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Graphical Abstract

Highlights
- VEGFR3 is required for adult hippocampal neurogenesis
- VEGFR3 signaling converts quiescent neural stem cells into progenitor cells
- VEGF-C/VEGFR3 activates human neural stem cells through ERK and AKT

In Brief
The mechanisms regulating neural stem cell activation in vivo are poorly understood. Han et al. show that, in the adult mouse hippocampus, neural stem cells require VEGF-C/VEGFR3 signaling to enter the cell cycle and convert into progenitor cells. The role of VEGFR3 signaling is conserved in human neural stem cells.

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Vascular Endothelial Growth Factor Receptor 3 Controls Neural Stem Cell Activation in Mice and Humans

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SUMMARY

Neural stem cells (NSCs) continuously produce new neurons within the adult mammalian hippocampus. NSCs are typically quiescent but activated to self-renew or differentiate into neural progenitor cells. The molecular mechanisms of NSC activation remain poorly understood. Here, we show that adult hippocampal NSCs express vascular endothelial growth factor receptor (VEGFR) 3 and its ligand VEGF-C, which activates quiescent NSCs to enter the cell cycle and generate progenitor cells. Hippocampal NSC activation and neurogenesis are impaired by conditional deletion of Vegfr3 in NSCs. Functionally, this is associated with compromised NSC activation in response to VEGF-C and physical activity. In NSCs derived from human embryonic stem cells (hESCs), VEGF-C/VEGFR3 mediates intracellular activation of AKT and ERK pathways that control cell fate and proliferation. These findings identify VEGF-C/VEGFR3 signaling as a specific regulator of NSC activation and neurogenesis in mammals.

INTRODUCTION

The adult mammalian brain continuously produces new neurons in two discrete regions, the subventricular zone (SVZ) lining the ventricles and the dentate gyrus (DG) of the hippocampus (Altman and Das, 1965; Doetsch et al., 1999). In rodents, hippocampal neurogenesis is enhanced by external factors, including an enriched environment and voluntary running exercise (Brown et al., 2003; Vivar and van Praag, 2013). A decline in hippocampal neurogenesis occurs with age and may underlie cognitive and mood alterations associated with aging and Alzheimer’s disease (Lazarov et al., 2010; Mu and Gage, 2011).

Hippocampal neurogenesis occurs within the subgranular zone (SGZ) of the DG and is initiated by neural stem cells (NSCs), which undergo a series of divisions to generate new granular layer interneurons that integrate into the hippocampal circuitry (Kempermann et al., 2004). NSCs include a quiescent population, which are radial glia-like cells (RGLs) (or type-1 cells) that are characterized by the expression of Nestin, GFAP, Sox2, and Hes5 (Bonaguidi et al., 2011; Encinas et al., 2011; Lugert et al., 2010; Suh et al., 2007). NSC activation is upon Ascl1 regulation (Andersen et al., 2014) and leads to generate proliferative progenitors, known as intermediate progenitors (IPCs), which in turn give rise to committed neuronal progenitors (neuroblasts). Whereas the steps of hippocampal neuron formation have been well characterized (Bonaguidi et al., 2012; Kempermann et al., 2004), the molecular mechanisms controlling this cellular progression remain poorly understood.

Several signaling pathways are known to maintain hippocampal NSC quiescence through inhibition of cell proliferation. Conditional disruption of the genes encoding BMP2 and 4, sFRP3, Notch/RBP-J, and REST in RGLs all result in rapid activation of NSC division, leading to a transient increase in IPC numbers and production of new adult hippocampal neurons (Ehm et al., 2010; Gao et al., 2011; Jang et al., 2013; Mira et al., 2010). In contrast to these repressors of NSC activation, only a few positive regulators of NSC division and progenitor cell production are known. These include sonic hedgehog/smoothened and BDNF/TrkB, but both of these
signaling pathways are also active in other subpopulations of the hippocampal niche (Li et al., 2008; Machold et al., 2003). Identification of NSC-selective positive regulators should allow prolonging or enhancing neurogenesis during aging and improve the efficacy of NSC-based repair therapies, especially in older patients.

