Early Growth and Adult Health
Programming of postprandial responses, food intake and salt sensitivity

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Individuals who were born with small body size are at increased risk of metabolic diseases, including type 2 diabetes and cardiovascular disease, in adult life. Unhealthy dietary habits are also closely linked with these diseases. This study investigated whether body size at birth is associated with food and nutrient intake later in life and whether birth weight modifies the relationship between salt intake and blood pressure. In addition, the impact of early growth on postprandial metabolism was examined.

Individuals born with small body size consumed lower amounts of fruits and berries, and had lower intake of carbohydrates and higher intake of fat reflecting unhealthier dietary habits. In addition, they were especially sensitive to the blood pressure-raising effect of salt. Therefore, they would especially benefit from the reduction of salt intake. Slow growth during early life had adverse effects on postprandial insulin and triglyceride responses. Unhealthy dietary habits and elevated postprandial responses may partly explain the association between early growth and increased risk for metabolic diseases in adulthood.
RESEARCH 124

Mia-Maria Perälä

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ACADEMIC DISSERTATION

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Helsinki, 2014
To my Family
Abstract


Background. Epidemiologic studies have shown that individuals who were born with small body size are at increased risk of metabolic diseases, including type 2 diabetes and cardiovascular disease, in adult life. Unhealthy dietary habits are also closely linked with these diseases. However, few results are available on whether dietary habits play a role in the association between birth size and disease risk in later life. Small body size at birth or infancy is also closely linked with cardiovascular disease risk factors, such as elevated fasting glucose and cholesterol levels. The association between small body size at birth and elevated fasting levels is surprisingly modest compared with the much stronger association between size at birth and disease risk. Postprandial levels of lipids and glucose have been proposed to be more important than fasting levels in disease process. However, only limited data is available on the long-term influences of early growth on postprandial responses.

Aims. The aim of this thesis was to determine whether body size at birth is associated with food and nutrient intake later in life and whether birth weight modifies the relationship between salt intake and blood pressure. In addition, the impact of early growth on postprandial metabolism was examined.

Subject and methods. The Helsinki Birth Cohort Study comprised 8760 individuals born during 1934–1944 in Helsinki. Of these, 2003 individuals participated in a clinical examination between the years 2001 and 2004. In the clinic, their weight, height and blood pressure were measured and they filled a validated food-frequency questionnaire. Of those who attended the clinical study, 12 obese individuals with a slow increase in body mass index (BMI) during the first year of life and 12 BMI- and age-matched controls were recruited to participate in the postprandial studies between the years 2009 and 2010. Each participant consumed six different test meals in random order. Blood samples were collected during the fasting state and 4-h postprandially. Glucose, insulin, lipids, incretins and appetite regulatory hormones were measured.

Results. Body size at birth was positively associated with consumption of fruits and berries and intake of carbohydrates, sugars and fibre. An inverse association between size at birth and fat intake was also observed. When participants were
divided into groups for the best obtained breakpoint to birth weight, it was observed that salt intake was related to systolic blood pressure among participants whose birth weight was ≤ 3050 g but not among participants whose birth weight was > 3050 g. Among low birth weight participants, a 1-g increase in salt intake was associated with a 2.48-mmHg higher systolic blood pressure. Salt intake was not significantly associated with diastolic blood pressure, either in the low birth weight or high birth weight groups. Early growth affected the postprandial responses and insulin and triglyceride responses were significantly higher in the group that grew slowly during early life than in the controls. Individuals with slow early growth also showed higher appetite regulatory hormone peptide YY responses than did the controls.

**Conclusions.** This study showed that individuals born with small body size may be programmed towards unhealthy dietary habits. In addition, they are sensitive to the blood pressure-raising effect of salt and therefore, may especially benefit from a reduction in salt intake. Slow growth during early life adversely affects postprandial insulin and triglyceride responses. Unhealthy dietary habits and elevated postprandial responses may be one underlying mechanism explaining the increased risk of metabolic diseases associated with nonoptimal early growth. Early growth may also alter appetite regulatory hormone secretion, which could be one explanation why individuals born small or who grow slowly during infancy are unlikely to become obese in later life.

Keywords: birth weight, early growth, DOHaD, food intake, salt intake, blood pressure, postprandial responses
Tiivistelmä


Tavoitteet. Tämän väitöskirjatyön tarkoituksena oli tutkia syntymäkoon yhteyttä aikuisiän ravinnonsaantiin ja selvittää, selittääkö syntymäkoko suolan saannin ja verenpaineen välistä yhteyttä. Lisäksi tutkittiin varhaiskasvun vaikutusta aterianjälkeisiin aineenvaihdunnan vasteisiin ylipainoisilla aikuisilla.

