

Angioedema Center, 3rd Department of Internal Medicine, Semmelweis University, Budapest, Hungary; ⁵the Department of Allergology, Navy Hospital, Athens, Greece; ⁶the Allergy and Lung Function Office, University of Medicine and Pharmacy, Tirgu-Mures, Romania; ⁷the Department of Dermatology and Allergy, Charité-Universitätsmedizin Berlin, Germany; and ⁸the Laboratory of Molecular Diagnostics, Hungarian National Blood Transfusion Service, Budapest, Hungary. E-mail: agermen@med.uth.gr.

*These authors contributed equally to this work.

This work was partially supported by an unrestricted grant from Shire Hellas.

Disclosure of potential conflict of interest: M. Speletas is employed by the University of Thessaly School of Health Sciences; has received research support unrelated to this manuscript from Greek Secretariat of Research in Greece; has received lecture fees from Amgen; and has received travel support from Shire, Behring, Bristol-Meyer, and Novartis. F. Psarros has received travel support from Shire. D. Moldovan has received travel support from Shire and CSL Behring; has received investigator fees from CSL Behring, Viropharma/Shire, and Pharming Technologies; and has received sponsorship of a local conference on HAE and PID from Pharming, Shire HGT, and CSL Behring. M. Magerl has received consultancy/honorarium fees from Shire, Viropharma, CSL Behring, and Sobi. E. Gramoustianou has received research support from Shire. A. E. Germeis has received research support from Shire and Novartis; has received lecture fees from Novartis and Amgen; and has received travel support from Shire. A. Avramouli has received research support from Shire. H. Varga has received travel support from CSL Behring, Pharming, and Shire. M. Maurer has received research support, consultancy and lecture fees, and travel support from Shire; has received consultancy and lecture fees from Biocryst and Viropharma; and is on the boards of Shire, Biocryst, and Viropharma. H. Farkas is on the boards of CSL Behring, Swedish Orphan Biovitrum, Shire, and Viropharma; and has received lecture fees from Swedish Orphan Biovitrum and Shire. The rest of the authors declare that they have no relevant conflicts of interest.

REFERENCES

1. Zuraw BL. Clinical practice. Hereditary angioedema. *N Engl J Med* 2008;359:1027-36.
2. Zahedi R, Bissler JJ, Davis AE 3rd, Andreadis C, Wisnieski JJ. Unique C1 inhibitor dysfunction in a kindred without angioedema, II: identification of an Ala443-Val substitution and functional analysis of the recombinant mutant protein. *J Clin Invest* 1995;95:1299-305.
3. Davis AE 3rd, Aulak K, Parad RB, Stecklein HP, Eldering E, Hack CE, et al. C1 inhibitor hinge region mutations produce dysfunction by different mechanisms. *Nat Genet* 1992;1:354-8.
4. Pappalardo E, Cicardi M, Duponchel C, Carugati A, Choquet S, Agostoni A, et al. Frequent *de novo* mutations and exon deletions in the C1 inhibitor gene of patients with angioedema. *J Allergy Clin Immunol* 2000;106:1147-54.
5. Speletas M, Boukas K, Papadopoulou-Alataki E, Tsitsami E, Germeis AE. Hereditary angioedema in Greek families caused by novel and recurrent mutations. *Hum Immunol* 2009;70:925-9.
6. Bors A, Csuka D, Varga L, Farkas H, Tordai A, Füst G, et al. Less severe clinical manifestations in patients with hereditary angioedema with missense C1INH gene mutations. *J Allergy Clin Immunol* 2013;131:1708-11.
7. Kalmar L, Hegedus T, Farkas H, Nagy M, Tordai A. HAEdb: a novel interactive, locus-specific mutation database for the C1 inhibitor gene. *Hum Mutat* 2005;25:1-5.
8. Farkas H. Pediatric hereditary angioedema due to C1-inhibitor deficiency. *Allergy Asthma Clin Immunol* 2010;6:18.
9. Bork K, Meng G, Staubach P, Hardt J. Hereditary angioedema: new findings concerning symptoms, affected organs, and course. *Am J Med* 2006;119:267-74.

Available online September 23, 2014.
<http://dx.doi.org/10.1016/j.jaci.2014.08.007>

Associations between the 17q21 region and allergic rhinitis in 5 birth cohorts

To the Editor:

The high heritability and comorbidity between allergic rhinitis and asthma suggest common etiologies and genetic susceptibility loci. Among the most robust signals for asthma is the chromosomal locus 17q21.¹ Studies that examined whether this strong asthma locus is also associated with allergic rhinitis

have yielded conflicting results. A recent large genome-wide association study (GWAS)² and a candidate gene study³ identified associations between genetic variants at the 17q21 locus and allergic rhinitis, in contrast to null findings in previous GWASs on allergic rhinitis^{4,5} and self-reported allergy.⁶ Large, well-defined studies are needed to clarify these inconsistencies.

We pooled data from 5 birth cohorts not included in the aforementioned GWASs (Children, Allergy, Milieu, Stockholm, Epidemiological Survey [BAMSE], Canadian Asthma Primary Prevention Study [CAPPs], German Infant study on the influence of Nutritional Intervention plus environmental and genetic influences on allergy development [GINIplus]/Lifestyle-related factors, Immune System and the development of Allergies in East and West Germany plus the influence of traffic emissions and genetics study [LISAplus], Prevention and Incidence of Asthma and Mite Allergy [PIAMA], and Study of Asthma Genes and the Environment [SAGE]; $N_{\text{total}} = 4624$ children, 92.7% white) and examined whether 7 single nucleotide polymorphisms (SNPs) at the 17q21 locus are associated with allergic rhinitis from early childhood to adolescence, and the effects of comorbidity with asthma on the associations.