Vascular endothelial growth factors (VEGFs) and their high-affinity tyrosine kinase receptors (VEGFRs) are potent regulators of the growth and maintenance of vascular and neural cells (Eichmann and Thomas, 2013; Zacchigna et al., 2008). In the hippocampus, VEGF-A increases angiogenesis, neurogenesis, and neuronal plasticity (During and Cao, 2006; Fournier and Duman, 2012; Licht and Keshet, 2013). However, it is not clear whether VEGF-A enhances neurogenesis directly, through its receptors VEGFR1 and 2 on neural cells, or indirectly, through factors released from newly formed blood vessels. The related growth factor VEGF-C is a potent regulator of lymphangiogenesis (Loehla et al., 2009). VEGF-C also induces angiogenesis but only weakly, as expression of its receptor VEGFR3 is mainly restricted to tip cells at the extremities of growing blood vessels (Tammela et al., 2008). In the brain, we have previously shown that VEGF-C stimulates neurogenesis via direct cell-autonomous actions of VEGFR3 in neural cells. Deletion of Vegf-c impairs neural development in both Xenopus and mouse embryonic brains, and conditional deletion of Vegf3 within NSCs affects neurogenesis in the adult mouse SVZ (Calvo et al., 2011; Le Bras et al., 2006). We hypothesized that VEGF-C-VEGFR3 signaling might affect hippocampal NSCs in mice and humans, thereby controlling neurogenesis.

Here, we examined the role of VEGFR3 and its mechanism of action in adult hippocampal NSCs. We show that the VEGF-C/VEGFR3 pathway is a positive signal that selectively promotes NSC activation and conversion into progenitor cells in mice. Moreover, VEGFR3 signaling is conserved in human NSCs where VEGF-C activates ERK- and AKT-signaling pathways. Taken together, these data identify VEGF-C/VEGFR3 as a novel signaling pathway in mammalian NSCs that may be targeted therapeutically to improve neurogenesis.

RESULTS

Vegfr3 and Vegf-c Expression in the Hippocampal DG

We characterized the expression of Vegfr3 in the adult hippocampus using Vegfr3::YFP BAC-transgenic mice (Calvo et al., 2011). YFP labeled capillaries and neural cells localized along the SGZ (Figure 1A). Flow cytometric analysis of dissociated DG cells confirmed that 16% of neural cells (CD31+/non-endothelial cells) were YFP-expressing cells, hereafter referred to as Vegfr3YFP cells (Figures 1B and S1A). Quantitative RT-PCR (qRT-PCR) showed that Vegfr3 transcripts are highly enriched in Vegfr3YFP cells compared to DG neural cells, whereas lower levels of the other two VEGFR family members Vegfr1 and Vegfr2 were found in Vegfr2YFP cells (Figures 1C and S1B). qRT-PCR also showed enrichment of ligand VEGF-C transcripts in Vegfr3YFP cells, suggesting possible autocrine VEGF-C/VEGFR3 signaling in these cells (Figure S1B). In addition, prominent VEGF-C expression was seen in DG cells surrounding the SGZ by X-gal staining of Vegfc1acZ+ knockin mice and antibody staining (Figures 1D and 1E), indicative of possible paracrine VEGF-C signaling to Vegfr3YFP cells.

To examine whether this ligand-receptor system functioned in hippocampal neurogenesis, we determined which type of neural cells expressed VEGFR3. About half of Vegfr3YFP cells (55%) exhibited a RGL morphology characteristic of NSCs (type-1a) and stained positive for GFAP, Nestin, or BLBP radial glia markers (Figures 1F–1H and S1C). Almost all Vegfr3YFP RGL cells expressed GFAP (99%), including a 17% subpopulation that lacked Nestin expression. Previous reports showed that GFAP+/Nestin– RGL cells are quiescent NSCs and suggested that GFAP+/Nestin– RGL cells are activated NSCs (De Carolis et al., 2013), which would indicate that Vegfr3YFP RGL cells include both quiescent and activated NSCs. The other 45% of Vegfr3YFP cells lacked radial processes (Figure 1G). Less than 10% of those cells exhibited weaker YFP and stained for the IPC markers Ascl1 and Tbr2, and the rest included S100b+ astrocytes (15%) and non-radial Tbr2– cells that may correspond to non-radial NSCs (20%). DCX+ neuroblasts as well as postmitotic interneurons were excluded from the Vegfr3YFP population (Figures 1F, 1H, and S1C). Flow cytometric analysis of Vegfr3YFP cells confirmed that they include a majority of S100b+/Glast+/GFAP+ astroglial cells (65%–70%) and Sox2+ (50%) cells (Figure S1D; Table S1). RNA-sequencing analysis (RNA-seq) also showed enrichment of transcripts encoding NSPC markers in Vegfr3YFP cells, and specific expression of GFAP, Nestin, Blbp, Ascl1, Tbr2, and Sox2 in Vegfr3YFP cells was confirmed by qRT-PCR and flow cytometric analysis (Figures S1E and S2A; Table S2).

