



Forest fires in Canadian permafrost region: the combined effects of fire and permafrost dynamics on soil organic matter quality

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Abstract Wildfires burn approximately 1% of boreal forest yearly, being one of the most significant factors affecting soil organic matter (SOM) pools. Boreal forests are largely situated in the permafrost zone, which contains half of global soil carbon (C). Wildfires advance thawing of permafrost by burning the insulating organic layer and decreasing surface albedo, thus increasing soil temperatures. Fires also affect SOM quality through chemical and physical changes, such as the formation of resistant C

compounds. The long-term post-fire effects on SOM quality, degradability and isotopic composition are not well known in permafrost forests. We studied the effect of forest fires on the proportional sizes of SOM pools with chemical fractionation (extracting with water, ethanol and acid) of soil samples (5, 30 and 50 cm depths) collected from a fire chronosequence in the upland mineral soils of the Canadian permafrost zone. We also determined the ^{13}C and ^{15}N isotopic composition of soil after fire. In the topsoil horizon (5 cm) recent fire areas contained a smaller fraction of labile SOM and were slightly more enriched with ^{15}N and ^{13}C than older fire areas. The SOM fraction ratios reverted towards pre-fire status with succession. Changes in SOM were less apparent deeper in the soil. Best predictors for the size of recalcitrant SOM

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fraction were active layer depth, vegetation biomass and soil C/N ratio, whereas microbial biomass was best predicted by the size of the recalcitrant SOM fraction. Results indicated that SOM in upland mineral soils at the permafrost surface could be mainly recalcitrant and its decomposition not particularly sensitive to changes resulting from fire.

Keywords Chemical fractionation · Microbial biomass · Wildfire

Introduction

Forest fires burn 5–20 M ha of boreal forest each year (Stocks et al. 2004; Sukhinin et al. 2004; Tan et al. 2007; Flannigan et al. 2009) and as much as 80% of the boreal forest region is located in the circumpolar permafrost zone (Helbig et al. 2016). Around half of the global soil carbon (C) pool resides in the permafrost soils of the Northern hemisphere and yet one-fourth of this permafrost is predicted to thaw by 2100 (Davidson and Janssens 2006). Thawing may be accelerated in multiple ways by fires; they not only release heat, but also may burn the insulating organic layer, leading to increased soil temperatures and, therefore, an increase in the active layer depth (Jorgenson et al. 2010; Bret-Harte et al. 2013; Brown et al. 2015; Gibson et al. 2018). Additionally, the albedo of the soil surface may change due to the destruction of vegetation and darkening of the soil surface (Bret-Harte et al. 2013; French et al. 2016). All of these factors can lead to further warming of the subsoil and thawing of permafrost (Yoshikawa et al. 2002; Randerson et al. 2006; Gibson et al. 2018). Thawing of permafrost means that soil organic matter (SOM) that was previously protected by subzero temperatures, becomes susceptible to decomposition (Dutta et al. 2006; Harden et al. 2012; Walz et al. 2017). The degree of SOM decomposition following permafrost thaw, and the resulting CO₂ emissions, are still under discussion (Waldrop et al. 2010), and depend on many factors, such as soil hydrology (Trumbore and Harden 1997; Lawrence et al. 2015). The thawing permafrost has been expected to cause significant positive feedback to climate change through greenhouse gases released from decomposing, previously frozen organic matter (Schädel et al. 2016).

On the other hand, in forest areas, the succession of vegetation after a fire has been shown to protect the permafrost, even when air temperatures increase (Jorgenson et al. 2010).

Especially stand-replacing forest fires can produce large amounts of partly burnt organic material, which becomes incorporated to SOM (González-Pérez et al. 2004; López-Martín et al. 2016). This, together with the burning of the organic layer, may affect the soil microbial communities (Mataix-Solera et al. 2009). In general, forest fires decrease microbial biomass (Dooley and Treseder 2012; Köster et al. 2016), as even relatively low soil temperatures (< 100 °C) during the fire may be fatal to microbes (Certini 2005). Further, if the fire causes soil hydrophobicity, the soil may dry, further diminishing the microbial activity (DeBano 2000). The loss of SOM, in turn, can decrease the nutrient availability and microbial growth (Wan et al. 2001), which can lead to diminished decomposition of SOM in burnt forest areas (DeBano and Conrad 1978; Certini et al. 2011). Conversely, a recent study from the Mediterranean has observed increased bacterial and fungal biomasses in burnt soils (Rodríguez et al. 2017, 2018).

Fire also affects physical and chemical properties of soil (Certini 2005; López-Martín et al. 2016). Burning causes significant C (Certini et al. 2011; De la Rosa et al. 2013) and nitrogen (N) loss from soil (DeBano and Conrad 1978; Vitousek and Howarth 1991). During or after the fire the quality of SOM may decrease since the roots, and exudates emitted from them, are possibly removed and previously labile SOM may change its form (Waldrop et al. 2010; Schädel et al. 2016). This is also reflected in the soil C/N-ratio, which is usually reduced after fire (Almendros and González-Vila 2012). Possible changes that SOM may undergo during the fire are dependent on the chemical and physical properties of the soil, as well as the intensity of the fire (González-Pérez et al. 2004; Knicker 2007). For example, very high temperature leads to the production of black carbon (Waldrop et al. 2010; Almendros and González-Vila 2012). Black carbon usually has a long residence time in soil (Krull et al. 2006) and can therefore act as a C sink (Czimczik et al. 2003). It may also indirectly facilitate degradation of labile C by acting as a habitat for microbes (Wardle et al. 2008).

During forest fires, soil temperatures at the soil surface usually vary between 200 and 300 °C, and

even low temperatures of 40–70 °C cause degradation of biological material (Knicker 2007). Temperatures reaching 300 °C may cause structural changes in SOM, resulting in the formation of pyrogenic compounds (Almendros and Leal 1990; Knicker et al. 1996). Macromolecules, such as cellulose and lignin, may go through transformations during fire even at lower temperatures, leading to the formation of compounds, which are more recalcitrant and thus become unrecognisable to certain enzymes (Almendros and González-Vila 2012). Furthermore, chemical changes in SOM functional groups may lead to hydrophobicity or stronger binding of SOM to mineral particles (Faria et al. 2012). This means that post-fire a certain amount of SOM becomes protected from microbial degradation (Faria et al. 2012). In summary, there could be significant, yet bidirectional impacts of forest fire on SOM decomposition in permafrost regions, with the overall short- and long-term impacts remaining yet unknown.