Tulokset. Syntymäkoko oli yhteydessä runsaampaan hedelmien ja marjojen kulutukseen. Lisäksi syntymäkoko oli yhteydessä pienempään rasvan saantiin ja suurempaan hiilihdraattiihin, sokerin ja kuidun saantiin. Kun tutkittavat jaettiin syntymäkoon perusteella ryhmiin parhaamman katkaisukohdan mukaan, havaittiin, että suolan saanti oli yhteydessä systoliseen verenpaineeseen ainoastaan henkilöillä, joiden syntymäraino oli ≤ 3050 g. Heillä 1 g suurempi suolan saanti oli yhteydessä 2.48 mmHg korkeampaan systoliseen verenpaineeseen. Vastaavanlaista suolan saannin ja verenpaineen välistä yhteyttä ei havaittu suurempikokosiina syntyneillä (> 3050 g). Suolan saanti ei ollut yhteydessä diastoliseen verenpaineeseen pienipainoisina tai suurempikokosiina syntyneillä. Henkilöillä, jotka olivat kasvaneet hitaasti ensimmäisen elinvuoden aikana, oli kontrolliryhmään verrattuna suuremmat aterianjälkeiset triglyseridi- ja insuliinivasteet sekä kylläisyyttä säätelevän hormonin peptidi YY:n pitoisuudet.


Avainsanat: syntymäkoko, varhainen kasvu, elämänkaarinaikokäytäntö, ravinnonsaanti, suola, verenpaine, aterianjälkeiset vasteet
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List of original publications

This thesis is based on the following original articles referred to in the text by their Roman numerals. In addition, some unpublished material is presented.


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### Abbreviations

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<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tbody>
<tr>
<td>ANOVA</td>
<td>Analysis of variance</td>
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<tr>
<td>BMI</td>
<td>Body mass index</td>
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<td>CCK</td>
<td>Cholecystokinin</td>
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<tr>
<td>CI</td>
<td>Confidence interval</td>
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<tr>
<td>CV</td>
<td>Coefficient of variation</td>
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<tr>
<td>CVD</td>
<td>Cardiovascular disease</td>
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<td>DBP</td>
<td>Diastolic blood pressure</td>
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<td>DNA</td>
<td>Deoxyribonucleic acid</td>
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<td>DOHaD</td>
<td>Developmental Origins of Health and Disease</td>
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<tr>
<td>DPP</td>
<td>Dipeptidyl peptidase</td>
</tr>
<tr>
<td>E%</td>
<td>Percentage of total energy intake</td>
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<tr>
<td>FF-meal</td>
<td>Fast-food meal</td>
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<td>FFA</td>
<td>Free fatty acids</td>
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<td>FFQ</td>
<td>Food-frequency questionnaire</td>
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<td>GI</td>
<td>Glycaemic index</td>
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<tr>
<td>GIP</td>
<td>Glucose-dependent insulinitropic peptide</td>
</tr>
<tr>
<td>GL</td>
<td>Glycaemic load</td>
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<tr>
<td>GLP-1</td>
<td>Glucagon-like peptide 1</td>
</tr>
<tr>
<td>HBCS</td>
<td>Helsinki Birth Cohort Study</td>
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<tr>
<td>HPA-axis</td>
<td>Hypothalamus-pituitary-adrenal axis</td>
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<td>IAOC</td>
<td>Incremental area over the curve</td>
</tr>
<tr>
<td>Abbreviation</td>
<td>Description</td>
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<tr>
<td>IAUC</td>
<td>Incremental area under the curve</td>
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<tr>
<td>MUFA</td>
<td>Monounsaturated fatty acid</td>
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<tr>
<td>OGTT</td>
<td>Oral glucose tolerance test</td>
</tr>
<tr>
<td>PI</td>
<td>Ponderal index</td>
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<tr>
<td>PP</td>
<td>Pancreatic polypeptide</td>
</tr>
<tr>
<td>PUFA</td>
<td>Polyunsaturated fatty acid</td>
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<td>PYY</td>
<td>Peptide YY</td>
</tr>
<tr>
<td>REC-meal</td>
<td>Meal macronutrient composition according to dietary guidelines</td>
</tr>
<tr>
<td>SBP</td>
<td>Systolic blood pressure</td>
</tr>
<tr>
<td>SD</td>
<td>Standard deviation</td>
</tr>
<tr>
<td>SES</td>
<td>Socioeconomic status</td>
</tr>
<tr>
<td>SFA</td>
<td>Saturated fatty acid</td>
</tr>
<tr>
<td>SGI</td>
<td>Group with slow growth during infancy</td>
</tr>
<tr>
<td>TG</td>
<td>Triglycerides</td>
</tr>
<tr>
<td>totAUC</td>
<td>Total area under the curve</td>
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<tr>
<td>VAS</td>
<td>Visual analogue scale</td>
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<td>WHO</td>
<td>World Health Organization</td>
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1 Introduction

Metabolic diseases, such as type 2 diabetes and cardiovascular disease (CVD), are common health problems, both in the developing as well as in developed countries, including Finland. Genetic and environmental factors, such as unhealthy dietary habits and physical inactivity, play key roles in the development of these disorders. There is strong epidemiological evidence that in addition to genetic and environmental factors, small body size at birth is also related to increased risk for the development of metabolic diseases (Barker 1995, Barker et al. 2005, Eriksson et al. 2007, Huxley et al. 2007, Mu et al. 2012, Osmond et al. 2007, van Abeelen et al. 2011).