Allergic rhinitis was defined as concomitantly reported rhinitis and positive sensitization to any aeroallergen (age range 4-16 years in pooled data). Seven SNPs, including top GWAS hits for asthma (rs7216389 and rs2305480),¹ were extracted from genotyped (BAMSE) or imputed genome-wide (other cohorts) data and coded as 0, 1, and 2 according to the number of effect alleles. Cohort-specific study designs, outcome definitions, and genotyping/imputation are provided in this article's [Methods section](#) and [Table E1](#) in the Online Repository at www.jacionline.org. The effect of each SNP on allergic rhinitis over time was analyzed using generalized estimating equation models (logit link and exchangeable correlation structure) adjusted for age and cohort (pooled models only). Associations were calculated per cohort and in the combined pooled data. Odds ratios with corresponding 95% CIs are presented.

Allergic rhinitis was reported at least once by 969 of the 4624 children ([Table I](#)). The proportion of cases generally increased with age (see [Table E2](#) in this article's [Online Repository](#) at www.jacionline.org). Effect allele frequencies were very similar across cohorts for all SNPs (see [Table E3](#) in this article's [Online Repository](#) at www.jacionline.org). The correlation between 5 of the SNPs (rs2305480, rs7216389, rs4065275, rs8076131, and rs12603332) was high ($r^2 > 0.7$, $D' > 0.9$) and more moderate between these 5 SNPs and the other 2 ($r^2 < 0.3$, $D' > 0.9$; rs17608925 and rs3744246).

Allergic rhinitis and 6 of the 7 studied SNPs were significantly associated in the pooled data ([Table I](#)). Because the SNPs are in high linkage disequilibrium, the associations are not independent. Cohort-specific risk estimates were consistent across all cohorts except the smallest cohort CAPPs (see [Table E4](#) in this article's [Online Repository](#) at www.jacionline.org). Associations stratified by sex were significant only among males, although the direction of effects was similar in males and females (data not shown).

Sensitivity analyses were conducted to assess whether associations with allergic rhinitis were modified by the copresence of asthma. Associations between the 17q21 variants and allergic rhinitis with concomitant asthma appeared more pronounced (although not significantly) than when allergic rhinitis or asthma was modeled independently ([Table I](#)). Risk

TABLE I. Pooled longitudinal associations between 7 SNPs at the 17q21 locus and health outcomes*

SNP	Gene	Effect allele	EAF	Allergic rhinitis† (case ever = 969 of 4624)		Asthma‡ (case ever = 571 of 4619)		Allergic rhinitis with concomitant asthma§ (case ever = 234 of 4019)		Allergic rhinitis without concomitant asthma§ (case ever = 816 of 4425)	
				N	OR (95% CI)	N	OR (95% CI)	N	OR (95% CI)	N	OR (95% CI)
rs2305480	<i>GSDMB</i>	A	0.45	4605	0.85 (0.77-0.94)	4600	0.83 (0.72-0.96)	4002	0.72 (0.59-0.88)	4407	0.89 (0.80-0.99)
rs7216389	<i>GSDMB</i>	T	0.49	4604	1.15 (1.04-1.27)	4599	1.15 (1.01-1.32)	4001	1.35 (1.12-1.63)	4408	1.10 (0.99-1.22)
rs4065275	<i>ORMDL3</i>	G	0.50	4421	1.14 (1.03-1.26)	4417	1.13 (0.98-1.29)	3859	1.35 (1.11-1.64)	4266	1.09 (0.98-1.22)
rs8076131	<i>ORMDL3</i>	G	0.45	4417	0.86 (0.77-0.95)	4413	0.85 (0.73-0.98)	3856	0.73 (0.60-0.89)	4262	0.89 (0.80-1.00)
rs12603332	<i>ORMDL3</i>	C	0.50	4618	1.15 (1.04-1.27)	4426	1.13 (0.98-1.29)	4014	1.34 (1.11-1.61)	4420	1.10 (0.99-1.23)
rs17608925	<i>ORMDL3</i>	C	0.11	4414	0.88 (0.74-1.05)	4410	0.99 (0.79-1.25)	3853	0.94 (0.67-1.32)	4259	0.89 (0.74-1.07)
rs3744246	<i>ORMDL3</i>	T	0.20	4607	0.86 (0.75-0.97)	4602	0.89 (0.75-1.06)	4004	0.83 (0.65-1.06)	4411	0.89 (0.77-1.02)

Case ever, Whether a child ever had a positive report at any follow-up; EAF, effect allele frequency; *GSDMB*, gasdermin B; N, number of children included in the model; OR, odds ratio; *ORMDL3*, ORM1-like protein 3.

*Models were adjusted for age and cohort.

†Age-specific controls were defined as those without allergic rhinitis. P values of significant pooled associations ranged from .002 for rs2305480 to .017 for rs3744246. Bonferroni-corrected P value corresponds to $.05/7 = .007$. The associated SNPs are in linkage disequilibrium ($r^2 > 0.2$, $D' > 0.9$).

‡Age-specific controls were defined as those without asthma.

§Age-specific controls were defined as those without allergic rhinitis or asthma.

||P < .05.