The isotopic composition of SOM may be used to describe SOM lability as processed, recalcitrant SOM, typically is enriched with heavier isotopes compared to fresh SOM (Natelhoffer and Fry 1988; Wynn 2007). The enrichment is mainly caused by microbial degradation of organic matter (Natelhoffer and Fry 1988; Ehleringer et al. 2000), but also other processes, such as differing mobility of dissolved organic compounds, may have a role in the enrichment (Ehleringer et al. 2000; Tu et al. 2004). In these processes, the heavier isotopes are left behind since they require higher activation energy in reactions (Wynn 2007). Wildfires have been found to enrich soil with ^{15}N to lighter ^{14}N (Cook 2001; Boeckx et al. 2005), which is caused by discrimination against heavier isotopes during volatilization of N (Frank et al. 2004) or nitrification process (Pardo et al. 2002). On the other hand, isotopic enrichment of soil may also be dependent on the burning of depleted litter during the fire (Högberg 1997). The effects of fire on soil $\delta^{13}\text{C}$ composition are not as consistent with studies reporting depletion, while some found no clear effect and some enrichment (Beuning and Scott 2002; Rumpel et al. 2007; Hyodo et al. 2013).

Several recent studies have focused on the degradability and quality of permafrost SOM (Dutta et al. 2006; Waldrop et al. 2010; Moni et al. 2015; Weiss et al. 2016; Walz et al. 2017). Nevertheless, studies concentrating on different SOM fractions in

permafrost areas are rare (Dutta et al. 2006; Höfle et al. 2013; Gentsch et al. 2015). To our knowledge, studies utilising chemical fractionation to study the quality of SOM in fire-affected permafrost forest soils are lacking. As forest fires and quality of SOM have significant effects on the release of C from forest covered permafrost soils, it is crucial for future predictions of possible CO_2 emissions to understand how fires change the proportions of different SOM fractions, as well as how permafrost SOM is generally divided between these fractions.

The aim of this study was to examine how the proportions of insoluble (recalcitrant) and soluble (labile) fractions in SOM change as a result of forest fire in the active layer on permafrost. We also aimed to study how the abundancies of stable isotopes $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ change along with depth and time since last fire. Our final aim was to identify the most important factors (active layer depth, soil temperature, biomass, C/N ratio) explaining the size of insoluble (recalcitrant) fractions of SOM post-fire, as well as to determine the SOM fractions most affecting the microbial biomass.

We hypothesised that the effects of fire could be seen in the composition of different chemical SOM fractions, as fire may decrease the amount of SOM, while at the same time also releasing organic matter from burning vegetation (possibly increasing the proportion of labile SOM) and on the other hand forming recalcitrant compounds (increasing the proportion of recalcitrant SOM). We also expected the changes in the proportions of the different chemical SOM fractions to be apparent along the fire chronosequence with advancing succession after fire. These changes could be observed as a larger fraction of recalcitrant SOM after the fire and comparably larger fractions of more labile SOM as succession proceeds and the effects of fire are diminishing with time. Also, we expected the fire to cause changes in the isotopic composition of SOM, as for ^{15}N there may be discrimination during volatilisation of N (Frank et al. 2004), while the effects of fire on $\delta^{13}\text{C}$ are not as clear (Beuning and Scott 2002; Rumpel et al. 2007; Hyodo et al. 2013).

Materials and methods

Study sites and sample collection

The study sites were located in the northern boreal forests of Canada (Fig. S1, Online resource), along the Dempster Highway close to Eagle Plains, Yukon (66°22'N, 136°43'W), and Tsiigehtchic, Northwest Territories (67°26'N, 133°45'W), also described in Köster et al. (2017). This area belongs to the taiga zone, with annual mean temperature and precipitation of -8.8 °C and 248 mm, respectively (Environment and Climate Change Canada 2017). The location is characterised by underlying, continuous permafrost. The permafrost area is affected by cryoturbation and soils are classified as cryosols (IUSS Working Group WRB 2014), or relict cryosols in the youngest fire area. The dominating soil texture was silt loam. The tree species in the area consist mainly of evergreen conifers, such as *Picea mariana* (Mill.) BSP and some *Picea glauca* (Moench) Voss. The ground vegetation is mostly formed by the following species: *Sphagnum* sp., *Cladonia* sp., *Cladina* sp., L., *Rhododendron groenlandicum* Oeder., *Rubus chamaemorus* L. and *Vaccinium uliginosum* L.

We established the study areas in 2015 in forest areas burnt 3 (FIRE₃), 25 (FIRE₂₅), 46 (FIRE₄₆) and over 100 years ago (FIRE₁₀₀), so that together they formed a fire chronosequence, as also described in Köster et al. (2017) (Fig. S1, Online resource). Fire areas forming the chronosequence were selected from unmanaged areas based on year of fire and reachability. These study areas were located based on GIS-maps of burnt areas in Yukon and Northwest Territories and further narrowed down during fieldwork based on finding areas with similar soil type, topography, tree species and tree diameter. Stand ages were confirmed from cored tree rings. Each fire area had three sample lines (length 150 m), with each line having three sample plots ($n = 9$) 50 m apart from each other in FIRE₃, FIRE₂₅ and FIRE₄₆. The lines were separated by a minimum of 200 m and they were placed at least 150 m away from roads to avoid any effects of roads on vegetation or permafrost depth (Gill et al. 2014). For comparison, we also established a similar 150 m long line (with three sampling plots in each) in a forest stand that had not been exposed to fire in the last 100 years. These lines were situated next to FIRE₃, FIRE₂₅ and FIRE₄₆, one line next to each area, to

represent the possible spatial geographical variation and to achieve a balanced comparison between fire areas in the analysis. These three lines together formed the area (FIRE₁₀₀) used in the analysis.

Soil samples were taken from soil pits, excavated on each sample plot, by inserting a cylinder (6 cm diameter and length) horizontally into the soil profile. Samples were collected from the depths of 5, 30 and 50 cm so that the point of sampling depth was at the center of the cylinder. Samples were then frozen until analysis. The soil temperature (Table S1, Online resource) for each sampling depth was measured from the pit excavated in each sampling plot. The active layer depth was measured simultaneously from each pit or, when deeper than 1.2 m, estimated by using a linear regression of soil temperature against the soil depths. Sampling depth of 50 cm was below permafrost layer in FIRE₄₆ and FIRE₁₀₀ areas (where also 30 cm sampling depth was just below the permafrost surface). The permafrost depths, together with other basic soil characteristics are presented in Table S1 and S2 (Online Resource).

Biomass for each area was also estimated by determining the tree and ground vegetation biomass (Table S2, Online resource) as described by Köster et al. (2017) in each plot. The ground vegetation biomass was measured from four 0.20×0.20 m² in each plot, while ground vegetation coverage was visually determined from two 0.75×0.75 m². We also measured tree characteristics from a 400 m² area for all trees over 1 m tall: stem diameter at 1.3 m height (or diameter at the base of trees shorter than 1.3 m in height), the height of a tree, crown height and crown diameter. Tree biomass was calculated following the allometric functions (Lambert et al. 2005) with a separate function for spruce (Wagner and Ter-Mikaelian 1999).

