The Rho GTPase Cdc42 regulates hair cell planar polarity and cellular patterning in the developing cochlea

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ABSTRACT

Hair cells of the organ of Corti (OC) of the cochlea exhibit distinct planar polarity, both at the tissue and cellular level. Planar polarity at tissue level is manifested as uniform orientation of the hair cell stereociliary bundles. Hair cell intrinsic polarity is defined as structural hair bundle asymmetry; positioning of the kinocilium/basal body complex at the vertex of the V-shaped bundle. Consistent with strong apical polarity, the hair cell apex displays prominent actin and microtubule cytoskeletons. The Rho GTPase Cdc42 regulates cytoskeletal dynamics and polarization of various cell types, and, thus, serves as a candidate regulator of hair cell polarity. We have here induced Cdc42 inactivation in the late-embryonic OC. We show the role of Cdc42 in the establishment of planar polarity of hair cells and in cellular patterning. Abnormal planar polarity was displayed as disturbances in hair bundle orientation and morphology and in kinocilium/basal body positioning. These defects were accompanied by a disorganized cell-surface microtubule network. Atypical protein kinase C (aPKC), a putative Cdc42 effector, colocalized with Cdc42 at the hair cell apex, and aPKC expression was altered upon Cdc42 depletion. Our data suggest that Cdc42 together with aPKC is part of the machinery establishing hair cell planar polarity and that Cdc42 acts on polarity through the cell-surface microtubule network. The data also suggest that defects in apical polarization are influenced by disturbed cellular patterning in the OC. In addition, our data demonstrates that Cdc42 is required for stereociliogenesis in the immature cochlea.

KEY WORDS: Planar polarity, Patterning, Development, Hair cell, Stereociliary bundle, Kinocilium, Microtubules, aPKC, Auditory

INTRODUCTION

The mammalian auditory sensory epithelium, the organ of Corti (OC) of the cochlea, consists of sensory hair cells and supporting cells. These cells are characterized by unique morphologies, specializations of their actin and microtubule cytoskeletons and complexity in cytoarchitectural organization as an epithelium (Raphael and Altschuler, 2003). One row of inner hair cells (IHCs) and three rows of outer hair cells (OHCs) together with interdigitated supporting cells form a checkerboard pattern, created by Notch signaling-mediated lateral inhibition (Eddison et al., 2000) and convergent extension (CE)-based cellular movements that are supported by cell-adhesive molecules (McKenzie et al., 2004; Wang et al., 2005; Togashi et al., 2011).

The mechanotransduction organelle at the hair cell’s apical surface, the stereociliary bundle, comprises rows of filamentous actin (F-actin)-rich stereocilia arranged in graded heights into a V-shaped structure. The vertex of the developing bundle is marked by the presence of a specialized primary cilium, the kinocilium, that is crucial for determining the orientation and morphology of the bundle (Ross et al., 2005; Jones et al., 2008; Cui et al., 2011; Sipe and Lu, 2011). The uniform orientation of hair bundles at the surface of the OC – the vertex of each bundle pointing laterally – defines planar cell polarity (PCP) at tissue level. In addition, hair cells show planar polarity at subcellular level, manifested as normally oriented bundles coupled with asymmetric bundle morphology, i.e. positioning of the kinocilium at the vertex, behind the longest stereocilia. We refer to hair cell intrinsic polarity in this case. In the OC of the mouse, planar polarity and cellular organization are established during late-embryogenesis. Structural maturation during the early postnatal life reinforces the polarized cellular structures, an event that is essential for proper sound perception (Forge et al., 1997).

The non-canonical Wnt/PCP pathway is required for the establishment of PCP in the OC, as evidenced by random hair bundle orientation in mutant mice in which the core components of this pathway are inactivated (Curtin et al., 2003; Montcouquiol et al., 2003; Wang et al., 2006). Much less is known about the molecules underlying the establishment of hair cell intrinsic polarity, a process that can function downstream or parallel of the core PCP pathway. Also, molecular interactions between the core PCP proteins and the regulators of intrinsic hair cell polarity are poorly understood. Recent studies suggest that, in addition to the mechanisms operating at the hair cell apex (Sipe and Lu, 2011; Sipe et al., 2013; Ezan et al., 2013; Tarchini et al., 2013), hair cell intrinsic polarity is influenced by mechanisms originating from the junctions between hair cells and supporting cells (Fukuda et al., 2014).

