

<https://helda.helsinki.fi>

Wooden Breast Myodegeneration of Pectoralis Major Muscle Over the Growth Period in Broilers

Sihvo, H. -K.

2017-01

Sihvo , H -K , Linden , J , Airas , N , Immonen , K , Valaja , J & Puolanne , E 2017 , '
Wooden Breast Myodegeneration of Pectoralis Major Muscle Over the Growth Period in
Broilers ' , Veterinary Pathology , vol. 54 , no. 1 , pp. 119-128 . <https://doi.org/10.1177/0300985816658099>

<http://hdl.handle.net/10138/310093>

<https://doi.org/10.1177/0300985816658099>

unspecified

acceptedVersion

Downloaded from Helda, University of Helsinki institutional repository.

This is an electronic reprint of the original article.

This reprint may differ from the original in pagination and typographic detail.

Please cite the original version.

Wooden breast myodegeneration of pectoralis major muscle over the growth period
in broilers

H.-K. Sihvo*, J. Lindén, N. Airas, K. Immonen, J. Valaja, E. Puolanne

Department of Food and Environmental Sciences, P.O. Box 66, FI-00014 University
of Helsinki, Finland (HKS, KI, EP)

Department of Veterinary Biosciences, P.O. Box 66, FI-00014 University of Helsinki,
Finland (HKS, JL, NA)

Department of Agricultural Sciences, P.O. Box 28, FI-00014 University of Helsinki,
Finland (JV)

* Corresponding author: Sihvo H.-K.

Address: Department of Food and Environmental Sciences, P.O. Box 66, FI-00014
University of Helsinki, Finland

Email: hanna-kaisa.sihvo@helsinki.fi

Tel: +358405193741 Fax: +358294158475

Abstract

Wooden breast (WB) myopathy of broiler chickens is a myodegenerative disease of an unknown etiology and is macroscopically characterized by a hardened consistency of the pectoralis major muscle. Our aim was to describe the development and morphology of WB over the growth period in broilers. Additionally, the effect of restricted dietary selenium on the occurrence of WB was examined by allocating the birds in two dietary groups: restricted and conventional level of selenium. The experiment included 240 male broilers that were euthanized at ages of 10, 18, 24, 35, 38 or 42 days and evaluated for WB; abnormal hardness of the pectoralis major muscle rendered the WB diagnosis. The severity and the distribution of the lesion and presence of white striping were recorded. The first WB cases were seen at 18 days; 13/47 birds (28%) were affected and the majority exhibited a mild focal lesion. In the following age groups the WB prevalence varied between 48% and 73% and the lesion was usually diffuse and markedly firm. White striping often coexisted with WB. Histological evaluation performed on 111 cases revealed a significant association of myodegeneration and lymphocytic vasculitis with WB. The vasculitis and perivascular cell infiltration were restricted to the veins. Restricted dietary selenium did not affect the occurrence of WB ($P = 0.44$). Our results indicate that WB starts focally and spreads towards a diffuse and more severe lesion.

Keywords

poultry, chickens, skeletal muscle, broiler, wooden breast myodegeneration, pectoralis major, vasculitis, white striping

Wooden breast (WB) myodegeneration is an emerging disease in broiler chickens, macroscopically recognized as hardness of the pectoralis major muscle, often accompanied with pale color and white striping.²⁷ In slaughter-age broilers, i.e. approximately 35 days and older, the main histological lesions of the breast muscle consist of chronic myodegeneration with regeneration and interstitial edema, accumulation of loose connective tissue, or fibrosis.²⁷ Replacement of the severely degenerated muscle fibers with connective tissue may partly account for the muscle hardness, but the hardened consistency seems to also occur in cases without excessive fibrosis.²⁷ Clinical signs have not been reported with WB and the lesion is usually detected post mortem. Although affected birds can exhibit diffuse and severe chronic myodegeneration of the pectoralis major muscle, it remains controversial whether the condition is painful.²⁷

The cause and pathomechanism of WB are currently unknown. There are several causes of myodegeneration in animals, such as deficiency of selenium or vitamin E, an excess of their antagonists, ionophore toxicity, exertional myopathy, hypoxia, and muscular dystrophies.^{14, 27, 29} However, those are not associated with the substantial hardness of the muscle tissue, which is the key feature and the current basis for the diagnosis of WB. In addition, many of the previously described causes also affect other organs, whereas WB seems to be restricted to the skeletal muscle.^{14, 29}

Selenium deficiency as a cause of oxidative damage leading to degenerative myopathy has been described in many animal species including broiler chickens.^{7, 12, 14} Grain is the usual content in the diet of broilers; the variation of the natural selenium content of the soil, as well as the use of fertilizers devoid of supplemented selenium are potential causes for insufficient selenium content of broiler feed.