Soil chemical fractionation

SOM quality was studied by using chemical fractionation following Berg and Ekbohm (1991) and Karhu et al. (2010) in which oven dried (40 °C) soil samples were separated into water (sugars, low levels of fatty acids and protein remains), ethanol (waxes, fats), sulphuric acid hydrolysable (hemicellulose, cellulose) and non-hydrolysable (Klason lignin and humified material) fractions (Berg and McLaugherty 2003; Trofymow and Group 1998). First, the soil-extraction

solutions were sonicated for 90 min in a water bath, after which the solution was filtered through glass crucibles (Schott Duran, pore size 10 µm) and oven dried at 105 °C. Dried samples were weighed using an analytical scale, with detected mass loss indicating the water-soluble fraction. Extraction was repeated accordingly with pure ethanol. Lastly, the remaining sample was extracted in 72% sulphuric acid, whereby the acid solution was sonicated for 60 min and the solution was diluted to 2.5% and autoclaved for 60 min at 125 °C. After autoclaving the samples were filtered, rinsed with ultrapure water, and dried as in the previous extractions. Mineral soils (30 and 50 cm) were extracted the same way as the organic soils but, due to fine soil particles, the samples were centrifuged in glass tubes instead of filtering through crucibles. SOM content was determined as loss on ignition (550 °C, 3 h) and the different soil fractions are expressed as a percent off total SOM. The different SOM fractions can be connected to corresponding SOM pools characterized by their turnover times: active pool (sugars), slow pool (waxes, fats, cellulose) and very slow/stable pool (lignin) (Bot and Benites 2005).

Determination of soil characteristics

The pH and electric conductivity of the soil (Table S1, Online resource) were determined from a 1:2.5 (v:v) soil/water solution that was left to settle overnight. The measurements were performed with a pH meter (PHM210, Radiometer Analytical, France) and an electric meter (JENWAY 4010 Conductivity, TER Calibration Ltd., Wigan, UK). Further, we measured soil particle size distribution for soil classification from oven dried, sieved (< 2 mm) samples with laser diffraction (LS 230, Beckman Coulter Inc. US).

Soil C and N concentrations (Table 1) and natural isotope abundances ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) were analysed from bulk soil. Subsamples were ground to fine powder and approximately 3 mg was packed into tin capsules. The analyses were performed with an elemental analyser (FlashEA 1112 Series, Thermo Fischer Scientific, Waltham, MA) coupled to an isotope ratio mass spectrometer (IRMS, DELTA plusXP[®], Thermo Electron Fischer Scientific, Waltham, MA). The isotopic values ($^{13}\text{C}/^{12}\text{C}$ and $^{15}\text{N}/^{14}\text{N}$ ratios) are expressed as milli-Urey (mUR) in relation

to international reference standards (Peedee Belemnite for $^{13}\text{C}/^{12}\text{C}$ and air for $^{15}\text{N}/^{14}\text{N}$) determined as:

$$\delta X = [(R_s/R_{sd}) - 1] \times 1000 \quad (1)$$

where X is the isotope, R_s is the isotope ratio of the sample (heavy to light), and R_{sd} is the ratio of the international reference standard. More details on the isotope analysis are found in Figueiredo et al. (2014).

Microbial biomass carbon

We also determined the microbial biomass C following the fumigation-extraction method by Vance et al. (1987). The extracted microbial C was measured with total organic C analyzer (TOC-V CPH, Shimadzu, Kyoto, Japan) and calculated as described by Beck et al. (1997). Here the microbial biomass also includes the fungal biomass.

Statistical analyses

We used linear mixed effects model for statistical analyses of the data. The analyses were performed using R (i386 3.3.2) with the lme4 package (Bates et al. 2015) and drop1-function (Chambers and Hastie 1992). Differences in fractions and isotopes between the age classes (or soil depths) were tested with multiple comparison of means, using the Tukey's HSD post hoc test. The fixed factor was either time since fire or soil depth (depending on if the analysis was comparing time since fire or soil depths). Sampling lines were treated as true replicates (random effects). Data was tested for normality of residuals with the Shapiro–Wilk's test, and in case of percentage values, transformed with the arcsine-transformation to meet the criteria of the tests. Linear mixed effects model was also utilised to determine the factors most affecting the size of the insoluble organic fraction in the soil. The best model was selected based on the Akaike's information criterion (AIC) (Chambers and Hastie 1992). This was achieved by dropping fixed factors from the model based on their AIC-values. The same model (Eq. 2) was fitted against the insoluble SOM fraction separately for each depth. The initial model was:

$$SOM_{non} = a + bAD + cBiomass + dT + e(T \times AD) + fCN + gRDM \quad (2)$$

Table 1 Mean (\pm SE) soil organic matter carbon and nitrogen concentrations and the C/N-ratios (n = 9)

Area	Depth (cm) (cm)	C kg m ⁻²	C (%)	N (%)	C/N-ratio	Soil type
FIRE ₃	5	2.61 \pm 0.5	42.8 \pm 7.69	1.1 \pm 0.2	38.1 \pm 2.4	Organic
	30	7.74 \pm 1.2	4.80 \pm 1.51	0.3 \pm 0.1	15.4 \pm 2.4	Mineral
	50	3.92 \pm 0.3	2.90 \pm 0.60	0.2 \pm 0.0	15.9 \pm 1.3	Mineral
FIRE ₂₅	5	7.46 \pm 3.7	33.5 \pm 6.89	0.8 \pm 0.1	44.7 \pm 9.3	Organic
	30	8.64 \pm 1.3	9.33 \pm 4.30	0.4 \pm 0.1	18.5 \pm 1.8	Mineral
	50	2.57 \pm 0.4	2.38 \pm 0.78	0.2 \pm 0.1	12.5 \pm 2.0	Mineral
FIRE ₄₆	5	5.27 \pm 1.3	42.3 \pm 3.42	1.4 \pm 0.0	30.4 \pm 1.2	Organic
	30	7.17 \pm 1.9	3.97 \pm 0.86	0.3 \pm 0.0	14.8 \pm 0.9	Mineral
	50	4.42 \pm 0.8	4.44 \pm 0.95	0.4 \pm 0.1	12.9 \pm 1.5	Mineral
FIRE ₁₀₀	5	7.09 \pm 1.4	38.9 \pm 2.43	1.1 \pm 0.2	48.4 \pm 4.1	Organic
	30	6.40 \pm 1.0	8.74 \pm 3.02	0.6 \pm 0.2	13.7 \pm 1.6	Mineral
	50	1.28 \pm 0.2	3.79 \pm 0.87	0.3 \pm 0.1	15.1 \pm 2.7	Mineral

where a is the intercept of the model and parameters from b to g are regression coefficients for the factors. The following fixed factors were used in the models: AD (m) is the depth of the active layer, $Biomass$ (kg m⁻²) is the above ground biomass consisting of tree and ground vegetation (shrubs, grasses, mosses and lichens), T (°C) is soil temperature at the time of sampling in each soil depth, $AD \times T$ is the cross effect of active layer depth and soil temperature and CN (%) is the soil C/N ratio. Sampling line was set as a random factor RDM . The factors and models are further

described in Table 2. Time since fire was not included in the model as it correlates with AD and $Biomass$ and its effects are apparent already through these factors.

