor interactions due to reduced molecular mobility and formation of hydrophobic sub-domains that are inaccessible to the triplet energy donor. This is effect is also manifested by the absence of TTA-UC using **C₂-DPA** as the acceptor. Nevertheless, we successfully achieved TTA-UC with three pairs of donor/acceptor systems **QDN/DPA**, **PdTPP/DPA**, and **PdTPP/C-DPA** with Φ_{UC} values of 2% (4%), 6% (12%), and 1% (2%), respectively. The present study brings new insight into molecular engineering and function of modified biorelevant dyes in order to achieve TTA-UC in biological environments.

ASSOCIATED CONTENT:

Data Availability: The data that support the findings of this study are available from the corresponding author upon reasonable request. In addition, details of the synthetic procedures for all precursor of **C–DPA** and **C₂–DPA**: ¹H and ¹³C NMR spectra, additional UV-vis absorption spectra, emission spectra, this material is available free of charge via the Internet at <http://aip.org>.

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Authors Contribution

These authors have contributed equally to this work

Corresponding Author

* aayitou@iit.edu

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