Recently, organic selenium has been shown to have a better bioavailability and antioxidative capacity than the inorganic selenium that traditionally has been used as a feed additive.^{5, 9, 23}

In addition to WB, there are other myodegenerative conditions of unknown etiology with a recent emergence in the broilers. White striping is a condition affecting the breast and thigh muscles of broilers and it occurs alone or concurrently with WB.^{17, 27} It is macroscopically characterized by pale striations up to a few millimeters wide parallel to the muscle fibers. Histologically it shares many features with WB, such as degeneration, regeneration, interstitial inflammatory infiltrates and fibrosis.^{17, 25, 27} A dorsal myodegeneration, very similar to WB both macroscopically and histologically, has been reported in the anterior latissimus dorsi (ALD) muscles of broilers in Brazil.³¹ Both the ALD myopathy and white striping have been associated with the heaviest birds or fillets of the flock or increased growth rate.^{3, 15, 16, 31}

The objective of this study was to characterize the time of onset of WB, its progression over the growth period, and the associated morphological features. Additionally, we tested the effect of reduced selenium content of the feed on the occurrence of WB.

Materials and Methods

Two hundred and forty one-day-old male Ross 508 broiler chicks were divided into 24 pens, ten birds per each, in one room of an experimental facility. They received no medical treatments or vaccinations; their feed, however, included a narasin-based coccidiostat at the level of 0.07% (Monteban, Elanco, IN, USA). The maintenance

and clinical observation were on daily basis. Due to heating lamps, the lighting was continuous for the first 7 days, from which onwards a daily 6-hour period of darkness was applied. The bedding consisted of wood chip litter on rubber mats on a concrete floor. Other husbandry conditions were adjusted according to the recommendations for conventional breeding.¹ Experimental procedures were not performed during life, and the birds were euthanized at the ages of 10, 18, 24, 35, 38 or 42 days (d) by mechanical cervical dislocation, followed by phlebotomy of the jugular veins. The selection of birds for euthanasia was as follows: two birds per pen were randomly selected for the first and second age group (10 and 18 d), one random bird per pen for the third age group (24 d), and then all remaining birds from 10 pens at 35 d, 10 pens at 38 d and the last 4 pens at 42 d (Table 1). This arrangement aimed at mimicking the density of birds per square meter near to that used in the conventional breeding. However, birds that died between the set age points were excluded from the study, which affects the number of birds per age group. For each age group, a balanced number of birds were chosen from both dietary treatments, which are described in detail below. Before euthanasia, each bird was weighed individually. Ethical permission for the experiment was issued by the Research Animal Resources Committee of the University of Helsinki (Decision number KEK12-045).

Dietary treatment

The birds were assigned for two ad libitum dietary treatments, SeLow and SeNorm, 12 pens each by random allocation. SeLow feed contained inorganic selenium (sodium selenite, Na_2SeO_3 , Anima Ltd., Krakow, Poland) 0.11 mg/kg for the first 7 d and then 0.13 mg/kg until the end of the experiment. SeNorm feed contained organic selenium yeast (Selsaf, Lesaffre Feed Additives, Marcq-en-Baroeul, France) 0.32

mg/kg for the first 7 d and then 0.30 mg/kg until the end of the experiment.² The selenium content of the feed was analyzed by the inductively coupled plasma–mass spectrometry (Nordic Committee on Food Analysis Method 161:1998, International Organization for Standardization 17294–2). The feed was based on wheat and soyabean. With the exception of selenium, the levels of other nutrients were adjusted according to recommendations and were similar for both groups.¹

Necropsy and WB diagnosis

Birds were necropsied immediately after death. The necropsy and sampling crew were blind to the dietary treatment. After the evaluation of the skin, head, oral cavity, wings, legs and footpads, the skin was bluntly dissected to reveal the pectoralis major muscles. Both pectoralis major muscles were visually evaluated for color change and white striping (present-absent), and palpated to evaluate the consistency of the muscle (normal, firm (mildly hardened) or markedly hardened (wood-like)). The right pectoralis major muscle was then cut off and a second person graded its consistency by palpation as follows: Grade WB0, normal consistency and macroscopically unaffected (Fig. 1); Grade WB1, hardened consistency and affected with WB (Figs. 2 and 3), including subcategories WB1mild, mildly hardened (firm) consistency and WB1hard, markedly hardened consistency. Additionally, the distribution of the WB lesion was recorded as follows: focal (FOC) included cases with a single hardened lesion either caudally (Fcaud) or cranially (Fcran) within a muscle of otherwise normal consistency (Fig. 2); and diffuse (DIFF) included cases with diffuse or extensive lesions with the presence of very small areas of normal muscle consistency, usually in the middle lateral area, (Fig. 3). When there was divergent scoring (n=3, normal vs. WB1mild), the two persons re-evaluated the cases

together to reach an agreement. The muscle was sampled for histology immediately after grading. The internal organs, as well as muscles of the left thigh and large joints of the left limb were macroscopically evaluated ($n=226$).