The best model for each soil depth was selected for sensitivity analyses of the variables. The sensitivity analyses were performed to determine how much the change of factors in the best fitting model would affect the predicted SOM_{non} . This was conducted using mean values for each factor and changing one factor at a time by $\pm 10\%$, while others remained unchanged. With every change, value for SOM_{non} was calculated

Table 2 Linear mixed effects models fitted against the insoluble SOM fraction (SOM_{non}) and experimental factors separately for each sampling depth (5, 30, 50 cm)

Model ID	Mixed effect model	Depth (cm)
Model 1 (initial model)	$SOM_{non} = a + bAD + cBiomass + dT + e(T \times AD) + fCN + gRDM$	
Model 2	$SOM_{non} = a + bAD + cBiomass + dT + fCN + gRDM$	5
Model 3	$SOM_{non} = a + bAD + cBiomass + fCN + gRDM$	5
Model 4	$SOM_{non} = a + bAD + cBiomass + gRDM$	5
Model 2b	$SOM_{non} = a + bAD + cBiomass + dT + fCN + gRDM$	30
Model 3b	$SOM_{non} = a + bAD + cBiomass + fCN + gRDM$	30
Model 4b	$SOM_{non} = a + cBiomass + fCN + gRDM$	30
Model 2c	$SOM_{non} = a + bAD + dT + e(T \times AD) + fCN + gRDM$	50
Model 3c	$SOM_{non} = a + bAD + dTf + CN + gRDM$	50
Model 4c	$SOM_{non} = a + bAD + fCN + gRDM$	50
Model 5c	$SOM_{non} = a + fCN + gRDM$	50

The best-fitted model for each depth is highlighted with bold lettering. Fixed factors are: AD = depth of the active layer, $Biomass$ = total biomass of trees and ground vegetation, T is soil temperature, $AD \times T$ is the cross effect of active layer depth and soil temperature and CN is soil carbon to nitrogen ratio. RDM is the random factor (sampling line)

using Eq. 2 and compared to a value where none of the factors were changed.

In addition, we compared different fractions as explaining factors for Microbial C (Eq. 3). Due to the fractions being dependent on each other, the model included only one fraction at a time and sampling line as a random factor. The best describing factor was chosen based on model *P* value and pseudo R^2 .

$$MC = a + b\text{Fraction} + cRDM \quad (3)$$

where *a* is the intercept, *b* and *c* are regression coefficients, *MC* is the microbial carbon (mg g^{-1}), *Fraction* is the size (%) of the fraction (water-, ethanol-, acid-soluble, or insoluble) and *RDM* is the random factor (sampling line).

Results

Soil organic matter fractions

Generally, the soil layers differed significantly in terms of the proportions of insoluble and soluble SOM fractions (Fig. 1). At 5 cm depth, the soluble fractions together formed on average 65% of SOM and the insoluble fraction around 35%, whereas at the 30 cm and 50 cm depths the insoluble fraction constituted to 75–84% of SOM and soluble fractions together only 16–25%.

The fire had effects on the size of the water- and ethanol-soluble fractions in the surface soil (5 cm) (Table S3, Online resource). These fractions were smallest mainly shortly after the fire and increased along with the forest succession. At 5 cm depth (Fig. 1a), areas FIRE₂₅ and FIRE₄₆ had a larger proportion of water-soluble SOM than area FIRE₃ ($P < 0.05$), but there was no difference between area FIRE₁₀₀ and area FIRE₃. In addition, the FIRE₄₆ area had higher water-soluble fraction than the FIRE₁₀₀ area ($P < 0.05$). Both younger fire areas, FIRE₃ and FIRE₂₅, had a smaller ethanol-soluble fraction than the older fire areas ($P < 0.001$). Further, the acid-soluble fraction and the insoluble fraction were not significantly different between fire ages, though there was a tendency towards an increase in the more recently burnt areas. *P*-values for comparison between fire ages at all depths are given in more detail in tables S3–S5 (Online Resource).

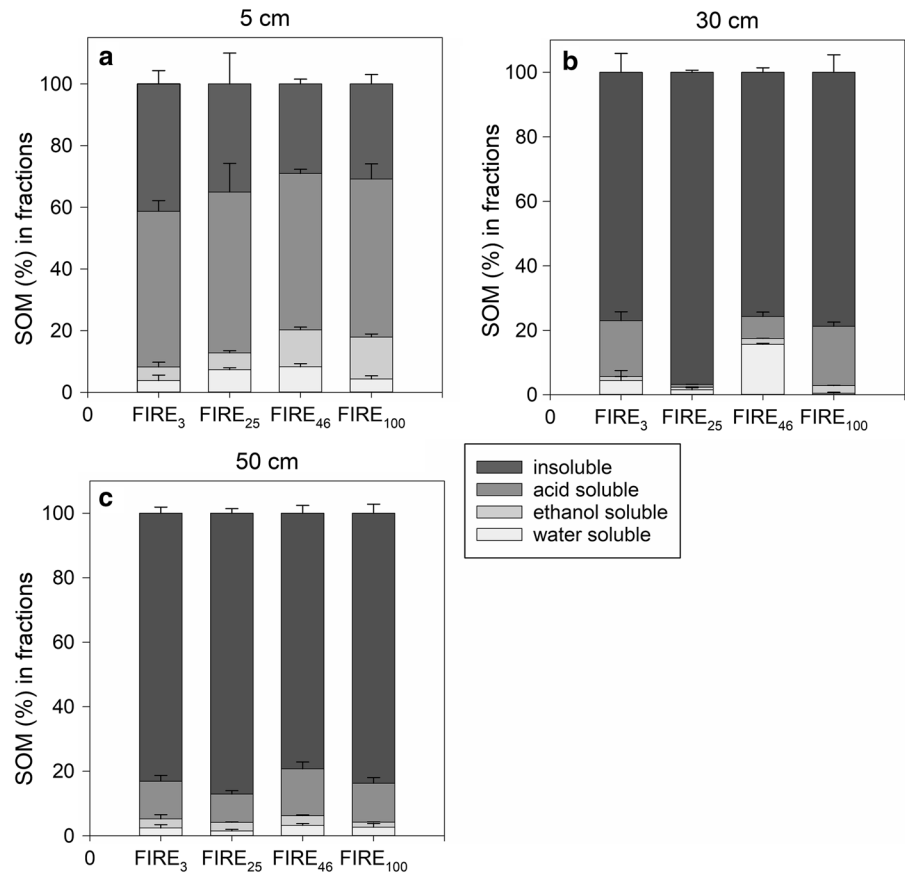
Deeper in the soil, the changes in SOM fractions along with the fire chronosequence were not as consistent as at the surface. In the 30 cm layer (Fig. 1b), area FIRE₄₆ had a larger water-soluble fraction than all the other areas ($P < 0.001$) (Table S4, Online resource). In addition, area FIRE₁₀₀ had a larger ethanol-soluble fraction than FIRE₂₅ and FIRE₃ ($P < 0.05$). The 30 cm layer also deviated from the two other layers in the size of the acid-soluble and insoluble fractions (Fig. 1b). Both FIRE₁₀₀ and FIRE₃ areas had greater acid-soluble fraction than areas FIRE₂₅ and FIRE₄₆ ($P < 0.001$), with FIRE₄₆ having a larger acid-soluble fraction than FIRE₂₅ ($P < 0.05$). Area FIRE₂₅ had a larger insoluble fraction than any of the other areas ($P < 0.001$). In the 50 cm layer (Fig. 1c), the only differences were in the ethanol-soluble fraction where area FIRE₄₆ showed larger values than FIRE₁₀₀ and FIRE₃ ($P < 0.001$), while in FIRE₂₅ the ethanol-soluble fraction was larger than in area FIRE₁₀₀ ($P < 0.001$) (Table S5, Online resource). The approximated amounts of each fraction from total soil C values (kg m^{-2}) varied with soil depth e.g. water-soluble fraction at the 5 cm depth was 0.13–0.43 kg m^{-2} and at 50 cm depth 0.02–0.15 kg m^{-2} (Table S6, Online Resource). For areas FIRE₄₆ and FIRE₁₀₀ this depth represents permafrost SOM.