In non-mammalian vertebrates, members of the Rho GTPase family mediate cell polarization through regulation of cytoskeletal reorganization, formation of junctional complexes, and activation and localization of polarity proteins (Schlessinger et al., 2009). In the developing OC of the mouse, the Rho GTPase Rac1 has been shown to regulate apical polarity of hair cells (Grimsley-Myers et al., 2009). We have previously shown that another Rho GTPase, Cdc42, is required for the development of apical polarity of supporting cells of the OC during the early postnatal life (Anttonen et al., 2012). In the present study, we postulate that Cdc42 regulates apical polarization of auditory hair cells as well. As hair cell polarity is established during late-embryogenesis, we concentrate on this prenatal period. Interestingly, a well-known effector of
Cdc42, the atypical protein kinase C (aPKC), has recently been shown to be involved in the establishment of apical polarity of embryonic hair cells (Ezan et al., 2013; Tarchini et al., 2013). We show here that Cdc42 is required for cellular patterning in the OC and for the establishment of planar polarity of hair cells. In addition, we demonstrate that Cdc42 regulates stereociliogenesis in the immature cochlea.

MATERIALS AND METHODS

Mice

Mice homozygous for the floxed Cdc42 allele (Cdc42fl/fl) were crossed with mice carrying the Fgfr3-iCre-ERT² transgene to obtain Cdc42fl/fl;Fgfr3-iCre-ERT² mice, encoding guanine nucleotide binding sequence is flanked by loxP sites (Wu et al., 2006). Cdc42loxPfl/fl;Fgfr3-iCre-ERT² and Cdc42loxPfl/fl mice were used as control animals. Genotyping by PCR was conducted as previously described (Wu et al., 2006; Young et al., 2010). To study the characteristics of iCre-mediated recombination Fgfr3-iCre-ERT² transgenic mice were bred with the ROSA26tm14(CAG-tdTomato) Cre-conditional reporter mice (obtained from the Jackson Laboratory) to generate Fgfr3-iCre-ERT²;Ai14(tdTomato) animals. The Ai14(tdTomato) transgene expression was detected by direct visualization of the tdTomato native fluorescence in tails of the animals. Timed pregnancies were established by the detection of vaginal plug, taken at the morning of plug observation as embryonic day 0.5 (E0.5). Both females and males were used in the analysis. Mice lines were maintained in a mixed background. All animal work has been conducted according to relevant national and international guidelines. Approval for animal experiments has been obtained from the National Animal Experiment Board.

Induction of iCre-mediated recombination

Pregnant mice were injected intraperitoneally with 3 mg of tamoxifen (Sigma, prepared as described earlier; Anttonen et al., 2012) at E13.5 and E14.5 or at E15.5 and E16.5. Dams were sacrificed and embryos dissected at E18.5. For postnatal studies, E18.5 embryos were surgically delivered and transferred to foster mothers. Pups were killed at postnatal day 6 (P6) and P18.

Whole mount specimens

For confocal microscopy, inner ears were dissected from E18.5 embryos and fixed in 4% paraformaldehyde (PFA) in PBS for 5 h. P6 and P18 cochleas were perilymphatically fixed with PFA before immersion in this fixative. For Cdc42 antibody staining, dissected inner ears were fixed in ice-cold 10% trichloroacetic acid (TCA) for 1 h. For immunofluorescence, whole mounts were blocked for 30 min with 10% normal serum in PBS containing 0.25% Triton X-100 (PBS-T), followed by incubation overnight at +4°C with the appropriate primary antibodies in PBS-T. The following primary antibodies were used: rabbit monoclonal acetylated tubulin (Cell Signaling Technology); mouse monoclonal acetylated tubulin; rabbit polyclonal gamma-tubulin; rabbit monoclonal Cdc42 (all from Molecular Probes/Invitrogen); mouse monoclonal Cdc42, exons 2 and 3 (Cell Signaling Technology); mouse monoclonal aPKCζ; rabbit monoclonal aPKCA/γ; rabbit monoclonal phospho-PKCζ/α; mouse monoclonal Rab11a (all from BD Biosciences); goat polyclonal Prox1 (R&D Systems); mouse monoclonal fodrin (Ylikoski et al., 1992); mouse monoclonal Rab11. Detection was performed using the Vectastain Elite ABC kit or Vectastain Mouse-On-Mouse kit and the diamobenzidine substrate kit (all from Vector Laboratories). Sections were counterstained with 3% methyl green and mounted in Permount (Fisher Scientific).

In situ hybridization was performed with [35S]-labeled Cdc42 (Cappello et al., 2006) and Fgfr3 (Peters et al., 1993) riboprobes on PFA-fixed paraffin sections. Hematoxylin was used for counterstaining. Sections were analyzed with a BX61 microscope (Olympus) using bright- and darkfield optics. Images were acquired through the DP70 CCD color camera and cell’F software (Olympus) and processed using Adobe Photoshop CS6 (Adobe Systems). For visualization in situ hybridization signals, autoradiographic silver grains in the darkfield image were selected, colored red and superimposed onto the brightfield image. For all antibodies and probes used in paraffin sections, a minimum of 4 cochleas per genotype were prepared for histological analysis.