The importance of prenatal growth on later health outcomes was demonstrated by David Barker and his colleagues almost 30 years ago (Barker and Osmond 1986). Based on animal findings as well as epidemiological observations, Barker formulated a theory that events occurring in early life could have long-term effects on health later in life. Since then, numerous epidemiological studies have been published, demonstrating that small body size at birth is associated with increased risk of chronic diseases later in life. The concept of long-term consequence of prenatal growth in later health is now widely accepted; e.g. the World Health Organization (WHO) acknowledged the importance of birth weight in preventing chronic diseases (World Health Organization 2011).

Even though dietary factors are well-known risk factors for metabolic diseases, few studies have focused on whether dietary factors interact with birth weight in predicting later health. Body size at birth has been found to modify the effect of some previously known risk factors for metabolic diseases, including lipid responses to dietary fat (Robinson et al. 2006) and the protective effect of dietary omega-3 fatty acids on carotid intima-media thickness (Skilton et al. 2013) as well as the protective effect of exercise on glucose tolerance (Eriksson et al. 2004).

Although epidemiological studies have shown that small body size at birth is closely linked with CVD risk factors, including elevated fasting glucose and cholesterol levels, the associations between small body size at birth and elevated fasting levels are surprisingly modest, compared with the association between size at birth and overall disease risk. With the exception of a few hours in the morning, we spend most of our waking hours in a postprandial state. Postprandial levels of lipids are also better predictors for the risk of development of CVD than fasting levels. Only a handful of studies (Byrne et al. 1997, Kensara et al. 2006, Schou et al. 2005) have examined postprandial responses in this context and, therefore, it is important to explore the long-term influences of early growth on postprandial responses.
The aim of this thesis was to focus on whether prenatal growth is associated with food intake and the relationship between salt intake and blood pressure and explore the effect of early growth on postprandial metabolism in adulthood.
2 Review of the literature

2.1 Developmental Origins of Health and Disease (DOHaD)

2.1.1 The DOHaD hypothesis

It was shown 80 years ago that death rates were more dependent on the decade of birth than on the decade of death (Kermack et al. 1934). Therefore, the authors proposed that living conditions during childhood explained mortality better than current circumstances. Forsdahl (1977) also observed in Norway that infant mortality rates were positively associated with mortality from CVD later in life. The association was so marked that he proposed that poor living conditions during early life followed by prosperity in adulthood are potential risk factors for CVD. In addition to mortality, Wadsworth and colleagues reported in 1985 that birth weight was inversely related to adult-life blood pressure (Wadsworth et al. 1985). Ravelli et al. (1976) observed that famine during the last trimester of pregnancy and the first months of life produced lower obesity rates, whereas famine during the first half of pregnancy resulted in higher obesity rates.

The importance of prenatal growth was also demonstrated by Barker and his colleagues in the 1980s who showed that there was a strong association between infant mortality rates and ischaemic heart disease, as well as an association between birth weight and rates of adult death from ischaemic heart disease (Barker and Osmond 1986, Barker et al. 1989). To explain these findings, they introduced a theory that fetal growth is associated with a number of chronic conditions later in life. The hypothesis is known as ‘Barker’s theory’ and ‘the Developmental Origins of Health and Disease (DOHaD) hypothesis’.

The theory was at first criticized by those who believed that recall bias, publication bias, socioeconomic conditions or lifestyles largely explained these findings (Huxley et al. 2002, Huxley 2006, Kramer and Joseph 1996). However, since the findings of Barker et al. (Barker and Osmond 1986, Barker et al. 1989), several epidemiological studies have been published and to date there is strong epidemiological evidence that low birth weight is related to an increased risk for the development of metabolic diseases, including type 2 diabetes (Johnson and Schoeni 2011, McNamara et al. 2012, Whincup et al. 2008), metabolic syndrome (Fall et al. 2008, Xiao et al. 2010) and CVD (Barker 1995, Barker et al. 2005, Eriksson et al. 2007, Huxley et al. 2007, Johnson and Schoeni 2011, Mu et al. 2012, Osmond et al. 2007, Stuart et al. 2013, van Abeelen et al. 2011). In addition to metabolic diseases, birth size is also linked with several other diseases, such as cancer (Xu et al. 2009),...
Review of the literature

osteoporosis (Martinez-Mesa et al. 2012), lung functioning (Lawlor et al. 2005) and asthma (Johnson and Schoeni 2011) in adulthood. The DOHaD field has also emerged into the mental health area, and associations between low birth weight and cognitive abilities (Rääkkönen et al. 2009, Shenkin et al. 2004), personality (Lahti et al. 2008), temperament (Pesonen et al. 2006), disruptive behaviour disorders (Latimer et al. 2012) and psychiatric symptoms (Indredavik et al. 2004, Lund et al. 2012, Monfils Gustafsson et al. 2009) have been described.