$\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ in bulk soil

In general, fire seems to have had an age- but no depth-wise effect on the soil ^{15}N isotopic values. The age-related trend in $\delta^{15}\text{N}$ -values (Fig. 2a) appeared at 5 cm with the FIRE₃ area being more enriched compared to area FIRE₁₀₀ ($P < 0.05$). The pattern was similar, though only marginally so ($P = 0.063$), at 30 cm depth (Fig. 2b). At 50 cm depth there were no significant differences (Fig. 2c). Moreover, we found no significant differences when we compared the ^{15}N δ -values between depths for each fire age.

The $\delta^{13}\text{C}$ values showed that the 5 cm depth was less depleted in $\delta^{13}\text{C}$ ($P < 0.05$) in the more recently burnt areas (FIRE₃ and FIRE₂₅) than in the older areas (FIRE₁₀₀ and FIRE₄₆). Yet, there was no difference between areas FIRE₂₅ and FIRE₃. At 30 cm depth (Fig. 3b) area FIRE₄₆ was more enriched in ^{13}C than all the other areas ($P < 0.05$). At 50 cm depth (Fig. 3c), we found no statistical differences. Contrastingly to $\delta^{15}\text{N}$, the $\delta^{13}\text{C}$ -values (Fig. 3a) were

Fig. 1 The mean chemical fractions of SOM from the 5, 30 and 50 sampling depths, presented as a percentage of total SOM. The error bars show standard errors of mean



significantly more depleted in the 5 cm ($P < 0.001$) layer than in the 30 and 50 cm layers. Also, in general, we found no significant differences in total soil C and N concentrations or C/N ratio between fire ages ($P > 0.05$).

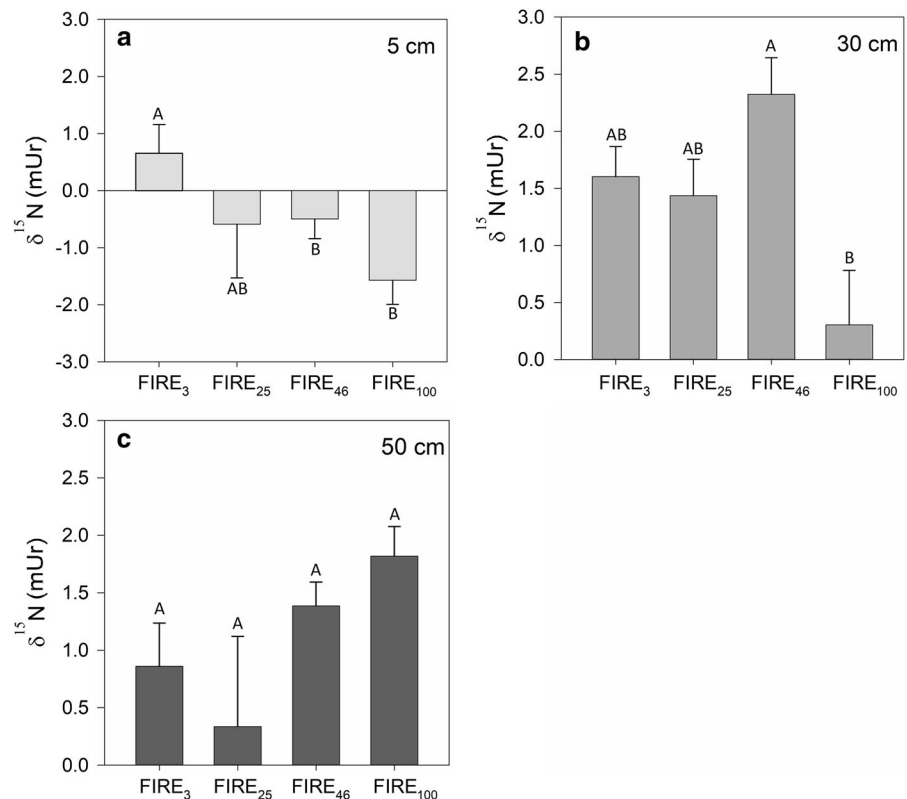
Linear mixed effects model analyses

Based on the linear mixed effect model analyses (Table 2), the size of the insoluble SOM fraction at 5 cm depth was best explained by the active layer depth and biomass (Model 4), which together explained 22% of the variation in the size of the fractions (Table S7, Online resource). At 30 cm depth the best explaining factors were biomass and C/N ratio (Model 4b), explaining 85% of the variation (Table S8, Online resource). At 50 cm depth, the best model (Model 5c), including only C/N, could explain 10% of the insoluble SOM fraction size (Table S9, Online resource).

Microbial C was best predicted by the size of the insoluble fraction at the 5 and 30 cm depths (Table S10, Online resource). At 5 cm depth the insoluble fraction explained 27% of the microbial C ($P = 0.046$), while at 30 cm depth ($P < 0.001$) it explained 97%. The size of the insoluble fraction and microbial biomass seem to have a negative correlation (Table S10, Online Resource). Still, at 50 cm depth none of the models were significant.

Sensitivity analyses of the best model (Model 4) at 5 cm soil depth revealed that changing either of the variables (*AD* or *Biomass*) results in a change of 1.5–2.0% in the size of the insoluble SOM fraction (Fig. S2, Online resource). The model was most sensitive to changes in the biomass. At 30 cm depth, where best model was 4b, the resulting change in insoluble SOM fraction size would be 0.1–0.6% (Fig. S2, Online resource) with the model again being most sensitive to changes in the biomass. Finally, at 50 cm soil depth (Model 5c) a change in the C/N ratio

Fig. 2 The natural $\delta^{15}\text{N}$ -values of bulk soil samples at 5, 30 and 50 cm depths from each fire area. The letters denote significant differences between age classes