Serial block-face scanning electron microscopy

Specimens were prepared and processed for serial block-face scanning electron microscopy (SBEM) and data was analyzed as previously described (Anttonen et al., 2014).

Hair bundle orientation

To measure the hair bundle orientation, a line was drawn from the place of the kinocilium through the center of the bundle (bisecting line). Another line was drawn parallel to the medio-lateral (planar polarity) axis. The angle of bundle orientation was defined as the angle formed between these lines. In control animals it is closer to 0°. Oriana 3 plots were drawn from the orientations of OHCs (OH1-OH3). The orientation was divided into 5 sections, each bar showing cell numbers within that 5° section. A total number of 119 and 88 OHCs from the medial coil of mutant (n=3) and controls (n=3) cochleas, respectively, were used for analysis.

Correlation between hair cell shape and bundle orientation

In the medial coil of the cochlea, OHC diameters were measured at the level below adherens junctions, and stereociliary bundle orientations defined as described in the previous paragraph, using the Image-Pro Plus software (Media Cybernetics) (n=76 OHCs, 4 mutant cochleas; n=21 OHCs, 2 control cochleas). Regression analysis was performed using the IBM SPSS Statistics and Microsoft Excel softwares.

RESULTS

Fgfr3-iCre-ERT² drives efficient recombination in the embryonic organ of Corti

Cdc42 is widely expressed and has several roles in developing hair cells (Melendez et al., 2011). Consistently, Cdc42 knockout mice show early-embryonic lethality (Chen et al., 2000). As we wished to reveal the role of Cdc42 during cellular differentiation in the late-embryonic OC, a conditional and inducible approach was required to bypass the likely effects of Cdc42 on dividing progenitor cells of the early otocyst. We used the Fgfr3-iCre-ERT² transgenic mouse line (Young et al., 2010) that was crossed with mice carrying the Fgfr3-iCre-ERT² transgenic mouse line (Young et al., 2010) that was crossed
with the Cdc42loxP/loxP mutant mice (Wu et al., 2006). We have previously shown that Fgfr3-iCre-ERT2 mice can be used for inducible, supporting cell-specific gene inactivation in the postnatal OC (Anttonen et al., 2012). To study recombination characteristics in the embryonic cochlea, Fgfr3-iCre-ERT2 mice were crossed with the ROSA26m14(CAG-tdTomato) reporter mice to generate Fgfr3-iCre-ERT2;Ai14(tdTomato) mice. Recombined cells were recognized by RFP immunohistochemistry in paraffin-embedded cross-sections through the cochlea (Fig. 1A–E). Recombination was induced at E13.5 and E14.5, at the stages when Fgfr3 starts to be expressed in the presumptive OC (Hayashi et al., 2010). At this age, precursor cells have exited the cell cycle and are initiating differentiation into hair cells or supporting cells (Ruben, 1967; Chen and Segil, 1999). At E18.5, basal and medial coils of the cochlea of the Fgfr3-iCre-ERT2;Ai14(tdTomato) double-transgenic mice showed recombination in OHCs and in two types of supporting cells, the Deiters’ and pillar cells (Fig. 1B–D). Importantly, the whole population of these cells showed recombination in the basal and medial coils, the regions of the cochlear duct analyzed in the present study. IHCs lacked RFP signal (asterisks in Fig. 1B–D). The apical cochlear coil showed recombination only in scattered sensory epithelial cells (Fig. 1E). This recombination pattern correlates well with the cell type-specific expression of Fgfr3 and with the dynamic Fgfr3 expression along the base-to-apex differentiation gradient in the developing cochlear duct (Hayashi et al., 2010). We conclude that Fgfr3-iCre-ERT2-mediated recombination efficiently targets OHCs, Deiters’ cells and pillar cells of the late-embryonic OC.

Cdc42 expression in the embryonic organ of Corti

To study Cdc42 mRNA expression in the developing cochlea, we used radioactive in situ hybridization on paraffin sections. As analyzed at E13.5, E15.5, E18.5 and P0, Cdc42 was ubiquitously expressed in the presumptive and, later, in differentiating hair cells and supporting cells (Fig. 2A,B; data not shown). Signal obtained by the Cdc42 antisense probe clearly surpassed the background signal obtained with the Cdc42 sense probe (Fig. 2B,C). Upon tamoxifen-induced recombination, the level of Cdc42 antisense signal in the OC of the Cdc42loxP/loxP;Fgfr3-iCre-ERT2 mice (hereafter termed as the mutant mice) was comparable to background signal, consistent with prior studies (data not shown; Anttonen et al., 2012).