The WHO defined low birth weight as that under 2500 g (World Health Organization Expert Commitee on Physical Status 1995). However, the above associations have also been observed within the normal birth weight range. In addition, associations of birth weight with disease risk in later life are not dependent on gestational age, even though the duration of gestation has a major impact on birth weight. Recent studies have proposed that not only low birth weight but also high birth weight are associated with some of these diseases, suggesting that the relationship between birth weight and later health is U-shaped (Dabelea et al. 1999, Eriksson et al. 2003, Harder et al. 2007).

In addition to prenatal life, early infancy is also a critical developmental period. Growth is rapid during infancy and development of tissues and organs, such as brain and liver, continue during this period. The importance of postnatal growth has been shown in a number of epidemiological studies which have shown that those who have experienced retarded growth in postnatal life have the highest risk of developing type 2 diabetes (Eriksson et al. 2003, Eriksson et al. 2006, Phillips et al. 2005), metabolic syndrome (Salonen et al. 2009a, Salonen et al. 2009b) and CVD in adulthood (Barker et al. 2005, Eriksson et al. 2007, Osmond et al. 2007). However, some studies have observed that accelerated growth during infancy is associated with increased risk of chronic disease in later life (Ibanez et al. 2006, Monasta et al. 2010, Salvi et al. 2012, Sonnenschein-van der Voort et al. 2012, Tzoulaki et al. 2010). Even though both retarded and accelerated growth during infancy is related to disease risk, there is no doubt that the first 1000 days after conception (gestation and first 2 postnatal years) is a critical time for programming of health in later life.

2.1.2 Factors influencing early growth

Several anthropometric measurements have been used to estimate prenatal growth. Birth weight is the most frequently used in epidemiological studies, because it is easy to measure and has been measured systematically for decades. It is, however, a crude indicator of prenatal growth. In addition to birth weight, birth length has also been used to measure prenatal growth. Length measurement contains a large amount of error which is partly explained by the activity level of the infants as well as difficulty in extending the infant’s leg completely due to natural flexion (Johnson et
Therefore, birth length is a less reliable measure than birth weight (Johnson et al. 1997). In addition, the ponderal index (PI, kg/m^3) and body mass index (BMI, kg/m^2), both of which indicate thinness at birth, can be calculated from birth weight and length. Other commonly used measures of prenatal growth include head circumference and placental size.

Both genetic and environmental factors affect prenatal growth (Kramer 1987, Ohlsson and Shah 2008). Therefore, not all babies who are small at birth have experienced growth retardation, but they may be genetically small. Some major factors associated with birth size are presented in Table 1. Genetic factors include sex of the fetus and genes that are passed on from the father and mother to the fetus. For example, maternal genes affect the size of the mother, as well as the mother’s capability of carrying a pregnancy. Environmental factors include maternal nutrition (mostly energy and protein intake), socioeconomic status (SES), endocrine factors, parity, stress, smoking, infections and placental functioning. Body size at birth is also largely dependent on the duration of gestation. It has been estimated that the fetus’ genes explain only 30–40% of birth weight and length, whereas environmental factors that are not related to the fetus’ genes determine 60–70% of the resulting birth weight (Clausson et al. 2000, Lunde et al. 2007).

In addition to prenatal growth, several factors also influence postnatal growth (Table 1). Studies have shown that although parity and placental weight influence birth weight and prenatal growth, they have little effect on postnatal growth (Hindmarsh et al. 2008). Postnatal growth is predominantly influenced by nutrition. In addition, genes affect postnatal growth more than prenatal growth (Pietiläinen et al. 2002). Some other factors that also affect postnatal growth include gestational age, size at birth and parental SES (Hindmarsh et al. 2008, Regnault et al. 2010).
Table 1. Factors influencing prenatal growth and birth weight, as well as postnatal growth (Hindmarsh et al. 2008, Kramer 1987, Ohlsson and Shah 2008, Regnault et al. 2010).