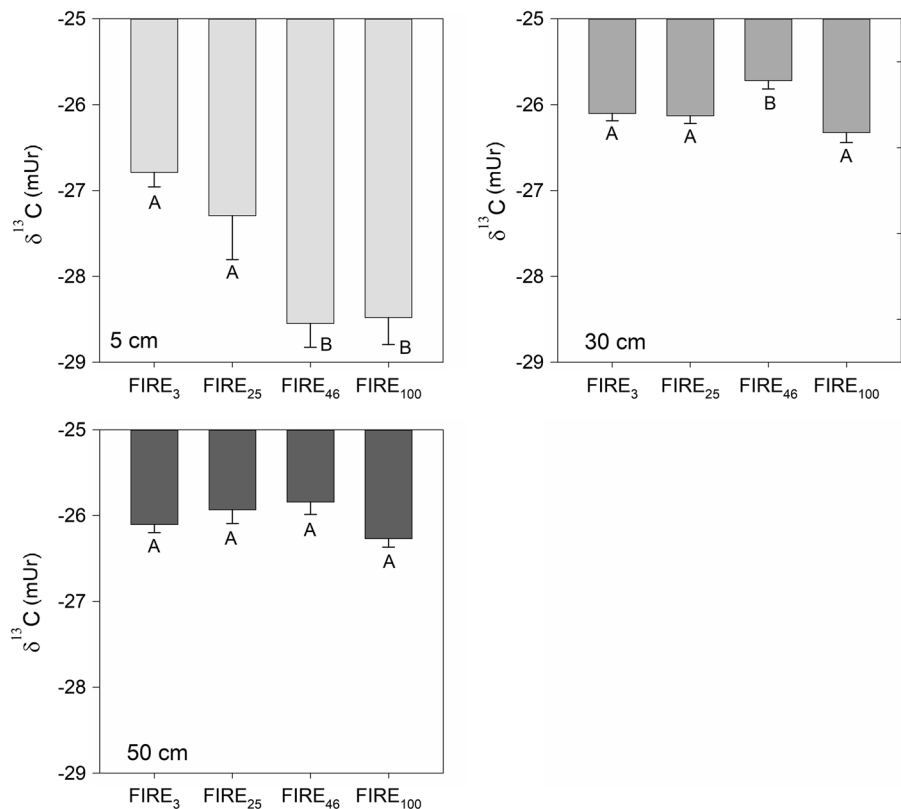
leads to a 1.2% change in the insoluble SOM fraction (Fig. S2, Online resource).

Discussion

The increased frequency of forest fires decreases both the ability of forests to store C and the degree of permafrost recovery (Hoy et al. 2016). Therefore, effects of fire on SOM quality, microbial activity and permafrost thaw could have a significant effect on greenhouse gas emissions during fire intervals. Our results indicated that the proportional sizes of the most soluble fractions (water and ethanol) in the surface soil (5 cm depth) were in general smaller after fire and increased with time. However, changes were not consistent in the two deepest layers. It may be that the effects of fire do not reach to depths of 30 and 50 cm, although fire has clearly changed the active layer depth (Köster et al. 2017), exposing previously frozen soil at 50 cm to thaw. Also study by Shibata et al. (2003) noted that fire did not affect the water-extractable C in the mineral soil.

The different SOM fractions can be thought to represent different SOM pools with differing turnover times. These SOM pools are decomposing with different rates in the soil and can be roughly divided from fast to slow in following order: sugars, starches, simple proteins, crude proteins, hemicellulose, cellulose, fats, waxes, resins and lignins (Osman 2013). Also decomposition models, such as Yasso and CENTURY, often divide SOM into pools, based on their turnover times. Yasso divides decomposing material to extractives, celluloses, lignin-like materials, humus 1 and humus 2 (Liski et al. 2005). In these pools the water-and ethanol extractable SOM could be thought to belong to the extractives pool with a decomposition rate of 0.48 year^{-1} (rates for conifer litter). The decomposition rate for cellulose is 0.30 year^{-1} and lignin-like compounds 0.22 year^{-1} , while humus 1 pool has a decomposition rate of 0.012 year^{-1} and humus 2 pool 0.0012 year^{-1} (Liski et al. 2005). In CENTURY the pools are divided to active pool (turnover time of months to few years), slow pool (turnover time of 20–50 years) and stable pool (turnover time of 400–2000 years)

Fig. 3 The natural $\delta^{13}\text{C}$ -values of bulk soil samples at 5, 30 and 50 cm depths from each fire area. The letters denote significant differences between age classes



(Century manual). For example, Bot and Benites (2005) have allocated different SOM groups to three pools: sugars, as easily decomposing, could be placed in the active cycling pool, while fats, waxes and cellulose represent the slow pool. Lignins, as resistant compounds, form the stable SOM pool (Bot and Benites 2005; Kutsch et al. 2009). Though these pools may not be straightforwardly comparable to the pools used in CENTURY, it can be deduced that there is a vast difference in the decomposition rates of these different SOM pools. Further, the degradability of a compound is affected by its molecular weight and structure: material with low molecular weight (e.g. sugars, aminoacids, phenols) is easier to decompose than material with high molecular weight (e.g. cellulose, lignin, cutin) (Cotrufo et al. 2013). SOM that is most readily available is characterised by its solubility to water (Marschner and Kalbitz 2003).

We observed no significant changes with time after fire in the insoluble fraction. In case of fire chronosequences, in an early succession forest (recently burnt) there may be a higher proportion of recalcitrant (charred) SOM (González-Pérez et al. 2004; Wardle

et al. 2008) as the organic layer may have been at least partially burnt. In late succession stage forests, on the other hand, incoming plant material, such as evergreen needles and woody material, is generally less easily decomposable (Brassard and Chen 2006). In our previous study, conducted in the same areas as this study, we observed that the organic layer depth continued to increase several decades after fire (Köster et al. 2017). Also the vegetation cover, both biomass and species composition, was affected by time since fire (Table S2, Online Resource). The youngest fire area had visible ashy, bare patches of ground, no living trees, less mosses and ground vegetation mostly consisting of *Equisetum sylvaticum* L. The older areas FIRE₄₆ and FIRE₁₀₀ (Table S2, Online Resource) had more lichen, mosses (*Sphagnum* sp., *Pleurozium* sp), shrubs (*Ledum groenlandicum* Oeder, *Vaccinium vitis-idaea* L., *V. Uliginosum* and some *Rubus chamaemorus* L.) and mature trees (Köster et al. 2017). The area FIRE₂₅ was similar to these, but with some remaining unvegetated patches and *Cladonia* and *Cladina* lichens. These changes in vegetation cover affect the quality of incoming organic matter to

soil, thus affecting also the different proportions of SOM fractions. This is because different species have differing decomposition rates. Such shrubs as *Vaccinium vitis-idaea* and *Rubus chamaemorus* L. decompose relatively slowly (Nilsson and Wardle 2005), while *Equisetum* sp species might decompose faster (Marsh et al. 2000). Further, though mosses decompose slowly, they have been observed to advance decomposition of other species by keeping up moisture levels in the litter layer (Wardle et al. 2003). Thus, in both recently burnt and late succession stage forests, there are possibly more sources for recalcitrant SOM than in mid-succession forests, perhaps explaining why we observed no significant changes with time in the insoluble fraction. We detected a trend (at 5 cm depth) of increasing proportions of recalcitrant SOM in younger, more recently burnt areas, indicating a strong effect of fire on SOM quality. Thus, charred SOM could have a noticeable role at some point in time. However, in an early succession forest, the input of resistant material may also be counteracted by the easily decomposable plant material deriving from the herbaceous plant species in the early succession stage (Chen and Shrestha 2012) or perhaps from the decomposing root material.