We next used immunocytochemistry to localize Cdc42 protein in the OC at E18.5. Comparable results were obtained with two commercial antibodies in TCA-fixed whole mount specimens. Cdc42 was localized to the OHC apex (Fig. 2D–G). Expression was found in the region around the basal body from where microtubules radiate at the cell surface (Fig. 2D). Weaker expression was found in the apical cytoplasm, on the medial side of the bundle (Fig. 2G). In addition, stereocilia showed strong expression (Fig. 2D) as well as the contact sites between supporting cells and OHCs at the level of adherens junctions (Fig. 2G). Cdc42 was also expressed in the apices of IHCs (data not shown) and supporting cells (Fig. 2G). Cdc42 immunostaining was abolished from OHCs of the Cdc42loxP/loxP;Fgfr3-iCre-ERT2 mice while the unrecombined IHCs showed maintained expression (Fig. 2E,E’; data not shown).
Cdc42 inactivation alters cellular patterning and cell shapes, but not global morphology of the organ of Corti

Whole mount specimens and paraffin-embedded cross-sections revealed an unaltered global morphology of the cochlea of the Cdc42 mutant mice at E18.5, including unaltered length and width of the OC (supplementary material Fig. S1A,B). To find out whether Cdc42 inactivation triggers cell cycle activity, expression of the proliferation marker Ki-67 and the cyclin-dependent kinase inhibitor p27Kip1, a negative cell cycle regulator, were studied on paraffin sections. The OC of both control and mutant mice showed strong p27Kip1 immunofluorescence in supporting cells and absence of Ki-67 staining in hair cells and supporting cells (supplementary material Fig. S1C–E). This suggests that Cdc42 is not involved in the regulation of the postmitotic state of the auditory sensory epithelial cells. Also cell survival was unaffected, based on the lack of staining for cleaved caspase-3, an apoptosis marker (data not shown). Further, expression of Prox1 in supporting cells (supplementary material Fig. S1D–E), and Gfi1 (data not shown) and myosin 6 in hair cells were similar in control and mutant specimens (supplementary material Fig. S1F,G). Thus, cell fate decisions are not affected by Cdc42 inactivation.

To study the possible involvement of Cdc42 in cellular patterning in the OC, whole mount specimens were labeled with phalloidin, marking F-actin. Cochlear specimens from control mice at E18.5 displayed precisely aligned hair cell rows, separated by supporting cell rows (Fig. 3A). In contrast, cochleas of the Cdc42 mutant mice showed occasional misplacement of OHCs between the cell rows, creating direct contacts between OHCs, as opposed to normal OHC patterning with intervening supporting cells (Fig. 3B). These defects were seen in the medial coil of the cochlea and were mostly absent in the basal and apical coils. The unrecombined IHCs were properly aligned along the length of the cochlear duct of the mutant mice (Fig. 3A,B).

Proper alignment of the cells of the OC relies on dynamic remodeling of cell-cell contacts, a process dependent on adherens junctions (Chacon-Heszele et al., 2012). Therefore, we next studied whether changes in the expression of the adherens proteins E-cadherin, β-catenin as well as nectin 2 and nectin 3 could explain the OHC patterning defects. At E18.5, these proteins showed comparable expression in the lateral contact sites between OHCs and supporting cells (Deiters’ cells) in both mutant and control specimens (Fig. 3C–F). Immunostaining also revealed comparable expression of the tight-junction-marker ZO-1 in OHCs of the two genotypes (data not shown). Thus, cellular disorganization triggered by Cdc42 depletion is not caused by disturbed recruitment of these junctional components to the plasma membrane.

In addition to patterning defects, cochleas of mutant mice showed abnormally shaped OHCs, revealed in phalloidin-labeled whole mount specimens at birth (Fig. 4A–H). OHCs with squeezed morphology were found (Fig. 4E,G) and their nuclei were positioned at variable heights in the apico-basal axis of the sensory epithelium, in contrast to OHCs of control animals with uniformly positioned nuclei (Fig. 4F,H). Similar to patterning defects, OHCs with abnormal morphology were concentrated to the medial coil and were intermingled with OHCs with a normal morphology. To find out whether the atypical OHC shape was linked with changes in the expression of actin-interacting-proteins, paraffin sections were immunolabeled for alpha-fodrin (non-erythroid spectrin) that cross-links actin filaments. Fodrin is a constituent of the cuticular plate and cortical lattice of hair cells (Ylikoski et al., 1992). In agreement, control specimens showed fodrin expression in these regions in the apical portion of hair cells (Fig. 4I,J). In the Cdc42 mutant mouse, the cuticular plate staining was unaltered, but fodrin as well as phalloidin labeling in the cortex extended towards the basal side of the squeezed OHCs. This was best seen in top-down views showing F-actin accumulation at the most shrunken level of the atypical, flask-shaped OHCs (Fig. 4D). These data suggest that Cdc42 regulates the cortical actin cytoskeleton and that defects in this regulation lead to cell shape abnormalities.