Analysis of proliferation showed that Vegfr3YFP RGL cells are mainly quiescent (Ki67+) whereas non-radial Vegfr3YFP cells include a few dividing cells (5% Ki67+; Figure 1I). Following a short BrdU-pulse (3 hr) to label proliferating NSCs and IPCs, 60% of BrdU+ cells were Vegfr3YFP cells (Figures 1J and 1K). Altogether, these data show that Vegfr3 expression characterizes NSCs and declines in neural progenitor cells in the adult hippocampal SGZ (Figure 1L).

VEGF-C/VEGFR3 Signaling Activates Hippocampal NSCs In Vitro

To test the stem cell properties of Vegfr3YFP cells, we sorted Vegfr3YFP and Vegfr3YFP-negative DG cells and analyzed their ability to form neurospheres, self-renew, and differentiate into neural cell types in vitro (Figure 2). We found that neurospheres formed exclusively from Vegfr3YFP cells, but not from the fraction of Vegfr3YFP-negative cells, which establishes that Vegfr3YFP cells are NSCs (Figure 2A). Neurospheres derived from Vegfr3YFP cells were able to self-renew for at least six passages, confirming the presence of NSCs with long-term self-renewal capacity (Figure 2B). Vegfr3YFP neurosphere cells transferred onto Matrigel differentiated into TuJ1+ neurons (13%) and GFAP+ astrocytes (71%) but rarely into O4+ oligodendrocytes (0.9%), which is consistent with previous reports showing that DG NSCs generate only very few oligodendroglial cells (Bona- guidi et al., 2011; Figure 2C). Vegfr3YFP cells are therefore NSCs, which maintain self-renewal and multilineage differentiation capacities.
Addition of recombinant VEGF-C (50 ng/ml) had no effect on cell survival (Figure 2D). However, VEGF-C-stimulated Vegfr3<sup>YFP</sup> cells generated more neurospheres (Figure 2E). The activation of Vegfr3<sup>YFP</sup> cells by VEGF-C was blocked by treatment with a blocking antibody against VEGFR3 (31C1), indicating that the response to VEGF-C is VEGFR3 dependent (Figure 2E).

To get a molecular readout of VEGF-C action on hippocampal NSCs, we performed RNA-seq. As shown in Figure S2B and Table S2B, VEGF-C-treated Vegfr3<sup>YFP</sup> cells, but not Vegfr3<sup>YFP-negative</sup> cells, strongly upregulated mRNA expression of regulators of the G1-S transition, such as cyclin D/E and cyclin-dependent kinases Cdk1/2, and of factors regulating chromosomal DNA replication. Furthermore, VEGF-C stimulation appears to drive NSCs from their quiescent state toward a proliferating progenitor state.

In VEGF-C-treated cells, the expression of NSC markers such as Notch1/2 (Ables et al., 2010), Hes5 (Imayoshi et al., 2013), Ctnnb1 (Lie et al., 2005), Bmpr1 (bone morphogenetic protein receptor) (Mira et al., 2010), Mhi1/2 (Mushashi1/2) (Okano et al., 2005), Bmi-1 (Fasano et al., 2009), and Rest (Abrajano et al., 2010) are downregulated whereas the expression of progenitor cell markers Ascl1 (Kim et al., 2008; Lugert et al., 2012), Sox21 (Matsuda et al., 2012), and Numb (Aguirre et al., 2010; Petersen et al., 2002) are upregulated (Figure S2B; Table S2B).

To confirm the effect of VEGF-C on cell cycle progression, we analyzed the cell cycle activity of Vegfr3<sup>YFP</sup> cells. After overnight culture in the presence or absence of VEGF-C, cells were stained with Pyronin Y/ Hoechst 33342 and analyzed by flow cytometry (Figure 2F). VEGF-C treatment of Vegfr3<sup>YFP</sup> cells, but not of Vegfr3<sup>YFP-negative</sup> cells, promotes a significant increase of cells in G1 phase (Figure 2F). Taken together, these data demonstrate that VEGF-C/VGFR3 signaling in NSCs initiates cell cycle entry and conversion of quiescent cells into proliferating progenitor cells.