In the labile fractions (water and ethanol) the most obvious changes were observed at 5 cm soil depth. FIRE₃ and FIRE₁₀₀ did not show differences in the size of the water-soluble fraction, but the middle-aged fire areas had higher fractions of water-soluble SOM. The similarity between FIRE₃ and FIRE₁₀₀ is probably related to the previously mentioned succession dynamics. Alternatively, there could be greater leaching, or loss as run-off, of water-soluble SOM in these areas compared to FIRE₂₅ and FIRE₄₆. In area FIRE₃ there was probably less evapotranspiration than in older fire areas due to smaller transpiring leaf biomass. This could mean that a proportionally larger amount of water is moving down in the soil profile compared to the older forests, where vertical movement is also limited by the higher permafrost surface. Further, some pyrogenic compounds are actually water-soluble (Norwood et al. 2013) and might show in the water-soluble fraction in addition to sugars. However, this trend was different in the ethanol soluble fraction. Both FIRE₃ and FIRE₂₅ had smaller ethanol-soluble fractions than FIRE₄₆ and FIRE₁₀₀, meaning that waxes and fats form a greater proportion of SOM during the later succession. This is probably also

related to changes in vegetation during the succession. In the later stage of forest succession, there is an increased biomass of coniferous trees and shrubs, with large amount of waxes in their cuticula, (Dickinson 1974; Brassard and Chen 2006), thus producing litter which increases the amount of fat and waxes in the SOM.

The fraction of insoluble SOM was higher in the 30 and 50 cm soil layers, which could be expected as deeper soils typically contains less readily decomposable SOM (Trumbore 2014). In addition, studies on black carbon have shown it to be easily translocated vertically in the soil and therefore it may have accumulated into subsoils (Dai et al. 2005; Rumpel et al. 2009). On the other hand, the ratios of SOM fractions are not only dependent on the formation of resistant compounds but are greatly affected by decomposers. Fungal species have an important role in degrading lignin (Waldrop et al. 2010). It has been found that if the fungal community becomes wiped out or reduced during the fire, the degradation of lignin will decrease (Waldrop et al. 2010), meaning that it could reside in the soil for a longer period. This could be apparent in the size of the insoluble SOM fraction for some time after the fire as Klason lignin found here is included in the recalcitrant fraction. For example, it takes several decades for the fungal biomass to recover after fire (Köster et al. 2016; Zhou et al. 2018). Yet we found that the length of time after fire had only a tendential, but insignificant, effect on the amount of insoluble and acid-soluble SOM, and can thus only speculate here about the role of fungi.

Our results show that the effects of fire on SOM are most prominent at the soil surface. The SOM fractions and isotopes showed very small, or no, differences between fire ages at 50 cm depth. This was further supported by the mixed effects model, where none of the predicting factors seemed to be significant and the only variable surviving to the best model was C/N ratio. Contrastingly, at both 5 and 30 cm depths the best model included biomass (trees, shrubs, grasses, mosses and lichens), and both models were most sensitive to changes in biomass. As the biomass in the model included above ground (tree- and ground vegetation), it could be that changes in them reached the 5 and 30 cm depths but not 50 cm. The effect of biomass on the size of the insoluble SOM fraction might occur through at least two opposing processes: loss of biomass during the forest fire and, on the other

hand, an increase of biomass and changes in litter quality with succession (Brassard and Chen 2006). At 5 cm, the effect of active layer depth on the insoluble SOM fraction was close to that of the biomass (Fig. S1, Online resource), which could be related to possible temperature changes caused by retreating/returning permafrost and the consequent effects on vegetation. At 30 cm depth the effect of the C/N ratio was also close to that of biomass, possibly linking these two together as C/N ratio is also an indicator of incoming litter quality (Johnson and Wedin 1997). Both biomass and C/N ratio correlated negatively with the size of the insoluble SOM fraction. Still, the lack of explaining factors at 50 cm is perhaps connected to the proximity of permafrost or high soil water content.

The mixed effects model analysis further showed that microbial biomass is, amongst the SOM fractions, best predicted by the size of the insoluble fraction (at 5 and 30 cm depth). Moreover, microbial biomass and insoluble fraction size appear to correlate negatively, so that when the size of the insoluble fraction is high, the microbial biomass is low. This indicates that SOM quality is more important than SOM quantity in northern boreal ecosystem, as microbes usually prefer labile SOM fractions as a C source (Rinkes et al. 2011). Microbial biomass at 50 cm depth, on the other hand, is probably limited by factor other than the availability of labile SOM, such as soil temperature, the presence of permafrost and anoxic conditions due to a high ground water table on top of the permafrost (Hobbie et al. 2000b).

We discovered an age-related trend for $\delta^{15}\text{N}$, with the most recently burnt area having a more enriched surface soil than area FIRE₁₀₀. During a fire, the ^{15}N depleted litter layer may burn almost completely, rendering the burnt area with material enriched with ^{15}N (Högberg 1997). For example, leaf litter of conifer trees seems to commonly be ^{15}N depleted (Hobbie et al. 2000a; Sah and Ilvesniemi 2007). Usually plants that do not fix N_2 have higher $\delta^{15}\text{N}$ values than plants fixing N_2 (He et al. 2009) and differences in leaf $\delta^{15}\text{N}$ values are caused by discrimination against heavier isotopes in N uptake and transfer from mycorrhiza to plant (Hobbie and Hobbie 2006, 2008). The discrimination is also affected by the type of mycorrhiza (Michelsen et al. 1998), which might become apparent after such disturbances, as fire, where microbial species compositions are possibly altered. A change in the species composition was discovered also in this

same fire chronosequence by Zhou et al. (2018). Previous studies have also noted that burning of the depleted litter is not necessarily the most dominating reason for enrichment with heavier isotopes. Instead, the enrichment may be attributed to volatilisation processes, which discriminate against heavier isotopes (Cook 2001; Boeckx et al. 2005) and root activity (Pumpanen et al. 2017). Forest fires generally also increase N leaching (Lamontagne et al. 2000), especially the leaching of ^{15}N depleted NO_3^- (Pardo et al. 2002), which may have contributed to ^{15}N enrichment in the FIRE₃ area. On the other hand, water-soluble pyrogenic compounds might also be enriched with ^{15}N and translocating down the soil profile. The soil ^{15}N enrichment tends to increase with soil depth (Högberg 1997), as a result of microbial degradation of SOM (ammonification, nitrification and denitrification processes), where enriched material is accumulating in the residues (Makarov 2009; Girona-García et al. 2018). Fire has also been shown to decrease the differences in ^{15}N enrichment along the soil profile, by causing enrichment in the upper 20 cm (Boeckx et al. 2005); a trend that was also somewhat apparent in our results. ^{15}N was enriched throughout the soil profile in area FIRE₃, while older fire areas were slightly depleted at the surface and enriched in the two deepest layers. However, as these differences were so small that they were not statistically significant, it is not possible to make conclusions based on them.