Cdc42 inactivation leads to stereociliary bundle misorientation in outer hair cells

The read-out of PCP in the OC is the uniform orientation of hair bundles in the plane parallel to the epithelial surface. In cochlear whole mounts prepared from control and mutant mice at E18.5, phalloidin labeling was used to visualize the F-actin-rich stereocilia. Acetylated tubulin was used to mark kinocilia (Fig. 5A–E). In control specimens, hair bundles were uniformly oriented across the epithelium and kinocilia were positioned at the vertex of the bundles (Fig. 5A–A’). Cochleas of mutant mice showed OHC bundles with random orientation, vertices pointing...
to many directions, demonstrating an abnormal planar polarity phenotype. Unrecombined IHCs showed unaltered bundle orientation (Fig. 5A–E). Similarly to patterning and shape abnormalities, bundle orientation defects were concentrated, but not restricted to the medial coil (Fig. 5B–B'). Quantification performed in the medial coil revealed a clear difference in OHC bundle orientations between mutant and control specimens (Fig. 5C). In the medial coil, the same OHC often displayed all abnormalities. In contrast, the basal coil showed OHCs with misoriented bundles lacking patterning and shape defects (Fig. 5D,E). Thus, bundle misorientation can occur independently of the defects at the level of the cell body. This was also found in experiments where induction of recombination was shifted from E13.5 and E14.5 to E15.5 and E16.5, assuming that this later onset of Cdc42 inactivation does not cause patterning defects in the medial coil. Despite the fact that there was a lack of patterning and shape defects, OHCs still showed a planar polarity defect when analyzed at E18.5 (data not shown).

To quantitate the relationship between disturbances in stereociliary bundle orientation and shape of OHCs, we measured cell diameters at the level of adherens junctions at E18.5 and correlated these values to bundle orientation (supplementary material Fig. S2). Orientation of hair bundles did not correlate with the diameter of cell bodies. Thus, these two phenotypical changes of OHCs of the Cdc42 mutant mouse can occur independently of each other.

**Expression of the core PCP components is not affected by Cdc42 inactivation**

Based on the altered PCP phenotype of OHCs of the Cdc42 mutant mice, we next examined the possible link between Cdc42 and the Wnt/PCP pathway (Fig. 5F–I). Cochlear surface specimens were prepared at E18.5 and stained for Vang-like 2 (Vangl2) (Montcouquiol et al., 2003) and Frizzled 6 (Wang et al., 2006), the core components of the PCP pathway. In control animals, Vangl2 was expressed at the level of adherens junctions, in contact sites between supporting cells and the medial side of the OHC plasma membranes (Fig. 5F,F'.G). By high resolution microscopy, this expression has been localized to supporting cells (Montcouquiol et al., 2006; Giese et al., 2012). The medial coil of mutant mice showed unaltered Vangl2 expression (Fig. 5H,H'). To exclude the effects of cellular disorganization on Vangl2 expression, we focused on the basal coil where OHCs displayed bundle disturbances exclusively. Triple-labeling for F-actin, acetylated tubulin and Vangl2 revealed unaltered Vangl2 expression, despite hair bundle abnormalities (Fig. 5I). Similarly, membraneous Frizzled 6 expression on the medial side of the OHC apex (Wang et al., 2006) was unaltered in mutant animals (data not shown). These findings exclude a direct relationship between Cdc42 and Wnt signaling.

**Cdc42 inactivation leads to stereociliary bundle dysmorphology and kinocilia mispositioning in outer hair cells**

In OHCs of the mutant mice, phalloidin labeling demonstrated dysmorphic hair bundles. In most cases, bundles were misoriented as well (Fig. 6A–D), but dysmorphic bundles lacking an orientation defect were also found (Fig. 6C,D). Abnormal bundle morphology, a read-out of altered hair cell intrinsic polarity (Sipe and Lu, 2011; Fukuda et al., 2014), was manifested as flat, wavy or inverted shape, often with no clear vertex. Importantly, bundle dysmorphology was often associated with abnormal positioning of the kinocilium relative to the bundle, as revealed by using acetylated tubulin as a kinocilium marker (Fig. 6C–D'). These findings were ultrastructurally confirmed by SBEM (Fig. 6E,F).