<table>
<thead>
<tr>
<th>Prenatal growth and birth weight</th>
<th>Postnatal growth</th>
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<tr>
<td>Genetic factors</td>
<td>Genetic factors</td>
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<tr>
<td>Maternal and paternal size</td>
<td>Maternal and paternal size</td>
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<td>Infant sex</td>
<td>Infant sex</td>
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<tr>
<td>Gestational age</td>
<td>Birth weight</td>
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<tr>
<td>Demographic factors</td>
<td>Gestational age</td>
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<tr>
<td>Maternal age and stress</td>
<td>Demographic factors</td>
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<tr>
<td>Parental SES</td>
<td>Parental SES</td>
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<tr>
<td>Nutritional factors</td>
<td>Nutritional factors</td>
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<tr>
<td>Gestational weight gain</td>
<td>Gestational weight gain</td>
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<tr>
<td>Energy and nutrient intake</td>
<td>Energy and nutrient intake</td>
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<tr>
<td>Physical activity</td>
<td>Breastfeeding</td>
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<td>Maternal morbidity</td>
<td>Infant morbidity</td>
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<tr>
<td>General morbidity</td>
<td>General morbidity</td>
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<tr>
<td>Infections</td>
<td>Infections</td>
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<tr>
<td>Gestational diabetes</td>
<td>Toxic exposure</td>
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<td>Pre-eclampsia</td>
<td>Cigarette smoking in pregnancy</td>
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<td>Obstetric factors</td>
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<td>Parity</td>
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<td>Maternal constraint</td>
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<td>Toxic exposure</td>
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<td>Cigarette smoking</td>
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<td>Alcohol consumption</td>
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<tr>
<td>Placental abnormalities</td>
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<td>Reduced blood flow</td>
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SES, socioeconomic status.
2.1.3 Mechanisms of programming

It has been proposed that during prenatal life and early postnatal life, a suboptimal environment could permanently influence organ development and functioning and thus has long-lasting influences on health. The process by which the early environment influences metabolic or endocrine changes in later life is called ‘programming’ (Lucas 1998). Changes in the early life environment may induce adaptations to ensure nutrient supply to the most vital organs, such as the brain, at the expense of other organs, such as liver, muscle and pancreas. These adaptations result in lifelong alterations in the structure and functioning of organs and may result in adverse health outcomes in adult life (Figure 1). For example, malnutrition during gestation may affect liver size (Winick and Noble 1966) and its microstructure (Burns et al. 1997) and functioning, such as changes in lipid metabolism (Cong et al. 2012, Lane et al. 2001, Liu et al. 2013) and liver damage (Fraser et al. 2008, Nobili et al. 2007). Growth retardation during prenatal or postnatal life also causes alterations in muscle mass (Eriksson et al. 2002) and muscle structure and functioning (Jensen et al. 2007, Jensen et al. 2008, Taylor et al. 1995), as well as alterations in adipose tissue by favouring abdominal fat accumulation and by changing its metabolic and hormonal functions (Boiko et al. 2005, Maiorana et al. 2007). In addition, suboptimal environments during early life may lead to structural changes in the brain (Coupe et al. 2010, Plagemann et al. 2000a) and lifelong changes in the functioning of different hormonal axes, including the hypothalamus-pituitary-adrenal axis (HPA-axis) and insulin-like growth factor system (Barker et al. 1993, Phillips et al. 1998, Reynolds 2013).
Based on animal models, convincing evidence suggests that epigenetic events serve as a memory of exposure in early life and thus mediate developmental programming by causing alterations in tissue-specific gene expression, as reported in a review by Attig et al. (2010) and Portha et al. (2013). These changes may further cause structural and regulatory effects on many organ systems and thus lead to disease in later life (Figure 1). Epigenetic modifications do not alter the heritable deoxyribonucleic acid (DNA) sequence but do affect gene expression by causing alterations to DNA or chromatin (Callinan and Feinberg 2006). Epigenetic modification may involve DNA methylation, in particular promoter regions of specific genes, or histone modification, such as acetylation (reviewed in Callinan and Feinberg 2006). In general, methylated genes are silenced and hypomethylated genes are induced. For example, maternal protein restriction during gestation reduces DNA methylation of the glucocorticoid receptor and peroxisomal
proliferator-activated receptor alpha genes in the offspring, and thus increases the expression of these genes and further affects protein functioning (Lillycrop et al. 2005). It has also been proposed that one of the mechanisms by which prenatal or postnatal environments may affect developmental programming is mitochondrial DNA methylation (McConnell and Petrie 2004). However, further studies are still needed in this area to understand how the early life environment causes these epigenetic modifications.

Early programming may be beneficial for survival under poor nutritional conditions. The long-term consequences may, however, be especially harmful if the postnatal environment differs from that predicted by the prenatal conditions. This has been demonstrated in several studies that individuals who have experienced retarded growth during prenatal and postnatal life and accelerated childhood weight gain are at increased risk of CVD and other metabolic diseases in adult life (Barker et al. 2005, Eriksson et al. 2003, Eriksson et al. 2006, Eriksson et al. 2007, Osmond et al. 2007, Phillips et al. 2005, Salonen et al. 2009a, Salonen et al. 2009b). In contrast, individuals who have experienced both prenatal and postnatal growth retardation, not followed by high nutrient intake in later life, do not have increased risk of developing these diseases (Stanner and Yudkin 2001).

The timing of food or nutrient restriction in pregnancy may also be critical. Findings from Dutch famine studies have shown that individuals exposed to malnutrition during early gestation have increased risk of many chronic diseases later in life, whereas those exposed to malnutrition during late gestation had lower risk (de Rooij et al. 2006, Ravelli et al. 1998).