**Maintenance of Hippocampal Neurogenesis In Vivo Requires Vegfr3**

We next investigated consequences of inducible Vegfr3 deletion in hippocampal NSCs in vivo, using Glast<sup>CreERT2</sup> to delete Vegfr3<sup>Roxi/fox</sup> allele (Glast<i>iJR3</i>) (Figures 3 and S3). The Glast<sup>CreERT2</sup> driver line allows the targeting of Glast<sup+i</sup> NSCs that express Vegfr3. Restricted expression of VEGF in glial cells excluded non-radial progenitor cells in the DG (Figure S3A; DeCarolis et al., 2013; Mori et al., 2006). Mice were sacrificed 1 or 5 months after Tx treatment to test the effects of Vegfr3 deletion in neurogenesis. Interccors of Glast<sup>CreERT2</sup> mice with ROSA26 and mT/mG reporter lines led to genetic recombination in >65% of SGZ cells, indicating that Glast<sup>CreERT2</sup> likely drove mosaic deletion of Vegfr3 in hippocampal NSCs (Figures S3A and S3B). Genetic recombination of Vegfr3 locus was assessed by PCR in Tx-treated Glast<i>iJR3</i> mice (Figure S3C). Glast<i>iJR3</i> and control (Vegfr3<sup>Roxi/fox</sup>) animals were given a 3 hr BrdU pulse prior to sacrifice to label activated NSCs (Figure 3A). One-month Tx-Glast<i>iJR3</i> and control mice showed similar numbers of Brdu<sup+</sup> SGZ cells and DCX<sup+</sup> neuroblasts (Figures 3B–3E). The number of GFAP<sup+</sup>Nestin<sup+</sup> RGL cells and of astrocytes in the hilus was similar between controls and Glast<i>iJR3</i> mice (Figures 3F and 3G). The number of RGL cells was also not altered in Glast<i>iJR3</i>; Vegfr3<i>YFP</i> mice expressing YFP in RGL cells (Figure S3D). Altogether, these data indicate that decline of hippocampal neurogenesis in Glast<i>iJR3</i> is not caused by a primary loss of NSCs or by an abnormal neuron/astrocyte differentiation balance. Anti-active caspase 3 antibody labeling and TUNEL staining of the DG showed only rare cells engaged in programmed cell death (PCD) in both Tx-control and -Glast<i>iJR3</i> mice, indicating that Vegfr3 deletion did not induce PCD in DG cells (data not shown). Therefore, Vegfr3 loss of function in NSCs does not compromise NSC survival and renewal but selectively impairs NSC activation and conversion into proliferative IPCs.

Decline of hippocampal neurogenesis is associated with cognitive impairments and an enhancement of fear and anxiety in both mice and humans (Encinas and Sierra, 2012; van...
Figure 2. VEGF-C/VEGFR3 Signaling Activates Hippocampal NSCs In Vitro

(A and B) Neurosphere cultures derived from sorted Vegfr3YFP and Vegfr3YFP-negative cells. The formation of neurospheres was only observed in Vegfr3YFP cell cultures that can self-renew for at least six successive passages.

(C) Representative images and quantification of neurosphere differentiation. Vegfr3YFP cells differentiate into TuJ1+ neuron (green), GFAP+ astrocyte (blue), and very few O4+ oligodendrocyte (red).

(D) Cell death was quantified by TUNEL staining.

(E) Representative images and quantification of neurospheres after treatment with VEGF-C (50 ng/ml) and a VEGFR3-function-blocking antibody (31C1).

(F) FACS profile and cell cycle analysis of control Vegfr3YFP cells and VEGF-C-treated Vegfr3YFP cells after Pyronin Y/Hoechst 33342 staining.

The scale bars represent 50 μm (A and C).

Student’s t test: p < 0.05 (*); p < 0.005 (**); not significant (ns). Bars: mean ± SEM; n = 3–5 independent experiments. See also Figure S2.
Vingaarden and Franklin, 2013). We reasoned that decreased neurogenesis in 5-month Tx-Glast iΔR3 mice might lead to behavioral defects. Five-month Tx-Glast iΔR3 and -age-matched control mice were subjected to two paradigms to assess anxiety and locomotor activity: the elevated plus maze (EPM) test (Pellow et al., 1985) and the open-field (OF) test (Gould et al., 2009), respectively (Figures 3H, 3I, and S3E–S3K). Five-month Tx-Glast iΔR3 mice displayed an anxiety phenotype in the EPM task, which was not due to a change in locomotor activity (Figures 3H, 3I, and S3E–S3K). Altogether, these results suggest a VEGFR3-dependent correlation between the decline of hippocampal neurogenesis and an increase of fear/anxiety status that manifests in middle-aged mice.