Prior studies have shown that normal hair bundle morphology depends on proper development and migration of the kinocilium/basal body complex (Ross et al., 2005; Jones et al., 2008; Cui et al., 2011; Sipe and Lu, 2011). During late-embryogenesis, this complex starts to develop at the center of the hair cell’s surface and it migrates towards the lateral cell periphery. In parallel, surrounding microvilli elongate to form stereocilia of graded heights. Around birth, the immature bundle is V-shaped with the kinocilium at the vertex, next to the tallest stereocilium (Jones and Chen 2008). Interestingly, Cdc42 has been shown to be crucial for the assembly of primary cilia in other cell types (Zuo et al., 2011; He et al., 2012; Choi et al., 2013). Based on these prior data, we next asked whether Cdc42 depletion influences OHC planar polarity by regulating the assembly or migration of the kinocilium/basal body.

Cochleas at E18.5 were stained for acetylated tubulin marking the axoneme of kinocilium, for γ-tubulin marking centrioles and...
for the GTPase Rab11a that has been shown to mark the basal region of primary cilia and to be critical for primary ciliogenesis (Knödler et al., 2010; Westlake et al., 2011; He et al., 2012). Cdc42 has been proposed to be involved in the process of primary ciliogenesis by targeting the exocyst and Rab11a-Rabin-Rab8 trafficking complexes (Zuo et al., 2011; He et al., 2012). We found strong, condensed Rab11a expression in the base of hair cell kinocilia, similar as reported in primary cilia of other cell types. This staining was found both in control and mutant specimens (Fig. 6A–D). Phalloidin labeling shows normal hair bundle orientation in the basal coil of the control cochlea. (E) The basal coil of the mutant specimen shows occasional OHCs with misoriented bundles (arrows), but hair cell rows are normally organized. (F–G) Triple-labeling for F-actin, Vangl2 and acetylated tubulin (kinocilia pseudocolored in yellow) shows Vangl2 expression in the contact sites between the OHC’s medial and Deiters’ cells in the control specimen. (H,H) In the medial coil of the mutant cochlea, Vangl2 expression pattern is influenced by cellular disorganization. (I) In the basal coil of the mutant cochlea where cellular disorganization is absent, Vangl2 expression domain is similar as in the control specimen (compare to Fig. 5G). Abbreviations: OHC, outer hair cell; IHC, inner hair cell. Scale bar shown in I: A–E, 6 μm; F–H, 5 μm; G,I, 2 μm.

**Cdc42 inactivation impairs microtubule organization and alters aPKC expression in the apices of outer hair cells**

In the apices of epithelial cells, astral microtubules radiate from the basal body and are captured by plus-ends to the cell cortex. Pulling forces provided by microtubules are essential for the migration of the basal body and primary cilium and for docking of these structures to their final position (Tang and Marshall, 2012). Recent studies on cochlear hair cells have linked abnormal positioning of the kinocilium to disorganized cell-surface microtubule network (Ezan et al., 2013; Sipe et al., 2013). Compared to control specimens (Fig. 7A–A1), acetylated tubulin-staining at E18.5 showed disorganized microtubules at the OHC surface in the Cdc42 mutant mice (Fig. 7B–B3). Both microtubules around the basal body and those radiating at the cell surface were abnormal. Further, both dysmorphology of the hair bundle and abnormal position of the basal body/kinocilium correlated with microtubule disorganization, a finding clearly after primary kinocilium/basal body migration (Lepelletier et al., 2013).
seen in high magnification views (Fig. 7B1–B3). These results suggest that altered planar polarity of OHCs is mediated by microtubular disturbances.

In several cell types, Cdc42 regulates polarization through aPKC, and this signaling has been shown to control cell polarity through the regulation of microtubule dynamics (Etienne-Manneville, 2004; Etienne-Manneville et al., 2005). Therefore, we next focused on aPKC expression at the OHC’s apical surface. It has been previously shown that the aPKC isoform λ/γ is not expressed in the OC around birth (P2), but becomes upregulated a few days thereafter (Anttonen et al., 2012). We found that the aPKCγ isoform is expressed in the hair cell apex during late-embryogenesis. Most prominent expression was found in the medial plasma membrane, opposite to the vertex of the hair bundle (Fig. 8A–A’). Weaker cytoplasmic expression was seen on the medial side at the cell surface (Fig. 8A–A’). These findings are consistent with prior data revealing that the hair bundle is positioned at the interface of aPKC-positive and -negative domains at the hair cell apex (Ezan et al., 2013; Tarchini et al., 2013). Based on triple-labeling with phalloidin and antibodies against ZO-1 and aPKCγ or phospho-aPKC (marking activated kinase), aPKCγ (data not shown) and phospho-aPKC (supplementary material Fig. S3) were localized to the level of tight junctions of OHCs.