Histological evaluation

Of the 226 macroscopically examined birds, 111 were evaluated histologically (WB0 $n=61$; WB1 $n=50$) (Table 2). Our focus was to describe the early development of WB, so all cases at 18 and 24 d of age (the phase of early macroscopic lesions) were histologically evaluated. Additionally, at least 25% of other age groups were randomly chosen for histological evaluation.

For histology, a sample of 1x1x1 cm was collected immediately after the macroscopic grading from the right side pectoralis major muscle. The sample was taken approximately 1 cm deep from the ventral surface of the muscle. The sampling sites were as follows: WB0 muscles were sampled from the middle region, the FOC cases from the focal lesion site, and diffusely affected muscles from the most severely affected region. Of FOC cases, a total of fifteen cases at 18, 24, 35 and 38d were also sampled from an area of normal consistency (non-lesion site) in addition to the lesion site, for the evaluation of intra-individual development of WB. The left biceps femoris muscle of five birds was sampled for histology; two WB0 and two WB1 cases with macroscopically normal biceps femoris muscles, and one WB1 case at 38 d with a hardened biceps femoris muscle. Of the internal organs, the heart, spleen, liver, kidney, lung, gizzard, duodenum and pancreas of twelve birds, one randomly selected WB0 and WB1 bird per each age group, were sampled and evaluated histologically.

The tissue samples were fixed in formalin by immersion, embedded in paraffin, and sectioned at 4- μ m thickness. Muscle samples were oriented transversally and longitudinally. Slides were stained with hematoxylin and eosin. CD3 antigen was detected by immunohistochemistry (polyclonal rabbit anti-human CD3, DakoCytomation, Glostrup, Denmark).

Histologically, the myodegenerative lesions were graded into four categories; DEG0 for absent or minimal, DEG1 for mild, DEG2 for moderate and DEG3 for severe to excessive myodegeneration (Figs. 4-7). Myodegenerative fibers by definition included hyalinized fibers with the loss of cross-striations as well as fragmented fibers that often were infiltrated or surrounded by macrophages or heterophilic granulocytes. Additionally, vasculitis and perivascular cell infiltration was graded into three categories according to the most severely affected vessel: V0 for normal vessel walls without surrounding cell infiltration, V1 for perivascular infiltrations of lymphocytes with or without mild intramural infiltration, and V2 for marked perivascular infiltration of lymphocytes with intramural infiltration sometimes obliterating the vascular wall (Figs. 8-10). The amount of perivascular adipose tissue was graded into four categories: A0 for absent, A1 for mild (thickness of the adipose tissue bed approximately similar to the largest diameter measure of the blood vessel), A2 for moderate ($\leq 2x$ blood vessel diameter), and A3 for marked ($\geq 3x$ blood vessel diameter) amount of adipose tissue. The presence of other histological features, such as regeneration, necrosis, inflammatory cell infiltration, and presence of loose connective tissue or fibrosis was also evaluated. The histological assessment was performed blinded to the macroscopic diagnosis and experimental dietary treatment. A board-certified veterinary pathologist (HKS) performed the necropsy and histological analyses.

Data analysis

Descriptive analyses were performed for the variables. The association between the histological grades and WB or white striping were assessed with Fisher's exact test twice; first only 18 and 24 d groups together, and then including all histologically evaluated birds. The relationships between WB severity and location or white striping were assessed with Fisher's exact test. The association between weight and WB or dietary treatment was assessed by the analysis of variance. Logistic regression analysis was performed to analyze the effects of dietary treatment and age on WB prevalence; WB was the outcome variable. P values less than .05 were regarded as significant. Statistical analyses were performed using IBM SPSS Statistics version 23 (IBM Corp., NY).

Results

Of the 240 birds, 14 (5.8%; 7 for each dietary treatment) died during the experiment, exhibiting signs of sudden death syndrome ($n=7$), ascites syndrome ($n=5$) or other ($n=2$) in the macroscopic post mortem examination.¹⁴ These birds were excluded from the study due to the development of rigor and post mortem changes. In the remaining 226 birds, no significant clinical signs were detected except in one bird with difficulty walking but no gross lesions. WB was not suspected through clinical observation of any of the birds.