**VEGF-C/VEGFR3 Signaling Activates Hippocampal NSCs In Vivo**

Because VEGF-C treatment drives NSC cell cycle entry in vitro, we investigated whether VEGF-C/VEGFR3 signaling also promoted NSC activation in vivo. VEGF-C was overexpressed during 2 to 3 weeks within the hippocampus of C57BL/6 adult mice by stereotaxic injection of adeno-associated virus (AAV) encoding full-length VEGF-C (AAV-VEGF-C) above the DG (Figure 4A). AAV-EGFP was used as a control, and GFP expression in the hippocampal region showed that all animals had been efficiently infected (Figure S4A). VEGF-C induced a 2-fold increase in the number of BrdU+ SGZ cells compared to control (Figure 5A). Running Vegfr3::YFP mice showed a robust increase in the number of BrdU+ Vegfr3::YFP cells compared to sedentary controls (Figure 5B). The total population of Vegfr3::YFP cells was not increased (Figure S5A), but BrdU+ Vegfr3::YFP cells significantly increased (Figures 5B and S5B). Therefore, running activity promotes activation of Vegfr3::YFP cells. This effect occurs in the absence of significant vascular remodeling (Figures S5C–S5E). As shown in Figure S5F, running activity had no significant effect on the level of Vegfc expression but increased Vegfr3 transcript expression in the DG. Vegfr3 expression likely amplifies in Vegfr3::YFP cells because the number of these cells remained stable. Altogether, these data indicate that the running-activity-induced hippocampal neurogenesis involves the activation of Vegfr3::YFP NSCs and the upregulation of Vegfr3 expression in these cells.

To determine whether VEGFR3 was required for the exercise-induced activation of Vegfr3::YFP NSCs, Glast iΔR3 and control mice were housed with a running wheel for 3 weeks after Tx treatment (Figure 5C). Glast iΔR3 mice displayed a normal running activity with similar running distance compared to control mice (Figure S5G). However, Glast iΔR3 mice were unable to activate running-induced neurogenesis, and the number of their BrdU+ SGZ cells was comparable with controls (Figures 5D and 5F). In contrast, running still increased the number of DCX+ neuroblasts in Glast iΔR3 mice similar to controls (Figures 5E and 5G). A possible explanation for the lack of effect on the DCX+ cells is that the loss of NSCs in Tx-Glast iΔR3 mice might be compensated for by the increased proliferation of early neuroblasts (Tbr2+DCX+), which is known to be induced by running activity (Hodge et al., 2008). Therefore, hippocampal NSC activation in response to running activity is Vegfr3 dependent but does not affect short-term production of newborn neurons.

**Vegfr3 Activates ERK- and AKT-Signaling Pathways in Human NSCs**

To determine whether Vegfr3 signaling is conserved in human NSCs and to identify downstream targets of Vegfr3, we analyzed Vegfr3 expression and signaling pathways in cultures of NSCs derived from human embryonic stem cells (hESCs) (H1 and H9 cell lines). hESCs were differentiated into free-floating NSC-containing spherical neural masses (SNMs) (Figures S6A–S6C). Western blot and immunostaining analyses demonstrate that SNMs derived from either H9 or H1 hESCs express Vegfr3, but not Vegfr2 (Figures 6A, 6B, and S6D). Vegfr3+ cells display elongated cytoplasmic processes that correspond mainly to BLBP-positive NSCs, but not DCX neuroblasts, under both 3D- and flat-culture conditions (Figures 6C and S6E).

hNSC treatment with VEGF-C stimulates cell division (Figure 6D). The mitotic response to VEGF-C is preceded by a rapid induction of Vegfr3 phosphorylation (Figure 6E). VEGF-C also strongly activates intracellular signals including ERK and AKT phosphorylation (Figure 6F). At a later time point (15 min), VEGF-C treatment stimulates AKT-downstream-signaling pathways including glycogen synthase kinase 3β (GSK3β) (Figure 6F). GSK3β is inhibited by pAKT and regulates cell cycle entry, cell proliferation, and cell fate (Hur and Zhou, 2010). MAZ51, an indoleamine that blocks VEGF-3 signaling (Kirkin et al., 2004), inhibits...
Figure 3. Vegfr3 Deletion in Adult Hippocampal NSPCs

(A) Schedule of Tx and BrdU administration to induce Vegfr3 deletion in NSCs and label-activated NSCs. (B and C) Representative images of coronal DG sections from control (upper panel) and Glast ΔR3 mice (lower panel) at indicated time points. Sections are stained for BrdU (red), DAPI (blue), and DCX (green). The scale bars represent 70 μm.

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VEGF-C-induced AKT phosphorylation in hESC-derived NSCs (Figure S6F). Therefore, VEGF-C/VEGFR3 regulates downstream signals controlling NSC activation, providing a molecular explanation for the NSC behavioral changes observed following VEGF-C/VEGFR3 signaling manipulation.