In OHCs of the Cdc42 mutant mice, the membraneous aPKC expression domain was altered and it corresponded to the degree of abnormal rotation of the hair bundle (Fig. 8B–D’). This is most clearly seen in the high magnification view of an OHC with a completely turned bundle (Fig. 8D’). Further, in OHCs where the bundle and kinocilium were abnormally positioned at the very lateral edge of the cell surface, membraneous and cytoplasmic aPKC expression was extended to this lateral side (Fig. 8D’). Based on triple-labeling with phalloidin and antibodies against aPKCγ and acetylated tubulin, microtubule disorganization was coupled with hair bundle defects and altered aPKC expression (Fig. 8C–C”). These results suggest that Cdc42 and its putative downstream effector, aPKC, regulate the establishment of hair cell planar polarity via the microtubule cytoskeleton.

**Planar polarity defect is maintained and stereociliogenesis impaired in outer hair cells of the Cdc42 mutant mice early postnatally**

To study the fate of OHCs with abnormal apical polarity, cochleas of the Cdc42<sup>loxP/loxP,Fgfr3-iCre-ERT2</sup> mice (tamoxifen administration at E13.5 and E14.5) were analyzed at P6. Phalloidin labeling showed a normal complement of auditory hair cells in these animals, demonstrating that Cdc42 inactivation does not abrogate the survival of juvenile hair cells. OHCs with misoriented and dysmorphic stereociliary bundles were maintained in these cochleas. Acetylated tubulin immunostaining revealed the presence of the kinocilium in OHCs of both mutant and control mice. Notably, the staircase pattern of stereociliary bundles was in many cases disrupted. Bundles also appeared fragmented, suggesting that stereociliogenesis itself was impaired (Fig. 9A–D), consistent with strong Cdc42 expression in stereocilia (Fig. 2D). To confirm the involvement of Cdc42 in stereociliogenesis, P18 mutant mice were analyzed. In contrast to OHCs of control specimens that showed long and precisely arranged stereocilia, OHCs of the mutant mice had fragmented hair bundles consisting of stereocilia of variable lengths, some being very short (Fig. 9E,F). The unrecombined IHCs with normal bundles served as controls (Fig. 9G,H). Interestingly, at P18, besides the defects in OHC stereociliogenesis, scattered OHC loss...
Cdc42 conditional, inducible shortly after birth (Anttonen et al., 2012). In that study, a regulate later developmental events, as shown in the cochlea Kesavan et al., 2009; Melendez et al., 2013). Cdc42 can also importance of Cdc42 in the establishment of apico-basal in vivo influences apical cell polarization (Etienne-Manneville, 2004). Cdc42 is involved in many cellular processes by regulating DISCUSSION evidence that Cdc42 regulates postnatal stereociliogenesis. was observed (Fig. 9F). Together, in addition to the control of hair cell patterning, shape and planar polarity, these results give evidence that Cdc42 regulates postnatal stereociliogenesis.

Fig. 8. Altered aPKC\(\epsilon\) expression in outer hair cells of Cdc42\textsuperscript{loxP/loxP}; Fgfr3-iCre-ERT\(^{2}\) mice. Confocal images of whole mount specimens dissected from the medial cochlear coil of control and mutant mice at E18.5 and labeled for F-actin, acetylated tubulin and aPKC\(\epsilon\). All images are shown in medial-lateral orientation, as indicated in (A). (A–A\(\prime\)) In the control specimen, aPKC is expressed in the cortical and, weaker, in the cytoplasmic domain at the OHC surface, medial to the hair bundle. Asterisks mark the lateral, aPKC-negative domain. (B–B\(\prime\)) In the mutant specimen, both cortical and cytoplasmic aPKC expression domains are rotated with respect to hair bundle misorientation. (C–C\(\prime\)) In the mutant specimen, triple-labeling for acetylated tubulin, phallolidin and aPKC shows that the rotation of aPKC expression parallels the rotation of the astral microtubule network and hair bundle misorientation. (D–D\(\prime\)) High magnification views of two OHCs with abnormal bundles. The OHC below has a completely turned bundle. Asterisks mark the concise aPKC-free area in these cells (compare to control OHCs in Fig. 8A\(\prime\)). Scale bar shown in D\(\prime\): A–C\(\prime\), 5 \(\mu\)m; D–D\(\prime\), 2 \(\mu\)m.