Macroscopic Findings

Palpatory hardness typical for WB was not exhibited by any of the excised pectoralis major muscles of the birds at 10 d of age. The first WB1 cases were seen at 18 d, with an age group prevalence of 28% (Table 1). From 24 to 42 d, the WB1

prevalence remained close to or above 50% per group. When divided into the subcategories, the majority of the WB1 birds fell into WB1mild category at 18 d, but at 24, 35 and 38 d over 60% of the affected cases were of WB1hard. The last age group, 42 d, slightly differed from this trend with 18% in WB1hard, but also had a markedly lower total number of cases. In addition to the hardened consistency, the WB lesion was characterized by pale color. Marked discrepancy between the right and left pectoralis major muscles in the presence or severity of the WB lesion were not noted.

The Fcaud location of the macroscopic WB lesion was detected only at 18 and 24 d; the Fcran and DIFF categories were more prevalent in the older age groups (Table 1). The severity of the muscle hardness, WB1mild vs. WB1hard, was associated with the location ($P < .001$); 2/13 cases (15%) of the Fcaud, 12/33 (36%) of the Fcran and 38/47 (81%) of the DIFF were of the WB1hard category.

White striping and WB were significantly associated ($P < .001$) (Table S1). When white striping was present, it was usually seen throughout the pectoralis major and was not restricted to the WB area.

A macroscopic lesion similar to WB was detected in the biceps femoris muscle of the thigh in one bird at 38 d of age; the muscle focally exhibited mildly hardened consistency, whereas the bird had a diffuse WB1hard lesion in the pectoralis major muscle.

Histologic Findings

Myodegeneration was significantly associated with WB ($P < .001$; Table 3, WB0-1 vs. DEG0-3). The myodegeneration grade for the 61 macroscopically unaffected (WB0)

muscles was either DEG0, represented by all age groups, or DEG1 that was seen only at ages 18 and 24 d. The majority of the WB1 cases exhibited DEG2 or DEG3, represented by all affected age groups. DEG0 and DEG1 were more common in the younger age groups of WB1 birds. In the WB1mild subcategory, all myodegeneration grades were rather equally represented, in contrast to the WB1hard, where 21/28 cases (75%) were of DEG3. Myodegeneration was segmental and accompanied by regeneration in all categories and age groups (Figs. 5-7). In a few cases, areas of extensive regeneration were seen (Fig. 11). In cross section, the affected areas showed marked variation in shape and diameter of the fibers (Fig. 12). Some perivascular adipose tissue was present in all histologically evaluated cases. Within the 18 and 24 d old birds, the amount of perivascular adipose tissue was not associated with WB, white striping, myodegeneration or vasculitis grade ($P > .05$ for all variables). However, when all age groups were included, the increased amount of perivascular adipose tissue was weakly associated with WB ($P = .01$) and with myodegeneration ($P = .004$) (Table 3). Vasculitis and the amount of perivascular adipose tissue were not associated ($P > .05$). A few interstitial aggregations of adipose tissue without visible blood vessel bundles were observed in three cases (two WB0 cases and one WB1mild case). Numerous spindle to oval shaped cells arranged in irregular vascular channel-like structures, with erythrocytes in the luminal area occasionally accompanying the myodegenerative changes (Fig. 13). The DEG2 and DEG3 grades were usually accompanied by moderate to marked interstitial edema and loose immature connective tissue, as well as fibrosis in the older age groups (35, 38, 42 d). However, fibrosis was absent in the WB1 birds at 18 and 24 days of age, including the WB1hard cases ($n=8$). Necrotic fibers were rare in general, but a few cases exhibited moderate to excessive numbers of necrotic fibers together

with myodegeneration (Fig.14). Features other than myodegeneration, perivascular inflammatory cell infiltration with vasculitis, and the amount of perivascular adipose tissue were not graded separately because they were either linked to myodegeneration or present in a minority of cases.

Vasculitis and WB were significantly associated ($P < .001$); none of the 61 WB0 cases, but 29/50 (58%) of the WB1 cases exhibited remarkable perivascular infiltration of lymphocytes with intramural infiltration, sometimes obliterating the vascular wall (V2) (Table 3). Vasculitis was especially frequent in the WB1hard category; only one case did not show vasculitis or perivascular lymphocyte infiltration, whereas in 21/28 (75%) cases those lesions were extensive. The ten WB1 cases that did not show any vascular lesions (V0) were either 18 or 24 d of age. Vasculitis scores V1 and V2 were exhibited by all affected age groups. Nine WB0 cases that had perivascular infiltrations of lymphocytes (V1) were of the youngest age groups: 10, 18 and 24 d.