DISCUSSION

We here identify a growth-factor-receptor-signaling pathway that promotes NSC activation during hippocampal neurogenesis. We show that Vegfr3 is expressed by mouse hippocampal NSCs and that VEGF-C/VEGFR3 signaling is necessary for activation and conversion of NSCs into progenitor cells. Furthermore, we demonstrated that VEGF-C/VEGFR3 signaling is evolutionarily conserved in human NSCs, where it promotes cell proliferation and activates ERK/akt pathways. To our knowledge, VEGF-C/VEGFR3 is the first known growth-factor-signaling pathway that selectively promotes NSC activation, suggesting that VEGF-C could be therapeutically used to enhance neurogenesis in humans.

We found that, like type-B astrocytes of the SVZ (Calvo et al., 2011), hippocampal NSCs exhibit Vegfr3YFP reporter expression and a specific enrichment of Vegfr3 transcripts. Expression of YFP in all RGL cells and the exclusive generation of neurospheres from Vegfr3YFP DG cells strongly suggest that Vegfr3 is characteristic for hippocampal NSCs. The Vegfr3::YFP mouse adds to the few models of transgenic reporter lines, such as Nestin- (Dranovsky et al., 2011; Encinas et al., 2006) and Hes5-reporter mice (Lugert et al., 2010), allowing the detection and sorting of adult NSCs. The level of YFP reporter expression was much higher in NSCs than progenitors, indicating that Vegfr3 is downregulated along with the asymmetric cell division of activated NSCs into progenitor cells.

Three independent lines of evidence demonstrate a direct requirement of VEGFR3 in NSC activation. First, VEGF-C treatment of Vegfr3YFP cells, but not Vegfr3YFP-negative cells, activates cell cycle entry; second, genetic deletion of Vegfr3 in NSCs impairs neurogenesis; and third, VEGF-C-induced NSC activation is lost in Vegfr3 mutant mice. Vegfr3 deletion in NSCs decreases the populations of activated NSCs and neuroblasts, without affecting the pool of NSCs in middle-aged mice. Vegfr3 activity is therefore not necessary for NSC maintenance but required for the generation of progenitors and neuroblasts, suggesting that VEGF-C/VEGFR3 signaling regulates activation of cell division in NSCs. This hypothesis is reinforced by RNA-seq data showing that VEGF-C stimulation shifts NSCs from their undifferentiated state toward a proliferating progenitor state. VEGF-C/VEGFR3 signaling in NSCs downregulates the expression of Bmi-1 and Rest, which interact with a broad array of transcriptional and epigenetic regulatory cofactors to maintain stemness and inhibit NSC differentiation (Abrajano et al., 2010; Fasano et al., 2009). This suggests that activation of VEGFR3 by VEGF-C de-represses the NSC differentiation program. VEGF-C/VEGFR3 signaling in NSCs is also characterized by upregulated expression of factors that are mandatory checkpoint regulators of the cell cycle. Although novel in the context of NSCs, this finding is not surprising when viewed from an angiogenesis perspective. VEGF-C is a well-known mitogen for VEGFR3+ lymphatic endothelial cells and mediates downstream signaling through activation of ERK and AKT (Koch and Claesson-Welsh, 2012), which suggests that these downstream signaling events are conserved between endothelial cells and NSCs.

The sources of VEGF-C for inducing NSC activation as well as the mechanisms regulating Vegfr3 expression in NSCs remain to be determined. NSCs contain Vegf-c transcripts, suggesting that, besides paracrine VEGF-C from DG cells, autocrine VEGF-C may activate VEGFR3 at the cell membrane. The continuous process of NSC activation also implies that VEGF-C availability may be regulated by translational or post-translational processes in NSC, perhaps through control of VEGF-C maturation by enzymes such as CCBE1 (collagen- and calcium-binding epidermal growth factor domains 1) (Le Guen et al., 2014). NSC activation is moreover likely to be controlled by a tight regulation of VEGFR3 expression levels. We found that Vegfr3 expression in DG cells is physical activity dependent, suggesting that it could be regulated by activity-dependent epigenetic mechanisms, such as DNA methylation that modulates neurogenesis (Guo et al., 2014; Jobe et al., 2012; Saharan et al., 2013) and also controls Vegfr3 expression in endothelial cells (Quentmeier et al., 2012). Notch signaling is another regulator that inhibits VEGFR3 expression in endothelial cells (Benedito et al., 2012), suggesting that Notch may maintain NSCs quiescence by regulating Vegfr3 expression. Conversely, we show that VEGFR3 activation in NSCs downregulates Notch and its downstream targets, suggesting that functional antagonism between VEGFR3 and Notch could switch NSC fate from quiescence to activation. Interestingly, Notch1 deletion in NSCs (Ables et al., 2010) induces similar phenotype to Vegfr3 deletion in NSCs, i.e., a delayed reduction in the populations of dividing NSPCs and neuroblasts.