### 2.2 Programming of food intake and appetite

#### 2.2.1 Factors affecting food choices and food intake

It is well known that diet plays an important role in the development of metabolic diseases. In addition to traditional nutrition recommendations that include recommendations for single nutrient intakes, food-based dietary guidelines describing how to choose food-items that both include the optimal amount of nutrients and are related to a decreased risk of metabolic diseases have been published (National Nutrition Council 2005, Nordic Council of Ministers 2013). These guidelines suggest that the diet should contain high amounts of vegetables, fruits and berries, nuts and wholegrain cereals. In addition, it is recommended to regularly eat fish and seafood and use vegetable oils, soft fats and low-fat dairy products. Table 2 summarizes the recommendations for healthy food choices.
Table 2. Guidelines for healthy food choices and recommended macronutrient intake according to the Nordic Nutrition Recommendation 2012 (Nordic Council of Ministers 2013).

<table>
<thead>
<tr>
<th>Increase consumption</th>
<th>Decrease consumption</th>
<th>Macronutrient intake</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vegetables</td>
<td>Red and processed meat</td>
<td>Carbohydrates 45–60 E%</td>
</tr>
<tr>
<td>Fruits and berries</td>
<td>Drinks with added sugar</td>
<td>Sucrose &lt; 10 E%</td>
</tr>
<tr>
<td>Nuts and seeds</td>
<td>Food products with added sugar</td>
<td>Fibre 25–35 g/d</td>
</tr>
<tr>
<td>Wholegrain cereals</td>
<td>Food products with added fat</td>
<td>Protein 10–20 E%</td>
</tr>
<tr>
<td>Fish and seafood</td>
<td>Salt and food products with added salt</td>
<td>Protein 10–20 E%</td>
</tr>
<tr>
<td>Alcohol</td>
<td></td>
<td>Total fat 25–40 E%</td>
</tr>
</tbody>
</table>

E%, percentage of total energy intake; SFA, saturated fatty acids; PUFA, polyunsaturated fatty acids; MUFA, monounsaturated fatty acids.

Although there are guidelines for healthy food choices, knowledge of what people should eat does not always affect what individuals will eat. Food choices are influenced by many factors, most importantly food availability, as reported in a review by Blundell et al. (2010). Figure 2 summarizes other important factors that influence food choices and food intakes. Briefly, physiologic factors, which include neural and hormonal signals, mostly affect how much people eat. Peripheral released physiologic factors may also affect the food reward system and thus food choices (Skibicka and Dickson 2011). Sensory factors, such as food taste, smell and texture, influence the liking of different foods and thus mostly affect what people eat but they also affect how much people eat. Environmental factors, including physiological state (e.g. thirst), psychological state (e.g. mood), social consequences and culture, as well as genes, interact with physiologic and sensory factors and thus affect food choices and food intakes (Mela 2001).
2.2.2 Appetite regulatory hormones

Obesity is the result of an imbalance between energy intake and energy expenditure. The regulation of energy balance is very complex; both genetic and environmental factors affect it (Guyenet and Schwartz 2012). The regulation of energy intake is also a complex process involving environmental and behavioural factors, feelings of hunger and fullness, and physiologic factors such as appetite regulatory hormones (Blundell et al. 2010). Although physiologic factors regulate appetite and hunger and thus energy intake, meals are mostly initiated by factors that are not based on energy needs, such as food availability, habit, time of day and social conventions (Woods and D'Alessio 2008). Physiological factors mainly control how much is consumed once a meal begins (Blundell et al. 2010). It has, however, been proposed that appetite regulatory hormones play a role in the development of obesity. Several studies have supported this by showing alterations in appetite regulatory hormones in obese individuals, as reported in a review by Suzuki et al. (2010).

Many peripherally released appetite regulatory hormones have been identified. In this thesis, the focus is on peptide YY (PYY), glucagon-like peptide 1 (GLP-1) and ghrelin (Table 3). Both PYY and GLP-1 decrease appetite and thus food intake, whereas ghrelin stimulates appetite and food intake. PYY is produced by the endocrine L-cells in the ileum and colon in response to food intake (Suzuki et al.
The PYY (1-36) released is rapidly metabolized by dipeptidyl peptidase 4 (DPP-4) to the active form, PYY (3-36). In addition to decreasing appetite, PYY has many other functions. For example, it inhibits insulin secretion and stimulates ileal break which means inhibition of upper gastrointestinal motility and gastric emptying by ingested food, thus restraining the rate of nutrient entry into the blood (Spiller et al. 1984).

Table 3. Summary of the characteristics of the appetite regulatory hormones.

