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An autochthonous case of cystic echinococcosis in Finland, 2015

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We report a case of pulmonary cystic echinococcosis in a child from eastern Finland with no history of travelling abroad. The cyst was surgically removed and the organism molecularly identified as *Echinococcus canadensis* genotype G10. This parasite is maintained in eastern Finland in a sylvatic life cycle involving wolves and moose; in the present case, the infection was presumably transmitted by hunting dogs.

In Fennoscandia (Finland, Norway and Sweden) and parts of western central Europe, cystic echinococcosis (CE) or hydatidosis is a rare disease seen in immigrants or other people who have resided in endemic countries. Here we present an unexpected autochthonous case of pulmonary CE in a Finnish child.

**Case description**

At the end of February 2015, an eight-year-old previously healthy child from eastern Finland had sudden abdominal pain and developed a vigorous generalised urticarial rash without other abnormal findings. The child had never travelled abroad. A week later, the child was admitted to hospital because of fever (38.5 °C) and a persistent cough. Upon admission, the patient was pale and their breath sounds were decreased over the left side of the chest. A chest X-ray revealed a large cavity, partially filled with fluid (Figure 1A). The ultrasound showed a considerable avocado-sized hollow (13.5 x 9 cm) with multiple lobulation. Consistent with these findings and an elevated C-reactive protein (58 mg/L; norm: 0-5 mg/L), and high normal leukocyte count (12.9 x 10^9/L; norm: 4.5-13.5 x 10^9/L), a lung abscess with parapneumonic empyema was set as the principal diagnosis. Elevation of the serum eosinophil leucocytes (4.7 x 10^9/L; norm: 0.1–0.4 x 10^9/L) was also observed.

Despite empirical treatment with intravenous cefuroxime (100 mg/kg/day divided in three doses) and clindamycin (40 mg/kg/day divided in three doses), the fever and cough persisted and the radiological findings did not resolve within a week. Computed tomography (CT) of the thorax on day 5 after admission revealed a pleural effusion and an empyema in the lower lobe of the left lung, and an abscess suspicion in the upper lobe (Figure 1B). A left-sided thoracotomy was performed on day 7 after admission. An empyema was detected and debridement of the pleural cavity and decortication was carried out. The suspected abscess cavity was opened and partially resected. Clinically, it proved to be a cystic structure with connection to small bronchi. This bronchocystic fistula was closed with sutures.

On the first postoperative day, the direct microscopic examination of calcofluor white-stained fragments of cyst wall and cyst content showed plenty of hooks and protoscolices, typical of *Echinococcus*. Hooks were detected also in Gram and Ziehl-Neelsen stained samples. After this finding, albendazole treatment (10–15 mg/kg/day divided in two doses) was started. Because of the obvious CE, a re-thoracotomy was performed on the second postoperative day. The cyst was close to the hilar structures and thus the upper lobe of the left lung had to be removed.
PCR for *Echinococcus* (partial mitochondrial cytochrome *c* subunit I gene [1]) was positive in specimens taken during the first thoracotomy from both the cyst and pleural empyema, indicating either spontaneous or intraoperative spillage of the cyst content into the pleural cavity. Sequencing revealed 100% identity with *Echinococcus canadensis* genotype G10 previously isolated in cervids from Finland [2].

Postoperatively, the lung function recovered slowly. A thorax and abdomen CT and an abdominal ultrasound did not reveal signs of hydatid cysts in the liver or other parts of the body. The findings in brain magnetic resonance imaging (MRI) were also normal. A serum sample taken six weeks after the first operation was strongly positive for IgG antibodies against *E. granulosus* by ELISA and IHA (Swiss Tropical and Public Health Institute, Basel, Switzerland). The tests are based on material of *E. granulosus* sensu stricto (s.s.) genotype G1 of sheep originating in Sardinia (Bruno Gottstein, University of Bern). Albendazole treatment was continued postoperatively for a total of three months.

**Investigation of close family members and dogs owned by the family**

The patient’s parents and two siblings were examined for *Echinococcus* infection. Chest X-rays and abdominal ultrasound results proved normal and serological tests were negative. The family lives in the countryside and has three dogs, which are used mostly for fowling. There was no history of feeding dogs with raw cervid viscera. The dogs were regularly dewormed, but not with anthelmintics effective against *Echinococcus*. Before treatment with praziquantel, faecal samples were collected from the dogs. Mitochondrial DNA was extracted directly from the faecal material [3] because the small size of the samples did not allow isolation of parasite eggs. A fragment of mitochondrial ribosomal DNA of the *E. granulosus* sensu lato (s.l.) complex [4] was detected by PCR in all the samples, but the short unspecific sequence did not reveal the exact species. Control specimens taken one month after deworming were PCR-negative.

**Background**

The most important aetiological agents of human CE are *E. granulosus* s.s. and *E. canadensis*, both of which were formerly included in the species complex of *E. granulosus* s.l [5]. Canids are the definitive hosts of these parasites, and various ungulates serve as intermediate hosts [6].

CE is found worldwide, prevailing in many endemic areas, typically in pastoral communities where people have close contact with dogs [6]. In the northern Fennoscandia, endemic human CE occurred in the reindeer herding area until the second half of the last century [7]. In Finland, the last published autochthonous case was diagnosed in 1963, in Sweden in 1967, and in Norway in 1977 [8]. Human infections derived from a synanthropic cycle involving dogs and reindeer, which broke up due to changes in traditional reindeer husbandry including decline of herding dogs [7,8]. A sylvatic wolf–moose cycle still exists in Finland, but it has not been linked to human infections [7].