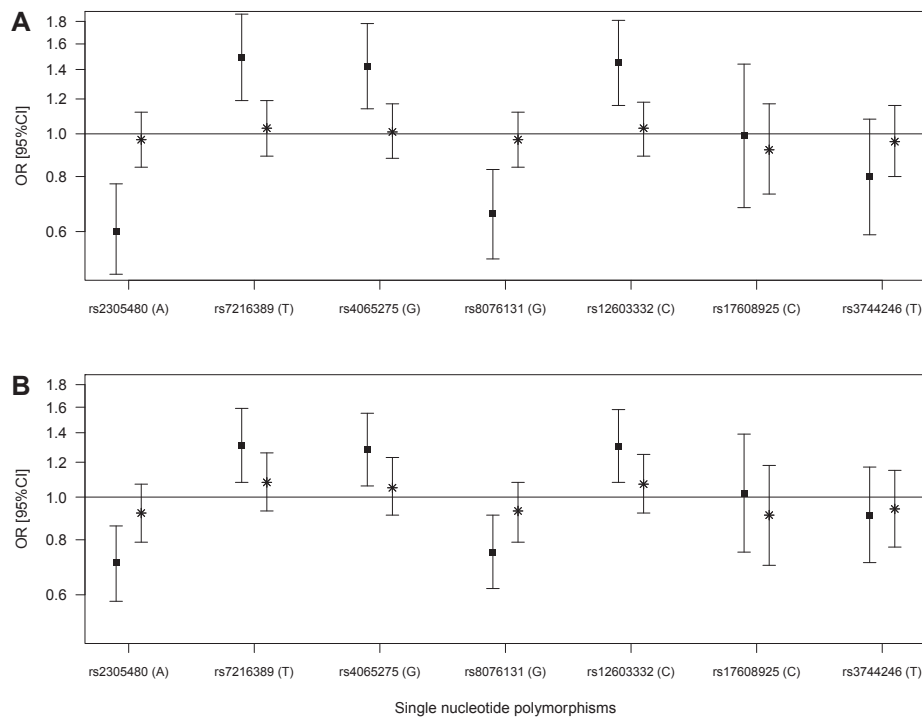


FIG 1. Pooled longitudinal associations between the 17q21 SNPs and allergic rhinitis, stratified by early-life (3-8 years) (A) and ever (B) asthma (black squares = yes; stars = no). Models were adjusted for age and cohort and only included cohorts with data on early-life asthma (BAMSE, GINI/LISA, and PIAMA). Controls were defined as children with no allergic rhinitis or asthma.

estimates for allergic rhinitis without concomitant asthma appeared smaller but were also significant/borderline significant for 5 SNPs in high linkage disequilibrium. Associations with allergic rhinitis were similar after adjustment for asthma during early life (3-8 years). However, stratified analyses indicated that associations between the SNPs and allergic rhinitis were significant only among those with a history of asthma (Fig 1).

In our study, risk estimates for the 17q21 locus with concomitant allergic rhinitis and asthma appeared largest, which is

intuitive given that significant associations were observed for asthma and allergic rhinitis modeled independently. These results have been observed by others² and may suggest that the 17q21 locus is involved in the development of multiple clinical manifestations or a more severe type of disease, possibly via a causal pathway involving asthma. An asthma-dependent mechanism is supported by the stratified analyses in which associations between the SNPs and allergic rhinitis were most pronounced for those with a history of asthma. Furthermore, the

17q21 locus appears to be a stronger risk factor for asthma than for allergic rhinitis.² Nevertheless, because weaker borderline significant longitudinal associations for allergic rhinitis without concomitant asthma were also observed, we are unable to exclude the possibility that any effect of 17q21 variation on allergic rhinitis may also be mediated through an asthma-independent pathway, as has been suggested by others.³ However, the null associations with allergic rhinitis for those without any history of asthma observed in this study do not support this hypothesis (Fig 1).

Allele-specific differences in gene expression at 17q21 are being investigated.^{7,8} However, how these genes (and other genetic, epigenetic, and gene-environment influences) may affect asthma or allergic rhinitis development remains under study. A functional role for ORM1-like protein 3 in regulating eosinophil trafficking, recruitment, and degranulation via the regulation of integrins and CD48 was recently identified.⁹ Because eosinophils are involved in immune and inflammatory responses to allergens, this may represent a possible mechanism by which 17q21 variation affects allergic asthma and allergic rhinitis.

Our study results are based on a substantially larger sample size and longer follow-up than any previous candidate-gene study for allergic rhinitis studying this region. It is noteworthy that the cohort-specific analyses, although consistent in trend, did not independently reach statistical significance in almost all cases. This highlights the need to carefully pool or meta-analyze homogenous data to achieve sufficient statistical power. It nevertheless remains possible that our study may have had less power to detect associations that were found to be without or with borderline significance. The SNPs studied were limited to those available for the largest subset of the cohorts, which included top hits from previous asthma GWASs.¹ It is reassuring that we observed significant associations for 6 of the 7 SNPs (which were in high linkage disequilibrium) and that associations were consistent across cohorts despite slightly different outcome definitions (doctor diagnoses [CAPPS, GINI/LISA, and SAGE], rhinitis symptom reports [PIAMA], or both [BAMSE]). This consistency also argues against the interpretation that the results are due to chance. Furthermore, associations between allergic rhinitis and the 17q21 locus would have remained significant after a very conservative Bonferroni correction (Table I).

In summary, genetic variants at the 17q21 locus were significantly associated with allergic rhinitis, primarily with concomitant asthma, in a pooled longitudinal analysis of 5 birth cohorts. These results support the hypothesis of a shared genetic susceptibility between asthma and allergic rhinitis.

We thank all the children and parents for their cooperation, and all technical and administrative support staff and medical and fieldwork teams. We acknowledge the Mutation Analysis Core Facility at Karolinska Institutet for genotyping in BAMSE, Dr I. M. Nolte and Dr O. Savenije for their contribution to the PIAMA imputation, and Denise Daley and the AllerGen Genetics team for assistance with CAPPS and SAGE data management and transfer.