Vasculitis and perivascular infiltrations were also significantly associated with higher myodegenerative scores ($P < .001$); 47/62 (76%) of the V0 cases were DEG0 and 25/29 (86%) of the V2 cases were of DEG3 (Table 3). Even though the majority of the V0 cases were associated with absent or minimal myodegeneration (DEG0), some of the DEG1 and DEG2 cases also lacked vascular changes; all these birds were either 18 or 24 d of age and WB0 or WB1mild macroscopically. All grade DEG3 cases had either perivascular infiltrations of lymphocytes (V1) or also vasculitis (V2). Perivascular infiltration of lymphocytes (V1) was seen with all myodegeneration grades, but extensive infiltrations and vasculitis (V2) were associated mainly with

grades DEG2 and DEG3. One DEG0 case also exhibited V2; this bird was 18 d of age and macroscopically was graded WB1mild.

Overall, vascular lesions and perivascular infiltrations were seen only in veins, typically venules or small veins; arteries did not exhibit significant changes even when the veins were severely affected. The cell infiltration around the veins formed dense cuffs and often was eccentric. Rare plasma cells or histiocytic cells were identified within the lymphocytic infiltration by morphology. Immunohistochemistry for the T cell marker CD3 revealed a strong positive staining of the perivenular lymphocytes in three tested cases with V1 or V2, 10 or 18 d of age. Spleen tissue from the experimental birds was used as a control for the CD3 staining; their T cell regions showed intense positive staining.

The non-lesion sites of the focally affected cases were sampled from 15 cases and exhibited histological myodegeneration scores of DEG0 ($n=1$; lesion site was WB1hard DEG3), DEG1 ($n=7$; lesion site WB1mild or WB1hard, DEG1-3) and DEG2 ($n=7$; lesion site WB1hard, DEG3); none of them exhibited DEG3. Vasculitis and perivascular lymphocyte infiltrations were either absent or mild (V0 $n=8$; V1 $n=6$) in the majority of the non-lesion sites; only one non-lesion site exhibited V2. All non-lesion sites exhibited either A1 or A2 grades of perivascular adipose tissue.

Macroscopic white striping was significantly associated with myodegeneration ($P < .001$) when all birds were included. The association remained significant when it was tested with WB0 cases only; first only the birds of 18 and 24 d of age ($P = .016$) and then all age groups ($P = .011$). Specific histological changes different from WB histology were not recognized for cases with macroscopic white striping.

In the internal organs, no significant pathological changes were seen. In the liver parenchyma and in the lamina propria of the duodenum, occasional aggregates of lymphoid cells were present. Perivascular lymphocytic infiltrations or vasculitis were not exhibited in any of the examined internal organs.

The biceps femoris muscle that had a macroscopic lesion similar to WB was histologically graded as DEG3, V2 and A1. The four histologically assessed but macroscopically normal biceps femoris muscles were of DEG0, V0 and A1 histologically.

Weight and dietary treatment

Mean live weights per age group are displayed in Table 1. The weight between the WB0 and WB1 categories or between WB0, WB1mild and WB1hard was not significantly different ($P > .05$). This was further analyzed by grouping together the birds near the conventional slaughter age (35, 38 and 42 d), but the differences were not significant ($P > .05$).

The dietary treatment SeLow did not have a significant effect on the occurrence of WB; the odds of having WB was 0.78 times lower for broilers that received SeNorm diet compared to SeLow diet (odds ratio 0.78; 95% confidence interval 0.42-1.46; $P = 0.44$) (Table S2). The weight of the birds did not significantly differ between the dietary treatments ($P > .05$) (Table S2).

Discussion

The morphology²⁷ as well as meat quality and processing technological properties^{19, 20} of the skeletal muscle affected with WB have been recently discussed, with focus on birds near slaughter-age. In the present study, we describe the progression of WB

myodegeneration in broilers over the breeding period until 42 d of age, and the associated morphological changes.

WB myopathy started to develop between 10 and 18 d of age. Some of the histological changes exhibited by the WB cases at 18 d, such as prominent regenerative fibers, take 1-2 days to appear, suggesting that the first birds succumb to histologically detectable WB at close to 15 d of age.¹¹ The severity of the palpatory hardness of the pectoralis major muscle, the macroscopic hallmark of the disease, was associated with both the age of the bird and the extent of the lesion. A milder and focal WB lesion was more common among the younger birds, whereas the birds close to the conventional slaughter-age usually had a diffuse and more severe lesion. This indicates that the WB lesion begins focally and spreads towards a diffuse form. This is further supported by the presence of myodegenerative lesions in the macroscopically unaffected (non-lesion) sites of the focally affected cases. The markedly hardened muscle consistency with lack of fibrosis histologically in several cases in the current study further corroborates previous findings that fibrosis is not the sole factor behind the hardening.²⁷

The WB lesion in the biceps femoris muscle of one bird in our study shows that the lesion is not strictly restricted into the pectoralis major muscles only. Anterior latissimus dorsi or other dorsal muscles were not evaluated in the present study and concurrent existence of a dorsal myopathy with WB cannot be excluded.