**Discussion**

Since our patient had never travelled abroad, a parasite infection was not initially considered in the differential diagnostics. In endemic areas with high prevalence of
clinical cases (e.g. in the Middle East or parts of Africa) the eosinophilia, generalised urticaria and pulmonary involvement would be expected to raise a suspicion of CE. The incidental finding of Echinococcus hooks in smears obtained for both mycological and bacterial staining highlights the importance of direct microscopic examination in this case.

The growth rate of hydatid cysts is slow (max. 13 cm per year, but usually much less) [9,10], depending on location, causative species and probably also patient’s age [9,11,12]. In the present case, the cyst had presumably been growing for at least one year and eventually constricted the lower lobe, thus predisposing to a secondary pulmonary infection. During this process, a fistula had ruptured and the cyst had been partially drained into the bronchial tree causing a persistent cough. Cyst leakage and exposure to echinococcal antigens probably account for the abdominal pain and urticarial rash.

Based on ultrasound, CT and clinical findings, the present case can be classified as CE4l according to the classification of the World Health Organization Informal Working Group on Echinococcosis (WHO-IWGE) [9,13]. In the classification, ‘4’ refers to an inactive stage (with degenerative contents and calcification in the present case) and ‘l’ to a large size [13]. Most cysts of this type are not fertile [13], but in this case protoscolices were present, indicating fertility.

Surgery still remains the main therapeutic option in pulmonary CE [6]. Here the cyst was not fully removed in the initial operation, since the parasitic aetiology was recognised only after microscopic examination. Due to the large size and difficult location of the cyst, the patient was re-operated and a lobectomy was performed for complete eradication of the parasite, and to avoid relapses and bacterial infections. An optimal length of postoperative albendazole treatment has not been established [9]. However, considering the formation of the fistula and the spilling during the initial operation, a prolonged course of albendazole was considered warranted. Furthermore, follow-up visits including serology are planned every three to six months over the first year and, thereafter, once a year by serology and MRI at least for five years if no clinical symptoms or findings develop.

According to reports from Alaska and Canada, ‘sylvatic’ (transmitted in wildlife) CE attributed to E. canadensis is more benign than classic ‘pastoral’ CE predominantly caused by E. granulosus s.s. in sheep-raising countries [11,14-16]. Relative mildness was also typical of endemic CE which occurred in the reindeer herding area of Fennoscandia [8]. The sylvatic form of CE is characterised by pulmonary involvement of relatively small hydatids with thinner laminated membrane (endocyst), and spontaneous cure because of cyst rupture into bronchi [11,14,15,17]. Patients are mostly asymptomatic and complications uncommon [11,14,15]. Percutaneous aspiration and drainage have been successfully accomplished for therapeutic and diagnostic purposes [12,15,17]. Anaphylaxis and secondary seeding have been extremely rare even in spontaneously ruptured cases [11,14,18]. Thus, in contrast to the global guidelines [6], in endemic cases in Alaska and Canada surgery is limited to symptomatic, infected or rapidly growing cysts [11,12,14-17]. Consistent with the clinical picture of our patient, children are more frequently symptomatic and prone to complications (e.g. bacterial secondary infections) from a cyst rupture, probably because the cysts occupy a large proportion of the lung volume, and the small calibre of bronchi prevents expectoration of parasite remnants [12].

The causative agents of sylvatic CE in the old clinical reports presumably represented the same genotypes (G8 and G10) that occur across the circumpolar north today. So far, G10 has been recorded only in three human cases: the first in southern Mongolia in 2011, the second in north-western Sakha Republic, Russia in 2013, and the most recent one in north-eastern China in 2015 [19-21]. All cases were treated surgically but the clinical course was not presented. The genotype G8 has been reported only once; the patient was operated in southeastern Alaska in 1999 [18,22]. The case, with multiple cysts disseminated in the peritoneal cavity, was atypical as sylvatic CE [18]. It is not known whether these different genotypes of E. canadensis correlate with differences in the clinical presentation of sylvatic CE.

![Figure 2](https://www.eurosurveillance.org/)

*Figure 2*  
Gram stain of the cyst wall showing two *Echinococcus* hooks, autochthonous cystic echinococcosis case in a child, east Finland, March 2015

Scale bar 30 µm.
The case presented here most likely originated in the sylvatic cycle via dogs. Given the slow growth of the larval stage and short lifespan of adult parasites, it is unclear whether the cyst in the patient was attributed to the infection detected in the dogs of the patient’s family. Although fed only with commercial dog foods, the dogs may have eaten raw viscera, e.g. during hunting, unnoticed by the owner.

Cervid offals are usually discarded at the shooting site. Even though wild scavengers operate rapidly, potentially infected offals are easily available during the moose hunting season for hunting dogs and others roaming freely in the woods. To prevent transmission, raw unrefrigerated or uncooked cervid viscera are not to be given to dogs, and cestocidic medication is to be regularly administered to hunting dogs before and after the hunting season [7,23]. Although sylvatic human CE is rare and sporadic, veterinary authorities should inform hunters and dog owners about these precautions and the potential risk of CE in areas where E. canadensis occurs in wildlife. In Europe, this includes Finland, the Baltic countries and Russia [7], and probably also some other countries which belong to the distribution range of wolves.

**Conflict of interest**

None declared.

**Authors’ contributions**

Drafting the manuscript: SH, AL; commenting on and revising the manuscript: AK, MA, TH, JK, KH, EB, KV, ET, TH-K, AO; attending physicians: SH, MA, TH, JH, EB, KV; consulting specialists: AK, TH-K; microbiological diagnosis: JK; genetic identification: AL; radiological interpretation: ET; veterinary diagnostics: AO.

**References**