Another novelty of our study is the finding that VEGFR3 expression is conserved in human NSCs, where it promotes proliferation and regulates ERK and AKT, GSK3β signaling cascades in response to VEGF-C. Through VEGFR3, VEGF-C activates several downstream signaling cascades promoting transcription, such as ERK for mitogen-stimulated proliferation (Phoenix and Temple, 2010), and translation, such as mTOR and S6K signals, which regulate cell survival and cell fate decisions (Bordey, 2014; Hur and Zhou, 2010). VEGF-C is therefore able to rapidly and simultaneously regulate cell cycle entry and differentiation processes in NSCs, which likely explains its potent role in mouse hippocampal neurogenesis.
Figure 4. VEGFR3 Mediates VEGF-C-Induced Proliferation in Hippocampal NSCs

(A) Intra-hippocampal injection of AAV9-GFP (control) and -VEGF-C. At 2 to 3 weeks after AAV infection, mice were given a 3-hr pulse of BrdU and analyzed to quantify proliferation and neuroblast production in the DG.

(B and C) Quantification of BrdU+ cells and DCX+ cells in the SGZ of control and VEGF-C-treated mice.

(D and E) Representative images and quantification of BrdU+ Vegr3YFP+ cells after VEGF-C overexpression in Vegfr3::YFP mice. VEGF-C treatment increases BrdU+ Vegr3YFP+ cells in SGZ (white arrows). The scale bars represent 35 μm.

(F and G) Representative images and quantification of BrdU+ Vegr3YFP RGL cells after VEGF-C overexpression in Vegfr3::YFP mice. VEGF-C treatment increases BrdU+ Vegr3YFP RGL cells in SGZ (white arrows). The scale bars represent 35 μm.

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We found that, in addition to impaired neurogenesis, Tx-Glast iΔR3 showed increased anxiety behavior, suggesting that VEGFR3 signaling in DG NSCs may contribute to emotional response. Although further research is required to reinforce this finding, it confirms previous genetic evidence that manipulation of neurogenesis alters behavior in models of anxiety and depression (Eisch and Petrik, 2012; Gross et al., 2002; Saxe et al., 2006; Snyder et al., 2011). However, VEGF3R is expressed by other Glast-expressing cells besides hippocampal NSCs, including tannocytes in the hypothalamic median eminence and a few astrocytes in the amygdala (Hou et al., 2011; Robins et al., 2013). These regions are involved in the regulation of the hypothalamic-pituitary-adrenal (HPA) axis that controls reactions to stress as well as mood and emotions (Canteras et al., 2010), suggesting that VEGF3R signaling in NSC/astrocytes may act on behavior by modulating the functioning of the limbic-HPA axis.

VEGF-C can promote angiogenesis (Gaál et al., 2013) but only weakly when compared to VEGF-A, because VEGF3R-signaling activity is mainly restricted to sprouting tip cells and inhibited in other endothelial cells by Notch signaling (Tammela et al., 2011). VEGF-C may thus be a candidate factor to enhance NSC activation, without unwanted effects on other cell types. Altogether, our findings are consistent with the possibility that VEGF-C activation of NSCs could improve age-related decline in hippocampal neurogenesis and associated mood defects, such as those seen in Alzheimer’s disease.

**EXPERIMENTAL PROCEDURES**

**Animals**

C57/B16 mice (Jackson Laboratories) were maintained in the Animal Research Center at Yale University. The Vegfr3::YFP (Calvo et al., 2011), GlastiΔR3/+, Vegfr3i−/−/+ (Haiko et al., 2008), and Vegf-c−/−/− (Karkkainen et al., 2004) mice have been described previously. GlastiΔR3/ mice were crossed with Vegfr3i−/−/+ mice to generate NSC-specific and Tx-inducible Vegfr3 mutant mice (Glast iΔR3). Animals were sacrificed the days indicated in each experiment. All experiments were approved by the IACUC of Yale University.

**TX Treatment**

To induce genetic recombination, 2-month-old adult mice were injected intraperitoneally (i.p.) with tamoxifen (TX) once per day for 10 consecutive days (as indicated in the experiments) at a dose of 2 mg per day.

**BrdU Administration**

BrdU (Sigma) was administered to the adult animals via a single i.p. injection (300 mg/kg) 3 hr before sacrifice. The brains were processed for immunohistochemistry as described below.