Histologically, WB presents as a polyphasic myodegeneration throughout the disease course, as the myodegeneration is accompanied with a variable amount of regenerative fibers in all affected age groups. We consider the polyphasic myodegeneration, usually accompanied with lymphocytic phlebitis, as a constitutive

microscopic lesion of WB. Other changes, such as connective tissue accumulation and changes suggestive of neovascularization, are likely to be secondary. Although the diagnosis of WB has until now relied almost entirely on the macroscopic findings, the diagnostic criteria should also include histological myodegeneration in the future, at least in studies of the pathomechanism. However, the microscopic changes alone are not specific for distinguishing WB from other myodegenerative diseases. For example, the presence of myofibers with an increased cross-sectional area (giant fibers), which are typically present in the pectoralis major of fast-growing broilers,⁴ should not be used as a sole diagnostic criterion for WB.

Minimal myodegeneration is regarded as a frequent or even normal feature in the skeletal muscle of the broiler, and also in our study several cases exhibited minimal myodegeneration (DEG0) in the absence of WB.^{10, 25, 28, 30} On the other hand, the DEG1 myodegeneration exhibited by macroscopically unaffected birds at 18 and 24 d of age may represent cases in which the development of WB has started, but is not yet macroscopically detectable. The 18 d prevalence of WB (28%) almost doubled by 24 d and remained rather steady and above 50% thereafter, indicating that many of the macroscopically unaffected cases at 18 d could have developed WB later on and should not be considered as a true unaffected control population.

Vasculitis and perivascular infiltrations of lymphocytes have been previously described in the WB lesions of slaughter-age broilers.²⁷ The present study confirms that similar CD3+ lymphocytic infiltration and vasculitis also affects the veins during the earlier phase of the WB disease. It remains unknown whether the phlebitis is primary or secondary in WB. In young birds, 18 and 24 d of age, DEG1 and DEG2 were seen without vasculitis, but extensive vasculitis without myodegeneration can

also be seen. In general, conditions with lymphocytic vasculitis without arterial or systemic involvement are rare. Enterocolic lymphocytic phlebitis in humans is one of the best described.^{22, 26, 26} It consists of a dense accumulation of lymphocytes around the veins and leads to thrombosis and ischemic lesions in the gut wall; the majority of the perivenular cells are CD3+ T lymphocytes. However, based on our study, thrombosis is not typical in WB and does not appear to be a primary cause of WB.

The potentially insufficient size or density of the capillary network to provide sufficient oxygen for the muscle, as well as remove the byproducts, has been under discussion recently as both the growth rate and the diameter of individual myofibers in the broiler have increased in recent decades.^{8, 13, 24} An increased expression of hypoxia-related genes in WB was also recently reported.²¹ Intriguingly, we found tubular structures reminiscent of neovascularization accompanying the myodegenerative lesions of WB, possibly indicating hypoxia, which is a strong stimulus for the formation of new blood vessels.⁶

The results on the condition known as white striping show that it is usually present when the muscle is affected with WB although it can occur without WB. In our study, different histological patterns could not be distinguished for WB and white striping, but white striping may result in the mild myodegenerative lesion in WB0 cases. The two conditions, WB and white striping, share very similar emergence temporally, both having become increasingly prevalent for at least half a decade now, as well as many similar histological features.^{3, 17, 25} These findings might suggest that white striping and WB are two macroscopic variations of one condition or share similar pathogenesis.

Although clinical signs could not be clearly related to WB by the routine daily observations of the birds, the presence of pain and welfare problems cannot be excluded and subtler studies are necessary. The mortality within the birds of our study was similar to what has been described in previous experiments with the same broiler hybrid, and the causes of death were usual for growing broilers.^{14, 18}

Restriction of dietary selenium did not affect the occurrence of WB in the current study. The choice to use inorganic selenium for the lower dose group and organic for the higher aimed at obtaining a higher bioavailability of the selenium for the higher group than the same concentration of inorganic selenium would have yielded.⁵ We consider this as a preliminary study on the effect of selenium on the WB condition.

In conclusion, the present study describes that WB is a polyphasic myodegeneration and its morphological changes start to develop around two weeks of age in broilers. The usual macroscopic features are hardened consistency of the muscle, pale color and white striping; and lymphocytic phlebitis accompanies the myodegeneration histologically. A mild and focal appearance is more common in the early phase, whereas birds close to the slaughter-age usually exhibit a more severe and diffuse lesion. In the focally affected muscles, mild myodegeneration can be seen histologically also in the macroscopically unaffected areas of the muscle. Fibrosis is not a sole explanation for the hardened muscle consistency in WB, but rather a secondary consequence of chronic myodegeneration, as markedly hardened muscle consistency without fibrosis is seen in the early phase of WB development. In the future, the diagnosis of WB should be based on both macroscopic and histologic findings, especially in birds younger than slaughter-age.