A similar trend as for ^{15}N was also found for ^{13}C abundance, with the two most recently burnt fire areas having less depleted values in the surface soil (5 cm) than in areas FIRE₁₀₀ and FIRE₄₆. Here we also noticed a trend where the ^{13}C enrichment increased with depth, which has been detected quite commonly across ecosystems (Brüggemann et al. 2011). Typically, higher $\delta^{13}\text{C}$ values indicate the presence of ^{13}C enriched compounds, such as polysaccharides and amino acids, whereas ^{13}C depletion could be caused by the existence of depleted materials, such as lignin and lipids (Rumpel and Kögel-Knabner 2011). The leaf isotopic values of C3 plants generally average around -27 mUr (Brodribb and Hill 1998; Cernusak et al. 2009; Orchard et al. 2010), whereas lipids and lignin tend to be -2 mUr to -3 mUr more depleted compared to average leaf isotope value (Ehleringer et al. 2000). Cellulose tends to have similar, or slightly higher (Loader et al. 2003), values as lignin (Hobbie and Werner 2004). Some studies have found that

during fire, cellulose is lost more than lignin and lipids, leading to lower $\delta^{13}\text{C}$ values in the heat modified OM (Alexis et al. 2010). On the other hand, post-fire litter of standing vegetation has been found to have higher $\delta^{13}\text{C}$ values than pre-fire litter, with post-fire litter forming much of the pyrogenic material (Alexis et al. 2010). Both of these factors may have contributed to the changes observed in the $\delta^{13}\text{C}$ values in this study. For example, in the top 5 cm, older fire areas had larger ethanol-soluble fraction (lipids etc.) and were more depleted. These findings seem to support each other and indicate that as succession proceeds after the fire, the size of the relatively labile SOM fraction increases.

In depth-wise comparison of the bulk soil the surface (5 cm) was more depleted of ^{13}C than the deeper layers (30 and 50 cm), which seems to be the norm with ^{13}C in forest soils (Balesdent et al. 1993; Krull et al. 2002; Boström et al. 2007). The slight enrichment along the depth profile is also supported by fractionation results, where deeper layers had greater recalcitrant SOM fractions. The ^{13}C enrichment with depth in forest soils has been connected to four different hypotheses: the Suess effect, microbial fractionation, microbial preference and soil mixing (Ehleringer et al. 2000). The Suess effect, depletion of atmospheric CO_2 of ^{13}C caused by the burning of depleted fossil fuels (Balesdent et al. 1990; Trolier et al. 1996), could lead to older/deeper soil layers having less depleted ^{13}C values corresponding to atmospheric $^{13}\text{CO}_2$ values of that time (Ehleringer et al. 2000). On the other hand, microbial fractionation prefers lighter isotopes, leading to enrichment in the residual SOM (Natlhoffer and Fry 1988; Ehleringer et al. 2000), while microbial preference is related to which kind of substrates the microbes favor (Ehleringer et al. 2000). Finally, C mixing suggests that during a loss of soil C there is mixing between fresh and old SOM, therefore causing changes in ^{13}C values. In addition to these, the SOM ^{13}C values are much dependent on the origins of the material, such as type of litter (Benner et al. 1987). Regardless of the reason, in our results, the differences in ^{13}C values between the surface and deeper layers were not very large, which could be at least partly explained by possible enrichment at the surface after the fire. At the same time, in the two oldest fire areas the deepest (50 cm) samples were below the permafrost, seemingly

indicating that permafrost SOM in this area is relatively recalcitrant.

Our results indicated that fires do decrease the quality of SOM in the surface layer, but that the effect of fire on soil at the permafrost surface is more indirect and happens through changes in soil temperature and deepening of the active layer. In the oldest fire areas, FIRE₄₆ and FIRE₁₀₀, the 50 cm (also 30 cm for FIRE₁₀₀) sampling depth was below the permafrost surface, showing that the permafrost soils in these areas were mostly very recalcitrant SOM and generally did not differ from the non-permafrost samples of the younger fire areas. While other studies have found somewhat differing results for the quality of permafrost SOM (Dutta et al. 2006; Waldrop et al. 2010; Moni et al. 2015), direct comparisons are difficult to make since there are differences between study sites, sampling depths and methods. As we did not sample the deep permafrost soils, some caution also needs to be taken while interpreting the results. However, our results indicated that, while in our study areas the permafrost may thaw, most of the permafrost SOM will possibly reside in the soil for a long period, due to its recalcitrant nature. Moreover, based on the sensitivity analyses, the permafrost/permafrost-affected soil does not seem to be very sensitive to changes, as the size of the recalcitrant fraction did not change significantly even if the factors affecting decomposition were changed in the sensitivity analysis of our linear mixed effect models. Despite this, it is possible that some of this recalcitrant SOM could decompose, as the input of fresh, easily decomposable C caused by fire could have a priming effect (Fontaine et al. 2007). Recalcitrant SOM may also be more temperature sensitive (Davidson and Janssens 2006; Karhu et al. 2010), unless protected by physical protection (Gillabel et al. 2010), meaning that the possible warming of soil, if the insulating organic layer is burnt and permafrost starts to thaw, may cause C emissions. Indeed, deeper active layer was found at the more recently burnt areas, which hints in that direction. Still, the formation of recalcitrant SOM by fire may act as a counter-affecting process. Possible C losses from these upland mineral forest soils are then largely dependent on whether in the warming climate the permafrost, lost after the fire, will continue to return as succession proceeds, and whether some following process will be able to degrade the potentially very resistant permafrost SOM.

Based on our results the effect of fires on SOM quality seems to be mostly on the surface soil. Initially after fire, the surface soil SOM quality is reduced and, as succession proceeds, the ratios of the SOM fractions revert towards pre-fire status. The SOM in permafrost soils in these forest areas seem to be mostly recalcitrant. These indicate that even if permafrost thaws, the scale and rate of C emissions from the upland mineral permafrost soils could be limited. However, further research is needed for quantifying the amount of SOM in different fractions and assessing the SOM quality of deep permafrost soils.

We conclude that a consequence of changing climate is that fire intervals become shorter and the forest rotation time is decreased, leading to less C being stored in the soil (Kaipainen et al. 2004; Hoy et al. 2016). While labile C is lost in fires, pyrogenic C is accumulated to the soil. At the same time, the pioneer species in vegetation might prosper relatively longer times compared to late succession species, which affects the quality of SOM via incoming litter. These factors may be seen in a proportional decrease in the size of the labile SOM fraction and increase in the size of recalcitrant fraction: while labile C is lost in fire, pyrogenic C is stored. This might lead to decrease in CO₂ emissions to the atmosphere from post-fire soil decomposition. At the same time, the permafrost has less time to recover and reach original depths (Hoy et al. 2016), meaning that the permafrost SOM, though possibly recalcitrant, will be vulnerable to decomposition for longer periods of time. The effect of these counteracting processes on permafrost C pools require further studies.

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