<table>
<thead>
<tr>
<th>Hormone</th>
<th>Primary site of production</th>
<th>Effect on appetite</th>
<th>Major target functions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ghrelin</td>
<td>X/A-like cells, gastric mucosa</td>
<td>Increase</td>
<td>Growth hormone release ↑; Gastrointestinal motility ↑; Insulin ↓</td>
</tr>
<tr>
<td>GLP-1</td>
<td>L-cells, ileum and colon</td>
<td>Decrease</td>
<td>Insulin ↑; Glucagon release ↓; Gastrointestinal motility ↓; Ileal break ↑</td>
</tr>
<tr>
<td>PYY</td>
<td>L-cells, ileum and colon</td>
<td>Decrease</td>
<td>Insulin ↓; Ileal break ↑</td>
</tr>
</tbody>
</table>

Like PYY, GLP-1 is produced by the endocrine L-cells in the ileum and colon and delays gastric emptying, contributing to ileal break. DPP-4 rapidly inactivates secreted GLP-1. GLP-1 is an incretin hormone. Incretins stimulate insulin secretion in response to a meal or glucose ingestion. This so-called incretin effect accounts for approximately 50–70% of the total insulin secretion after a meal (Kazakos 2011). GLP-1 also inhibits glucagon secretion and increases β-cell growth (Cummings and Overduin 2007). Orexigenic ghrelin is released from the gastric mucosa. In addition to its short-term effect on appetite regulation and food intake, it also affects long-term body weight regulation (Cummings and Overduin 2007). In contrast to PYY and GLP-1, ghrelin increases gastrointestinal motility. It also decreases insulin secretion and stimulates the release of growth hormone (Hosoda et al. 2006). In addition to these peptides, other commonly measured appetite regulatory hormones include small intestine-released cholecystokinin (CCK) and pancreas-secreted pancreatic polypeptide (PP), both of which decrease food intake in short-term periods (Badman and Flier 2005).

Appetite regulatory hormones transmit information relating to energy stores or recent energy intake to the hypothalamus and the hindbrain (Figure 3). These peripheral released appetite regulatory hormones can transfer information to the central nervous system through the vagus nerve to the hindbrain or directly to the hypothalamus or hindbrain. In the arcuate nucleus of the hypothalamus, there are two particularly important categories of neurons: appetite-stimulating neurons,
which contain neuropeptide Y and agouti-related peptides and appetite-inhibiting neurons, which contain pro-opiomelanocortin peptide and cocaine- and amphetamine-stimulated transcript peptide. This area has been extensively studied and reviewed in the past decade (Badman and Flier 2005, Cummings and Overduin 2007, Suzuki et al. 2010, Vincent et al. 2008).

**Figure 3.** Simplified figure of physiologic factors affecting short-term appetite regulation. Ghrelin is released from the stomach preprandially and stimulates appetite through the central nervous system via circulation (to the hypothalamus) or vagus nerve. Peptide YY (PYY), glucagon-like peptide 1 (GLP-1) and cholecystokinin (CCK) are released the intestine postprandially and reduce appetite and food intake through signals to the hypothalamus, hindbrain and vagus nerve. Pancreatic polypeptide (PP) is released from the pancreas postprandially and reduces appetite and food intake through signals to the hindbrain or vagus nerve. Modified from Vincent et al. (2008).
2.2.3 Body size at birth and food intake

There is strong evidence that lifestyle factors such as unhealthy dietary habits and physical inactivity are independent and modifiable risk factors for CVD and type 2 diabetes (Ignarro et al. 2007, Rees et al. 2013, Schellenberg et al. 2013). Therefore, it has been proposed that one possible mechanism behind the association between small body size at birth and increased risk for development of chronic diseases could be by early programming of these lifestyle factors (Portella et al. 2012). Animal models (Bellinger et al. 2006, Vickers et al. 2003) and meta-analysis of adult individuals (Andersen et al. 2009) support this hypothesis by showing that low birth weight is related to physical inactivity. Thus, early growth may influence disease risk in later life through direct biological effects as well as modifying adult behaviour.

In addition, there is evidence from animal studies that the early environment may program dietary habits. One of these studies showed that rats whose mothers were fed a low-protein diet during gestation had a preference for a high-fat diet (Bellinger et al. 2004); however, not all studies support this finding (Bellinger and Langley-Evans 2005, Cambraia et al. 2001). Fat intake also increased with decreasing birth weight in young children (Shultis et al. 2005, Stafford and Lucas 1998). Similarly, two different Dutch study groups observed that famine during prenatal life was related to preference for a high-fat diet in adulthood (Lussana et al. 2008, Stein et al. 2009). However, only one of these studies examined the association between birth weight and macronutrient intake in adult life and found no relationships (Lussana et al. 2008). Contrasting results were reported by Barbieri et al. (2009), who showed that in young Brazilian women, severe intrauterine growth restriction was related to higher intake of carbohydrates. They also studied prenatal growth restriction and food intake and found no effect of intrauterine growth restriction on food consumption. A recently published study, in which birth weight and food intake in later life were also investigated, showed that young Finnish adults who were born preterm at below 1500 g had similar macronutrient intake compared with term controls although they consumed less fruits, berries, vegetables and milk products (Kaseva et al. 2013).