Acknowledgements

The study was supported by HKScan Finland Ltd and Atria Finland Ltd.

References

1. Aviagen. *Ross Broiler Management Handbook*. Newbridge, UK: Aviagen; 2014:11-53, 71-96.
2. Aviagen. Appendices 1-3. In: *Arbor Acres Broiler Nutrition Supplement*. Scotland, UK: Aviagen; 2009:16-19.
3. Bauermeister L, Morey A, Moran E, et al. Occurrence of white striping in chicken breast fillets in relation to broiler size. *Poult Sci*. 2009;88(Suppl. 1):33.
4. Branciarri R, Castellini C, Dal Bosco A, et al. The occurrence of giant fibres in different muscles of two chicken genotypes. *Br Poult Sci*. 2014;55(2):181-185.
5. Briens M, Mercier Y, Rouffineau F, et al. Comparative study of a new organic selenium source v. seleno-yeast and mineral selenium sources on muscle selenium enrichment and selenium digestibility in broiler chickens. *Br J Nutr*. 2013;110(4):617-624.
6. Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature*. 2000;407(6801):249-257.
7. Dam H, Sondergaard E. Prophylactic effect of selenium dioxide against degeneration (white striation) of muscles in chicks. *Experientia*. 1957;13(12):494.

8. Dransfield E, Sosnicki AA. Relationship between muscle growth and poultry meat quality. *Poult Sci.* 1999;78(5):743-746.
9. Hall JA, Van Saun RJ, Bobe G, et al. Organic and inorganic selenium: I. Oral bioavailability in ewes. *J Anim Sci.* 2012;90(2):568-576.
10. Harvey AI Marshall IG. Skeletal Muscle. In: Causey Whittow G ed. *Sturkie's Avian Physiology*, 5th ed. California: Academic Press; 2000:123-139.
11. Hawke TJ, Garry DJ. Myogenic satellite cells: physiology to molecular biology. *J Appl Physiol (1985)*. 2001;91(2):534-551.
12. Huang JQ, Ren FZ, Jiang YY, et al. Selenoproteins protect against avian nutritional muscular dystrophy by metabolizing peroxides and regulating redox/apoptotic signaling. *Free Radic Biol Med.* 2015;83:129-138.
13. Joiner KS, Hamlin GA, Lien AR, et al. Evaluation of capillary and myofiber density in the pectoralis major muscles of rapidly growing, high-yield broiler chickens during increased heat stress. *Avian Dis.* 2014;58(3):377-382.
14. Klasing K, Crespo R, Shivaprasad H, et al. Noninfectious Diseases. In: Saif YM, Fadly AM, Glisson JR, et al eds. *Diseases of Poultry*, 12th ed: Wiley-Blackwell; 2008:1121-1258.
15. Kuttappan VA, Brewer VB, Apple JK, et al. Influence of growth rate on the occurrence of white striping in broiler breast fillets. *Poult Sci.* 2012;91(10):2677-2685.

16. Kuttappan VA, Brewer VB, Mauromoustakos A, et al. Estimation of factors associated with the occurrence of white striping in broiler breast fillets. *Poult Sci.* 2013;92(3):811-819.
17. Kuttappan VA, Shivaprasad HI, Shaw DP, et al. Pathological changes associated with white striping in broiler breast muscles. *Poult Sci.* 2013;92(2):331-338.
18. Lippens M, Room G, De Groote G, et al. Early and temporary quantitative food restriction of broiler chickens. 1. Effects on performance characteristics, mortality and meat quality. *Br Poult Sci.* 2000;41(3):343-354.
19. Mazzoni M, Petracci M, Meluzzi A, et al. Relationship between pectoralis major muscle histology and quality traits of chicken meat. *Poult Sci.* 2015;94(1):123-30.
20. Mudalal S, Lorenzi M, Soglia F, et al. Implications of white striping and wooden breast abnormalities on quality traits of raw and marinated chicken meat. *Animal.* 2015;9(4):728-734.
21. Mutryn MF, Brannick EM, Fu W, et al. Characterization of a novel chicken muscle disorder through differential gene expression and pathway analysis using RNA-sequencing. *BMC Genomics.* 2015;16:399.
22. Ngo N, Chang F. Enterocolic lymphocytic phlebitis: clinicopathologic features and review of the literature. *Arch Pathol Lab Med.* 2007;131(7):1130-1134.
23. Rajashree K, Muthukumar T, Karthikeyan N. Comparative study of the effects of organic selenium on hen performance and productivity of broiler breeders. *Br Poult Sci.* 2014;55(3):367-374.