**Tissue Collection and Immunohistochemistry**

Animals were anesthetized with isoflurane, and skull holes were drilled overlying the hippocampus on both hemispheres. The site of infusion was at the following coordinates relative to bregma: anteroposterior, −1.8 mm; mediolateral, −1.8 mm; and dorsoventral, −1.8 mm. Two microliters of control AAV-Egf (control) or AAV-VEGF-C were infused at a flow rate of 0.5 μl/min. Two to three weeks after injection, the brains were processed for immunohistochemistry as described below.

**Stereological Analysis**

The marker (BrdU, DCX, and YFP)-positive cells were counted in a one-in-six series of five sections (40 μm, 240 μm apart) throughout the rostrocaudal extent of the dorsal DG. The sections stained with DAPI were used to measure DG area using Velocity software (Perkin-Elmer). The total number of marker+ cells was determined by summing the number of marker+ cells
and multiplying this number of cells by the number of serial sections (number of cells/series x 6). Total volume was determined by summing the traced DG areas for each section multiplied by the distance between sections sampled (area x 240 μm). To calculate the number of marker+ cells per volume (1 mm³), the total number of marker+ cells was divided by the total volume.

**hESC-Derived NSC Cultures**

We have progressively differentiated hESCs into free-floating SNMs that contain self-renewing, multipotent NSCs according to a protocol (Cho et al., 2008). All experiments have been performed on SNMs derived from two different H1 (male) and H9 (female) hESC lines (Allegrucci and Young, 2007). Detailed information is provided in Supplemental Experimental Procedures.

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**Figure 5. VEGFR3 Is Required for Exercise-Induced NSPC Proliferation**

(A) Schedule of voluntary running activity and BrdU administration for Vegfr3::YFP mice.

(B) Quantification of BrdU+ cells and BrdU+Vegfr3YFP cells in the SGZ of sedentary control and running Vegfr3::YFP mice.

(C) Schedule of Tx treatment, voluntary running activity, and BrdU administration in control and GlastΔR3 mice.

(D–G) Representative images and quantification of BrdU+ SGZ cells and DCX+ cells in control (upper panel) and GlastΔR3 mice (lower panel), either sedentary (left) or running (right). BrdU+ cells (red), DAPI (blue), and DCX+ cells (green) are shown. Running induces proliferation of NSCs in control mice, but not in GlastΔR3 mice. A similar number of neuroblasts is generated in control and GlastΔR3 mice. The scale bar represents 35 μm.

Student’s t test: p < 0.05 (*); p < 0.001 (***); not significant (ns). Bars: mean ± SEM.

See also Figure S5.
A two-tailed, unpaired Student's t test or Mann-Whitney test were done to determine statistical significance (Graph-Pad Prism 6). Differences were considered statistically significant if the p value was 0.05 (*p < 0.05; **p < 0.01; ***p < 0.005). Error estimates were displayed as SEM.

According to the MIAME guidelines, all transcriptomic data were submitted to a MIAME compliant repository, NCBI’s Gene Expression Omnibus (GEO), and are accessible through the GEO accession number GSE55621.

**Figure 6. VEGFR3-Signaling Pathway in Human NSCs**

(A) Western blot analysis of VEGFR2 and VEGFR3 expression in H1 human-ESC-derived NSCs. VEGFR3 migrates as three molecular weight species: 195, 175, and 125 kDa (arrows). EB, embryoid bodies; ESC, embryonic stem cells; HUVEC, human umbilical vein endothelial cells; NP, neural progenitors; SNM, spherical neural mass.

(B) SNMs stained with anti-hVEGFR2 and -hVEGFR3 antibodies as indicated. The scale bars represent 100 μm.

(C) SNMs stained for VEGFR3 (green), BLBP (red; astroglia/NSC), or DCX (red; neuron) and DAPI (gray). The scale bars represent 20 μm.

(D) Representative image of phospho-histone H3 staining. The scale bar represents 100 μm.

(E) Phospho-VEGFR3 analysis by immunoprecipitation.

(F) Western blot analysis of the phosphorylation state of ERK, AKT (tyrosine 398 and serine 473), and GSK3β in SNMs after VEGF-C stimulation. Student’s t test: p < 0.05 (*). Bars/lines: mean ± SEM.

See also Figure S6.
SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, six figures, and two tables and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2015.01.049.

AUTHOR CONTRIBUTIONS

J.H. contributed to the in vivo studies and manuscript; C.-F.C. and M.L. contributed to the in vitro studies; K.L.B. contributed to the in vivo studies; T.H.K. contributed to the RNA-seq analysis and hNSC studies; J.-H.P. and C.P. contributed to the immunostaining; H.N. and K.L.B. contributed the AAVs and mice; R.S.D. contributed methodological advice and practical support for behavior studies; A.C.E. contributed to the research design and manuscript; and J.-L.T. contributed to the research design and manuscript.

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