Elaine Fuertes, MSc^{a,b}
Cilla Söderhäll, PhD^c
Nathalie Acevedo, MD, MSc^{c,d}
Allan Becker, MD^e
Michael Brauer, ScD^{a,f}

Moira Chan-Yeung, MB^f
F. Nicole Dijk, MD^g
Joachim Heinrich, PhD^b
Johan de Jongste, MD, PhD^h
Gerard H. Koppelman, MD, PhD^g
Dirkje S. Postma, MD, PhDⁱ
Juha Kere, MD, PhD^{c,j,k}
Anita L. Kozyrskyy, PhD^l
Göran Pershagen, MD, PhD^m
Andrew Sandford, PhD^f
Marie Standl, PhD^b
Carla M. T. Tiesler, MSc^{b,n}
Melanie Waldenberger, PhD^{o,p}
Marit Westman, MD^q
Christopher Carlsten, MD, MPH^{a,f}
Erik Melén, MD, PhD^{m,r}

From ^athe School of Population and Public Health, University of British Columbia, Vancouver, British Columbia, Canada; ^bthe Institute of Epidemiology I, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany; ^cthe Department of Biosciences and Nutrition, and Center for Innovative Medicine, Karolinska Institutet, Huddinge, Sweden; ^dthe Translational Immunology Unit, Department of Medicine Solna, Karolinska Institutet and University Hospital, Stockholm, Sweden; ^ethe Department of Pediatrics and Child Health, University of Manitoba, Winnipeg, Manitoba, Canada; ^fthe Department of Medicine, University of British Columbia, Vancouver, British Columbia, Canada; ^gthe Department of Pediatric Pulmonology and Pediatric Allergology, Beatrix Children's Hospital, Groningen Research Institute for Asthma and COPD, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; ^hthe Department of Pediatrics/Respiratory Medicine, Erasmus MC - Sophia Children's Hospital, Rotterdam, The Netherlands; ⁱthe Department of Pulmonology, Groningen Research Institute for Asthma and COPD, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; ^jFolkhälsan Institute of Genetics, University of Helsinki, Helsinki, Finland; ^kMolecular Neurology Research Program, University of Helsinki, Helsinki, Finland; ^lthe School of Public Health, University of Alberta, Edmonton, Alberta, Canada; ^mthe Institute of Environmental Medicine and Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; ⁿthe Division of Metabolic Diseases and Nutritional Medicine, Ludwig-Maximilians - University of Munich, Dr von Hauner Children's Hospital, Munich, Germany; ^othe Research Unit of Molecular Epidemiology, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany; ^pthe Institute of Epidemiology II, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany; ^qthe Division of Ear, Nose and Throat Diseases, Department of Clinical Science, Intervention and Technology, Karolinska Institutet, Karolinska University Hospital, Stockholm, Sweden; and ^rSach's Children's Hospital, Stockholm, Sweden. E-mail: erik.melen@ki.se.

The BAMSE study was supported by the Swedish Research Council, the Swedish Heart-Lung Foundation, Stiftelsen Frimurare Barnhuset i Stockholm, Matsumura's donation, the Stockholm County Council, the Swedish Environmental Protection Agency, the Swedish Society for Medical Research, and the Swedish Foundation for Strategic Research. The PIAMA study is supported by The Netherlands Organization for Health Research and Development, The Netherlands Organization for Scientific Research, The Netherlands Asthma Fund, The Netherlands Ministry of Spatial Planning, Housing, and the Environment, and The Netherlands Ministry of Health, Welfare and Sport. Part of the genotyping of PIAMA was supported by BBMRI-NL (grant no. CP2010-29) and the GABRIEL study (FP6 grant, European Commission, 018996). The GINIplus study was supported for the first 3 years by the Federal Ministry for Education, Science, Research and Technology (intervention arm) and Helmholtz Zentrum Munich (former Gesellschaft für Strahlenforschung [GSF]; observation arm). The 4-, 6-, and 10-year follow-up examinations of the Munich study center were covered by the Helmholtz Zentrum Munich (former GSF), Ludwig-Maximilians-Universität Munich, and Technische Universität Munich. The LISApplus study (Munich study center) was supported by the Federal Ministry for Education, Science, Research and Technology and the Helmholtz Zentrum Munich (former GSF). The imputation for GINIplus and LISApplus was supported by the Munich Center of Health Sciences as part of the Ludwig-Maximilians University Munich LMU initiative. The CAPPS was supported by the Canadian Institutes of Health Research, the British Columbia Lung Association, and the Manitoba Medical Service Foundation. The SAGE was supported by the Canadian Institutes of Health Research. The "Traffic Asthma and Genetics" collaboration was supported by the AllerGen Networks of Centres of Excellence. E. Fuertes was supported by the AllerGen Networks

of Centres of Excellence (Canadian Allergy and Immune Diseases Advanced Training Initiative) and the Canadian Institutes of Health Research (Sir Frederick Banting and Charles Best Canada Graduate Scholarship and Michael Smith Foreign Study Supplement Award). The aforementioned funding sources had no involvement in the study design, in the collection, analysis, and interpretation of data, in the writing of the report, and in the decision to submit the article for publication.

Disclosure of potential conflict of interest: D. S. Postma's institution has received consultancy fees from AstraZeneca, Chiesi, Boehringer Ingelheim, GlaxoSmithKline, TEVA, and Takeda; she has received or has grants pending from Chiesi, from which she has also received payment for delivering lectures. G. Pershagen's institution has received funding from the Swedish Research Council and the Swedish Environmental Protection Agency. M. Brauer's institution has received funding from AllerGen Networks of Centres of Excellence, as has E. Fuentes, who has also received funding from the Canadian Institutes of Health Research. G. H. Koppelman's institution has received funding from the Dutch Lung Foundation. The rest of the authors declare that they have no relevant conflicts of interest.