The CE process of cell movements consists of directed migration and polarized rearrangement of cells (Wallingford et al., 2002). CE has an important role during cochlear development. Between E14.5 and E18.5 in the mouse, cells of the OC become arranged into precise rows by means of CE, leading to thinning and lengthening of the cochlear duct (McKenzie et al., 2004; Wang et al., 2005). Interestingly, Cdc42 has been shown to regulate CE during gastrulation in non-mammalian species (Choi and Han, 2002; Yeh et al., 2011). It is likely that the possible role of Cdc42 in CE in the cochlea could be revealed with our gene inactivation strategy, not only because of the timing of onset of Cdc42 inactivation (between E13.5 and E14.5), but also because the inactivation took place in OHCs and two types of supporting cells, the pillar and Deiters’ cells, constituting a broad region of the cochlear epithelium. We did not see changes in dimensions of the OC of the Cdc42\textsuperscript{loxP/loxP}; Fgfr3-iCre-ERT\(^{2}\) mice, excluding a primary role for Cdc42 in the regulation of global CE events in the cochlea. This suggestion is supported by unaltered expression of the adherens junction components E-cadherin, \(\beta\)-catenin and nectins in the OC of the mutant mice, taking into account prior data that E-cadherin and N-cadherin expressions are abolished in hair cells when CE is impaired (Chacon-Heszele et al., 2012). However, we cannot exclude the possibility of a fine-grained function of Cdc42 in CE in the cochlea.

OHCs of the Cdc42 mutant mice showed misoriented and dysmorphic stereociliary bundles together with mispositioned kinocilia, the characteristic features of an abnormal hair cell intrinsic polarity phenotype. The kinocilium is essential for driving proper hair bundle morphogenesis (Ross et al., 2005; Sipe
In the mutant mice, we did not find defects in kinocilium assembly or primary migration of the kinocilium/basal body complex to the lateral side of OHC surface. The basal body serves as a microtubule-organizing center (MTOC) for cytoplasmic microtubules. Microtubules are connected to the cell cortex and have a crucial role in defining cell shape and polarity (Li and Gundersen, 2008). Interestingly, during the establishment of planar hair cell polarity, defects in cell-surface microtubules have been shown to lead to kinocilium/basal body mispositioning and improper bundle morphology and orientation (Sipe and Lu, 2011; Sipe et al., 2013, Ezan et al., 2013). We found that the microtubule network at the apex of Cdc42-depleted OHCs was disorganized, suggesting that the abnormal hair cell intrinsic polarity phenotype could result from perturbed microtubule dynamics. This suggestion is supported by our localization data, revealing Cdc42 expression in the pericentrosomal region, the region from where microtubules radiate (Mogensen et al., 1997). One candidate factor mediating the effect of Cdc42 in this region is dynein, a member of the microtubule motor protein complex. Similar to Cdc42, dynein has been localized in developing hair cells to the region around the centrosome and has been suggested to be involved in basal body positioning (Sipe et al., 2013). Interestingly, in other cellular models, Cdc42/dynein interaction orients microtubules radiating from the MTOC (Palazzo et al., 2001).

In addition to the expression of Cdc42 in the basal body region, we localized Cdc42 to the medial side of the OHC’s apical surface, so that its expression flanked the hair bundle position. Such an expression pattern of Cdc42 was comparable to that maintained on the lateral side of the OHC surface, indicating that Cdc42 is not a primary regulator of sustained planar polarity, as opposed to Lis1 (Sipe et al., 2013). Further, as misoriented hair bundles were maintained in these animals, the machinery responsible for polarity refinement (Copley et al., 2013) cannot overcome the polarity defects triggered by Cdc42 inactivation at late-embryogenesis.

In conclusion, consistent with the involvement of Cdc42 in multiple signaling cascades and in the regulation of cytoskeletal dynamics, the present results show that Cdc42 regulates various aspects of hair cell differentiation. Our results suggest that Cdc42 is involved in OHC stereociliogenesis early postnatally, likely through the regulation of actin dynamics (Ridley, 2006). Our data also demonstrate the role of Cdc42 in cellular patterning and in mediating planar polarity of embryonic OHCs. Together, these data show that Cdc42 is important for the establishment of proper hearing function.

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Competing interests
The authors have no competing or financial interests to declare.

Author contributions
A.K. and U.P. conceived and designed the experiments. A.K. performed the experiments. A.K. and UP analyzed the data and wrote the paper. M.L. and T.A. helped drafting the manuscript and all the authors reviewed the manuscript.

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