24. Remignon H, Gardahaut MF, Marche G, et al. Selection for rapid growth increases the number and the size of muscle fibres without changing their typing in chickens. *J Muscle Res Cell Motil.* 1995;16(2):95-102.
25. Russo E, Drigo M, Longoni C, et al. Evaluation of White Striping prevalence and predisposing factors in broilers at slaughter. *Poult Sci.* 2015;94(8):1843-1848.
26. Saraga E, Costa J. Idiopathic Entero-colic Lymphocytic Phlebitis: A Cause of Ischemic Intestinal Necrosis. *Am J Surg Pathol.* 1989;13(4):303-308.
27. Sihvo HK, Immonen K, Puolanne E. Myodegeneration with fibrosis and regeneration in the pectoralis major muscle of broilers. *Vet Pathol.* 2014;51(3):619-623.
28. Soike D, Bergmann V. Comparison of Skeletal Muscle Characteristics in Chicken Bred for Meat or Egg Production. I. Histopathological and Electron Microscopic Examination. *J Vet Med Ser A Physiol Pathol Clin Med.* 1998;45(3):161-167.
29. Van Vleet JF Valentine BA. Muscle and tendon. In: Maxie MG ed. *Jubb, Kennedy, and Palmer's Pathology of Domestic Animals*, 5th ed: Saunders Ltd.; 2007:185-277.
30. Williams S, Fletcher O, Abdul-Aziz T. Muscular System. In: Fletcher O ed. *Avian Histopathology*, 3rd ed. Florida: American Association of Avian Pathologists; 2008:80-95.
31. Zimermann FC, Fallavena LCB, Salle CTP, et al. Downgrading of heavy broiler chicken carcasses due to myodegeneration of the anterior latissimus dorsi: Pathologic and epidemiologic studies. *Avian Dis.* 2012;56(2):418-421.

Figure legends

Figures 1-7. Skeletal muscle, right pectoralis major muscle, broiler chicken.

Figure 1. Unaffected muscle of normal color and consistency. Grade WB0.

Figure 2. Focal manifestation of wooden breast myodegeneration. Hardened consistency and pale color affect the cranial area of the muscle (lined by arrowheads) which is surrounded by unaffected muscle. White stripes follow the muscle fiber orientation (arrow). Grade WB1, location Fcran.

Figure 3. Diffuse wooden breast myodegeneration. The muscle is diffusely hardened and pale. White stripes of variable width are present (arrow). Grade WB1, location DIFF.

Figure 4. Myodegenerative fibers are not present. Typical histology of grade DEG0. Hematoxylin and eosin (HE).

Figure 5. Some myodegenerative fibers are present between the unaffected muscle fibers. Degeneration is accompanied by regenerative fibers. Typical histology of grade DEG1. HE.

Figure 6. Several degenerative and regenerative fibers are present. The myofibers are slightly separated by accumulation of acellular eosinophilic material, loose connective tissue and mild cell infiltration. Typical histology of grade DEG2. HE.

Figure 7. Extensive number of muscle fibers are degenerated, accompanied by regeneration. The interstitial area is expanded by acellular eosinophilic material (asterisk), and loose and organized fibrous tissue. A cell infiltration surrounds a vein (arrow). Typical histology of grade DEG3. HE.

Figures 8-14. Skeletal muscle, pectoralis major muscle, broiler chicken. Illustrations of various grades of vasculitis and additional histologic features occasionally seen in wooden breast. Hematoxylin and eosin (HE).

Figure 8. An artery and a vein without perivascular cell infiltration. Typical presentation of grade V0.

Figure 9. A moderate infiltrate of mononuclear cells surrounds a vein. A few mononuclear cells are present within the vascular wall. Endothelial cells of the vein are reactive. Grade V1.

Figure 10. Extensive infiltrate of mononuclear cells surrounds and penetrates a vein. Grade V2.

Figure 11. Numerous regenerative fibers surround fragments of degenerated myofibers. Cell infiltration and acellular eosinophilic material is present in the interstitium between the regenerative fibers. A cell infiltration surrounds a vein (arrow). Inset: Detail of the regenerative fibers.

Figure 12. The shape and diameter of the myofibers varies from small and angular to enlarged, rounded or hypereosinophilic fibers. Small fibers with increased basophilia and prominent nuclei are also present (regenerative fibers). Loose connective tissue and inflammatory cell infiltrates accumulate in the endo- and perimysium.

Figure 13. Plump spindle cells with an oval nucleus form several loose, irregular tubular structures (arrows) with occasional intraluminal erythrocytes. Homogenous or finely granular eosinophilic acellular material and fibroblasts are intertwined with the tubular structures and the myofibers undergoing degeneration.

Figure 14. Numerous necrotic myofibers, composed of pale eosinophilic fibrillar or fragmented material with loss of the cross-striated architecture, are present. A cellular infiltrate surrounds a vein (arrow).