REFERENCES

- Moffatt MF, Kabesch M, Liang L, Dixon AL, Strachan D, Heath S, et al. Genetic variants regulating ORMDL3 expression contribute to the risk of childhood asthma. *Nature* 2007;448:470-3.
- Ferreira MAR, Matheson MC, Tang CS, Granell R, Ang W, Hui J, et al. Genome-wide association analysis identifies 11 risk variants associated with the asthma with hay fever phenotype. *J Allergy Clin Immunol* 2014;133:1564-71.
- Tomita K, Sakashita M, Hirota T, Tanaka S, Masuyama K, Yamada T, et al. Variants in the 17q21 asthma susceptibility locus are associated with allergic rhinitis in the Japanese population. *Allergy* 2013;68:92-100.
- Andiappan AK, Wang DY, Anantharaman R, Parate PN, Suri BK, Low HQ, et al. Genome-wide association study for atopy and allergic rhinitis in a Singapore Chinese population. *PLoS One* 2011;6:e19719.
- Ramasamy A, Curjuric I, Coin LJ, Kumar A, McArdle WL, Imboden M, et al. A genome-wide meta-analysis of genetic variants associated with allergic rhinitis and grass sensitization and their interaction with birth order. *J Allergy Clin Immunol* 2011;128:996-1005.
- Hinds DA, McMahon G, Kiefer AK, Do CB, Eriksson N, Evans DM, et al. A genome-wide association meta-analysis of self-reported allergy identifies shared and allergy-specific susceptibility loci. *Nat Genet* 2013;45:907-11.
- Verlaan DJ, Berlivet S, Hunninghake GM, Madore A-M, Larivière M, Moussette S, et al. Allele-specific chromatin remodeling in the ZPBP2/GSDMB/ORMDL3 locus associated with the risk of asthma and autoimmune disease. *Am J Hum Genet* 2009;85:377-93.
- Hao K, Bossé Y, Nickle DC, Paré PD, Postma DS, Laviolette M, et al. Lung eQTLs to help reveal the molecular underpinnings of asthma. *PLoS Genet* 2012;8:e1003029.
- Ha SG, Ge XN, Bahaie NS, Kang BN, Rao A, Rao SP, et al. ORMDL3 promotes eosinophil trafficking and activation via regulation of integrins and CD48. *Nat Commun* 2013;4:2479.

Available online September 26, 2014.
<http://dx.doi.org/10.1016/j.jaci.2014.08.016>

Increased noneosinophilic nasal polyps in chronic rhinosinusitis in US second-generation Asians suggest genetic regulation of eosinophilia

To the Editor:

Chronic rhinosinusitis (CRS) with nasal polyps (CRSwNP) is an inflammatory disease with a high impact on quality of life and is characterized by the presence of polyps in an individual with diagnosis of CRS. Although most of the nasal polyps (NPs) in patients with CRSwNP seen in Western countries are eosinophilic,^{1,2} noneosinophilic NPs comprise a significant percentage of CRSwNP seen in East Asian countries including China, Korea, Japan, and Malaysia.³⁻⁸ Studies comparing polyps in Chinese and Belgian patients have shown significantly lower eosinophil numbers^{3,9} and lower eosinophil cationic protein (ECP) levels⁹ in NPs in Chinese patients.

In the last decade, there has been a trend toward increasing eosinophilic nasal polyposis in Asian populations.^{5,10} A Korean study has shown that eosinophilic NPs have increased from

24% of the total NPs resected in 1993-1994 to 50.9% in 2010-2012.⁵ However, recent studies still show that about half of the CRSwNP cases in East Asian countries have a noneosinophilic pathology.⁴⁻⁸ This raises the possibility that this type of NPs are influenced by different pathogenesis elements that are more common in that area and/or that genetic factors play a role in the level of eosinophilia in NPs in Asian populations.

In an attempt to test whether the propensity of Asian populations to manifest noneosinophilic polyps is due to genetic factors, we evaluated the eosinophilic marker ECP and evidence for eosinophilia in the pathology report of polyp and sinus tissue collected during functional endoscopic surgery in a group of second-generation Asian patients with CRS in Illinois and compared them with those in patients of other ethnicities in Illinois. We focused on study patients with both parents from Asian countries who were born and raised in the United States. Such patients come from the same genetic pool as their parents but have presumably been exposed to environmental factors present in the United States. Our findings demonstrated reduced eosinophilia in Asian patients with CRSwNP who were born and raised in the United States.

A consecutive series of 296 patients with CRSwNP who underwent surgery at Northwestern University from 2005 to 2013 were included. Patients with self-reported Asian ancestry and born in the United States were identified as second-generation Asian. Twenty-three patients had identified themselves as Asian (including Chinese, Korean, Japanese, and Malaysian), out of which 11 were second generation and were included. The rest were excluded from the study. Original pathology reports of polyp and sinus tissue obtained during surgery were reviewed for reported eosinophilia. The tissue would have been reported as eosinophilic by the pathologists if eosinophils comprised more than 10% of inflammatory cells in the studied area. Furthermore, in 161 of the enrolled cases, tissue homogenates of polyp and/or uncinat tissue (UT) were available in a prospectively collected biorepository and were analyzed for ECP. UT was used as representative of sinus and upper nasal tissue. Previous studies have shown that NPs and UT of patients with CRSwNP from the United States have a significantly higher number of eosinophils than does control UT.² ECP levels were measured by ELISA using the Mesacup ECP Test Kit from MBL (Woburn, Mass). The ECP concentration was measured as ng/ μ L. The level was adjusted to total protein concentration in each sample, which was measured as mg/ μ L. All ECP levels were reported as ng/mg of total protein. Physician diagnosis of asthma and atopy was recorded as part of the study and was double-checked by using chart review. Atopy was defined by evidence of allergic sensitization to aeroallergens by using skin prick test or Immunocap testing. The study was approved by the Northwestern University institutional review board.

Comparisons among groups in terms of ECP levels were assessed by using Kruskal-Wallis and Dunn's multiple comparisons tests. Comparisons between groups in terms of eosinophilic polyps, asthma, and atopy were done by using Fisher exact test. Adjusting for age and sex for the above analyses was performed by logistic regression. These statistical analyses were performed using IBM SPSS, version 22. A *P* value of less than .05 was considered statistically significant.

Levels of ECP in polyp (mean, 179.4 vs 1256 ng/mg; *P* < .001) and UT samples (mean, 133.6 vs 838.6 ng/mg; *P* < .001) were significantly lower in tissue from second-generation Asian versus

METHODS

Outcome definitions

The cohort-specific rhinitis and asthma definitions, aeroallergens tested, and frequencies of follow-up are provided in Table E1. Parent-completed questionnaires were used to collect data on rhinitis and asthma. Aeroallergen sensitization was assessed as any specific IgE level of 0.35 kU/L or more for BASME, GINI/LISA, and PIAMA. For CAPPs and SAGE, sensitization was assessed by using skin prick testing. A positive reaction was defined as having a wheal diameter of 3 mm or more. Although not all cohorts had information on all aeroallergens, several aeroallergens were tested in each cohort (Table E1). Each cohort received ethical approval from its local authorized institutional review board. Statistical analyses were conducted in R, version 2.13.1 (www.r-project.org).

Genetic analyses

The 7 SNPs examined in this study were in Hardy-Weinberg equilibrium ($P > .01$). For BAMSE, these SNPs were genotyped in a subset of the cohort ($N = 2033$ with DNA extracted from blood samples available) by Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (SEQUENOM, Inc, San Diego, Calif). For the CAPPs, GINI/LISA, PIAMA, and SAGE cohorts, information on these 7 SNPs was extracted from imputed genome-wide data. All imputed allele dosage data were recoded to hard coding (0, 1, and 2) before further analysis. As a sensitivity analysis, the longitudinal model results for the GINI/LISA cohort were compared using hard and allele dosage coding for the SNPs and the results were very consistent.

For GINI/LISA, DNA samples from 1027 children and 69 children were analyzed using the Affymetrix Human SNP Arrays 5.0 and 6.0, respectively. Genotypes were called using the BRLMM-P (Affymetrix 5.0) or BIRDSEED V2 algorithm (Affymetrix 6.0). The genotype data were subjected to quality control filters on the variant and individual levels. Variants were excluded on the basis of the following criteria: a call rate below 95%, a minor allele frequency below 1%, or a Hardy-Weinberg equilibrium P value below 1×10^{-5} (~18% excluded). Only those individuals who had a call rate above 95%, a heterozygosity value within ± 4 SDs of the mean, and who passed a sex check and the similarity quality control step based on multidimensional scaling plots were retained for subsequent imputation. The genotype data were prephased using SHAPEIT v2^{E1,E2} and imputed using IMPUTE v2^{E3} against reference haplotypes from the 1000 Genomes Project (Phase I integrated variant set [v3; includes individuals from all ancestries]; March 2012, updated August 26, 2012; limited to variants with more than 1 minor allele copy). All 7 SNPs used for the present analysis were imputed with good imputation quality (IMPUTE v2 INFO > 0.94).

For PIAMA, DNA was extracted from blood or buccal swabs. DNA of 1377 children was genotyped on the Illumina Omni Express Exome Chip and DNA of 288 children was genotyped with the Omni Express chip, both at the Genomics Facility of the University Medical Center Groningen. DNA of 404 children was genotyped at the Centre National de Genotypage (CNG, Evry, France) as part of the GABRIEL consortium.^{E4} SNPs were harmonized by base-pair position annotated to genome build 37, name, and annotation of strand for each platform. Discordant or duplicate SNPs or SNPs that showed large differences in allele frequencies (>15%) were removed. After quality

control, a total of 1968 individuals remained and imputation was performed per platform using IMPUTE 2.0 against the reference data set of the CEU panel of the 1000 Genomes project (version March 2012). SNPs of high quality (info-score IMPUTE ≥ 0.7) were merged into 1 data set using GTOOL and used for further analysis.

For CAPPs and SAGE, data on only 4 SNPs (rs2305480, rs7216389, rs12603332, and rs3744246) were available. In total, 956 samples from parents and children were genotyped using the Illumina HumanHap550 SNP Array. Genotypes were subjected to the following quality control filters for variants and individual samples. Variants were excluded on the basis of the following criteria: a call rate below 95%, a minor allele frequency below 1%, failure of Hardy-Weinberg equilibrium P value below 1×10^{-4} , and SNPs with more than 2 Mendelian errors. Individual samples were retained if they had a call rate of more than 97% and a heterozygosity value within ± 3 SDs of the mean, and samples additionally had to pass gender and Mendelian transmission error checks. Multidimensional scaling was used to check for population stratification and to identify monozygotic twins ($n = 2$) and duplicate samples. Imputation was performed using MaCH^{E5} and HapMap2 r22 reference population. SNPs needed to be imputed with an $r^2 \geq 0.99$ to be used in the analysis.

REFERENCES

- E1. Delaneau O, Marchini J, Zagury J-F. A linear complexity phasing method for thousands of genomes. *Nat Methods* 2012;9:179-81.
- E2. Delaneau O, Zagury J-F, Marchini J. Improved whole-chromosome phasing for disease and population genetic studies. *Nat Methods* 2013;10:5-6.
- E3. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet* 2009;5:e1000529.
- E4. Moffatt M, Gut I, Demenais F, Strachan D, Bouzigon E, Heath S, et al. A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med* 2010;363:1211-21.
- E5. Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet Epidemiol* 2010;34:816-34.
- E6. Wickman M, Pershagen G, Nordvall SL. The BAMSE project: presentation of a prospective longitudinal birth cohort study. *Pediatr Allergy Immunol* 2002;13:11-3.
- E7. Chan-Yeung M, Manfreda J, Dimich-Ward H, Ferguson A, Watson W, Becker A. A randomized controlled study on the effectiveness of a multifaceted intervention program in the primary prevention of asthma in high-risk infants. *Arch Pediatr Adolesc Med* 2000;154:657-63.
- E8. Filipiak B, Zutavern A, Koletzko S, von Berg A, Brockow I, Gröbl A, et al. Solid food introduction in relation to eczema: results from a four-year prospective birth cohort study. *J Pediatr* 2007;151:352-8.
- E9. Heinrich J, Bolte G, Hölscher B, Douwes J, Lehmann I, Fahlbusch B, et al. Allergens and endotoxin on mothers' mattresses and total immunoglobulin E in cord blood of neonates. *Eur Respir J* 2002;20:617-23.
- E10. Brunekreef B, Smit J, de Jongste J, Neijens H, Gerritsen J, Postma D, et al. The prevention and incidence of asthma and mite allergy (PIAMA) birth cohort study: design and first results. *Pediatr Allergy Immunol* 2002;13:55-60.
- E11. Kozyrskij AL, HayGlass KT, Sandford A, Pare PD, Chan-Yeung M, Becker AB. A novel study design to investigate the early-life origins of asthma in children (SAGE study). *Allergy* 2009;64:1185-93.

TABLE E1. Characteristics of participating cohorts

Cohort (country)	Full name of cohort	Study type	Recruitment	Sample size*	Ages (y) available†	Rhinitis definition	Asthma definition	Aeroallergens tested
BAMSE (Sweden)	Children, Allergy, Milieu, Stockholm, Epidemiological Survey ^{E6}	Population-based birth cohort with wheeze nested case-control	1994-1996	1953	4, 8, 16	Symptoms (sneezing, runny or blocked nose, itchy, red, and watery eyes) after exposure to furred pets or pollen or a medical diagnosis of allergic rhinitis since previous questionnaire	At least 4 episodes of wheeze in the last 12 mo or at least 1 episode of wheeze with occasional or regular use of prescribed inhaled corticosteroids in the last 12 mo	Birch, cat, dog, house dust mite (<i>Dermatophagoide s pteronyssinus</i>), mold (<i>Cladosporium herbarum</i>), mugwort, timothy grass
CAPPS (Canada)	Canadian Asthma Primary Prevention Study ^{E7}	Randomized controlled study with asthma intervention	1995	80	7	Medical diagnosis of allergic rhinitis assessed at 7-y follow-up	Medical diagnosis of asthma assessed at 7-y follow-up	<i>Alternaria</i> , cat, cockroaches, dog, feathers, grass, house dust mites, mold (<i>C herbarum</i>), ragweed, trees, weeds
GINIplus (Germany)‡	German Infant study on the influence of Nutritional Intervention plus environmental and genetic influences on allergy development ^{E8}	Population-based birth cohort. Subset for nutritional intervention	1995-1998	612	6, 10	Medical diagnosis of allergic rhinitis or hay fever during the last 12 mo	Medical diagnosis of asthma during the last 12 mo	Birch, cat, dog, house dust mite (<i>D pteronyssinus</i>), mold (<i>Cladosporium herbarum</i>), mugwort, rye, timothy grass
LISApplus (Germany)‡	Lifestyle-related factors, Immune System and the development of Allergies in East and West Germany plus the influence of traffic emissions and genetics study ^{E9}	Population-based birth cohort	1997-1999	484	6, 10	Medical diagnosis of allergic rhinitis or hay fever during the last 12 mo	Medical diagnosis of asthma during the last 12 mo	Birch, cat, dog, house dust mite (<i>D pteronyssinus</i>), mold (<i>C herbarum</i>), mugwort, rye, timothy grass
PIAMA (The Netherlands)	Prevention and Incidence of Asthma and Mite Allergy ^{E10}	Population-based birth cohort. Subset for mattress cover intervention	1996-1997	1386	4, 8, 11/12	Sneezing, runny/blocked nose during the last 12 mo without cold or flu	Medical diagnosis of asthma during the last 12 mo	<i>Alternaria</i> ,§ birch, cat, <i>Dactylis</i> , dog,§ house dust mite (<i>D pteronyssinus</i>)
SAGE (Canada)	Study of Asthma Genes and the Environment ^{E11}	Population-based cohort with asthma nested case-control	1995	109	8	Medical diagnosis of allergic rhinitis assessed at 8-y follow-up	Medical diagnosis of asthma assessed at 8-y follow-up	Cat, dog, feathers, grass, ragweed, trees, weeds

*Number of children with health outcome information for at least 1 time point and genotype data.

†Ages at which both rhinitis and aeroallergen sensitization data were available, and therefore allergic rhinitis could be defined.

‡Data from only the Munich center from these 2 combined German cohorts were included in the current analysis, referred to as GINI/LISA throughout the article, because genome-wide genetic data were available only for that study area (Wesel, Leipzig, and Bad Honnef were excluded).

§Available only at ages 4 and 8 y.

TABLE E2. Pooled and cohort-specific numbers of cases and controls (cases/controls)

Age (y)	Pooled	BAMSE	CAPPS	GINI/LISA	PIAMA	SAGE
Allergic rhinitis*						
4	152/1675	102/1319	-	-	50/356	-
6	80/791	-	-	80/791	-	-
7	22/58	-	22/58	-	-	-
8	437/2398	253/1426	-	-	159/888	25/84
10	125/593	-	-	125/593	-	-
11/12	154/505	-	-	-	154/505	-
16	411/828	411/828	-	-	-	-
Allergic rhinitis with concomitant asthma†						
4	37/1594	28/1246	-	-	9/348	-
6	12/780	-	-	12/780	-	-
7	9/53	-	9/53	-	-	-
8	115/2313	63/1367	-	-	37/876	15/70
10	25/585	-	-	25/585	-	-
11/12	25/502	-	-	-	25/502	-
16	69/797	69/797	-	-	-	-
Allergic rhinitis without concomitant asthma†						
4	114/1594	73/1246	-	-	41/348	-
6	68/780	-	-	68/780	-	-
7	13/53	-	13/53	-	-	-
8	318/2313	186/1367	-	-	122/876	10/70
10	98/585	-	-	98/585	-	-
11/12	129/502	-	-	-	129/502	-
16	333/797	333/797	-	-	-	-

*Age-specific controls were defined as those without allergic rhinitis.

†Age-specific controls were defined as those without allergic rhinitis or asthma.

TABLE E3. Genetic information for the 7 SNPs at the 17q21 locus

SNP	Gene	Noneffect allele/effect allele	Position (build 37)	BAMSE*		CAPPS†		GINI/LISA†		PIAMA†		SAGE†	
				N	EAf	N	EAf	N	EAf	N	EAf	N	EAf
rs2305480	<i>GSDMB</i>	G/A	38062196	1934	0.47	80	0.47	1096	0.45	1386	0.44	109	0.40
rs7216389	<i>GSDMB</i>	C/T	38069949	1933	0.48	80	0.48	1096	0.49	1386	0.51	109	0.54
rs4065275	<i>ORMDL3</i>	A/G	38080865	1939	0.50	-	-	1096	0.50	1386	0.52	-	-
rs8076131	<i>ORMDL3</i>	A/G	38080912	1935	0.47	-	-	1096	0.45	1386	0.44	-	-
rs12603332	<i>ORMDL3</i>	T/C	38082807	1948	0.49	79	0.50	1096	0.50	1386	0.52	109	0.56
rs17608925	<i>ORMDL3</i>	T/C	38082831	1932	0.10	-	-	1096	0.11	1386	0.11	-	-
rs3744246	<i>ORMDL3</i>	C/T	38084350	1936	0.19	80	0.19	1096	0.21	1386	0.20	109	0.20

The SNPs are in linkage disequilibrium ($r^2 > 0.2$, $D' > 0.9$).

EAf, Effect allele frequency; *GSDMB*, gasdermin B; N, number of children with health outcome information for at least 1 time point and genotype data; *ORMDL3*, ORM1-like protein 3.

*Genotypes derived from genotyped data.

†Genotypes derived from imputed data.

TABLE E4. Cohort-specific associations between 7 SNPs at the 17q21 locus and allergic rhinitis*

SNP (effect allele)	BAMSE (up to 16 y)		CAPPS (at 7 y)		GINI/LISA (up to 10 y)		PIAMA (up to 12 y)		SAGE (at 8 y)	
	N	OR (95% CI)	N	OR (95% CI)	N	OR (95% CI)	N	OR (95% CI)	N	OR (95% CI)
rs2305480 (A)	1934	0.87 (0.75-1.00)	80	1.05 (0.52-2.13)	1096	0.80 (0.64-1.01)	1386	0.87 (0.72-1.06)	109	0.71 (0.36-1.40)
rs7216389 (T)	1933	1.15 (1.00-1.33)	80	0.98 (0.46-2.05)	1096	1.16 (0.93-1.44)	1386	1.14 (0.94-1.38)	109	1.25 (0.64-2.42)
rs4065275 (G)	1939	1.15 (1.00-1.33)†	-	-	1096	1.09 (0.87-1.36)	1386	1.13 (0.93-1.38)	-	-
rs8076131 (G)	1935	0.87 (0.76-1.01)	-	-	1096	0.81 (0.65-1.02)	1386	0.87 (0.71-1.06)	-	-
rs12603332 (C)	1948	1.18 (1.03-1.36)†	79	1.00 (0.47-2.12)	1096	1.09 (0.87-1.35)	1386	1.13 (0.93-1.37)	109	1.15 (0.60-2.22)
rs17608925 (C)	1932	0.97 (0.77-1.22)	-	-	1096	0.79 (0.52-1.20)	1386	0.83 (0.60-1.15)	-	-
rs3744246 (T)	1936	0.91 (0.76-1.08)	80	1.16 (0.49-2.76)	1096	0.88 (0.65-1.18)	1386	0.78 (0.61-1.00)	109	0.54 (0.21-1.40)

N, Number of children included in the model; OR, odds ratio.

*Models were adjusted for age except for CAPPS and SAGE for which only 1 time point was available. The SNPs are in linkage disequilibrium ($r^2 > 0.2$, $D' > 0.9$).

† $P < .05$.