What are the underlying mechanisms explaining these altered food choices? It has been proposed that the early life environment may alter physiologic factors that are related to food choices. Animal models have shown that a low-protein diet during gestation may alter the size and neuronal density of hypothalamic structures and expression of neuropeptides that are involved in the regulation of food intake (Breton et al. 2009, Erhuma et al. 2007, Plagemann et al. 2000a). These peptides, such as neuropeptide Y, may also control macronutrient selection behaviour.
Programming of dietary related risk factors by early growth (Primeaux et al. 2006, Smith et al. 1997, Tanaka and Kido 2008). However, since these results are based on animal models, it is still unknown whether these peptides play a role in macronutrient selection behaviour in humans as well. In addition, prenatal flavour experiences may enhance the acceptance and enjoyment of similarly flavoured foods during postnatal life. This was detected in an animal model (Bayol et al. 2007) and in a study of young children (Mennella et al. 2001) in which maternal diet during pregnancy influenced postnatal preference for the same diet.

Early growth may also affect sensory factors that are related to food choices. Subjects who have elevated leptin levels need higher concentrations of sweeteners, such as glucose, to detect the sweet stimulus, than subjects with lower leptin levels (Nakamura et al. 2008, Umabiki et al. 2010). It has been proposed that this type of alteration in detecting the sweet stimulus may lead to a reduced consumption of sweet food (Shigemura et al. 2004). Increased leptin secretion has been observed among participants who were born with low birth weight (Lissner et al. 1999, Phillips et al. 1999). Thus, altered leptin level could potentially be involved in food consumption. Early growth may also affect preference for salty foods as observed in one experimental study of young infants whose birth weight was inversely related to salty taste preference. The authors suggested that preference for salty foods among low birth weight individuals could lead to increased salt intake and subsequently elevated blood pressure (Stein et al. 2006). Another possible mechanism by which early growth could influence food choices later in life is programming of the sensitivity to the food reward, such as pleasure, that is associated with the consumption of palatable food. One recently published study supports this by showing that intrauterine growth retardation was associated with the consumption of palatable foods in preterm infants (Ayres et al. 2012).

2.2.4 Early growth and appetite regulation

Although low birth weight and small body size in infancy are related to increased risk for developing metabolic diseases in later life, large body size at birth and rapid growth during infancy are primarily associated with subsequent risk of obesity, as reported in systematic reviews and meta-analyses (Baird et al. 2005, Monasta et al. 2010, Monteiro and Victora 2005, Ong and Loos 2006, Schellong et al. 2012, Yu et al. 2011b, Zhao et al. 2012). Therefore, it has been suggested that growth retardation during the prenatal or postnatal periods may produce changes in the central control of appetite, such as altering appetite regulatory hormone secretions, and thus affect the risk for developing obesity. Consistent with this hypothesis, animal models have shown that both prenatal and postnatal growth retardation cause alterations in appetite regulation pathways, including alterations in hypothalamic nuclei structure and neuropeptide expression (Breton et al. 2009, Coupe et al. 2009, Desai et al.
Review of the literature


The evidence for early growth and appetite regulation systems in humans comes mostly from studies of young children. These studies indicate that infants who were born with low birth weight (Chen et al. 2012, Siahanidou et al. 2005) or born preterm (Berseth et al. 1992, Chen et al. 2012, Siahanidou et al. 2005, Siahanidou et al. 2007) have elevated levels of fasting PYY reflecting greater satiety. In addition, decreased fasting ghrelin concentrations have been reported in young children who were born with low birth weight (Siahanidou et al. 2005) and elevated ghrelin levels in young adolescents with rapid growth during infancy (Larnkjaer et al. 2010). However, not all studies support these findings (Chen et al. 2012, Darendeliler et al. 2009, Iniguez et al. 2002, Kyriakakou et al. 2009, Park 2010, Sahin et al. 2012). To date, the effect of early growth on postprandial responses of any appetite regulatory hormones has been investigated in only one previous study (Schou et al. 2005). In that study, GLP-1 responses were measured and no effect of birth weight on GLP-1 levels was observed. As mentioned above, GLP-1 affects appetite regulation by delaying food transit time from the stomach to the duodenum (Edholm et al. 2010), as well as through the central nervous system. Clearly, further studies are needed in this area.

2.3 Early growth and blood pressure

2.3.1 Blood pressure and salt intake

Elevated blood pressure (hypertension) is a common health problem in both the developing and developed countries. The WHO has estimated that the worldwide prevalence of elevated blood pressure among adults ≥ 25 years of age is 29.2% in men and 24.8% in women (World Health Organization 2011). Definitions for elevated blood pressure are presented in Table 4. Elevated blood pressure is also common in Finland; based on the FINRISK 2007 study, 52.1% of men and 33.6% of women have systolic blood pressure (SBP) over 120 mmHg and diastolic blood pressure (DBP) over 80 mmHg (Kastarinen et al. 2009). Elevated blood pressure is a major risk factor for CVD, including coronary heart disease and stroke (Mancia et al. 2007). It has been estimated that elevated blood pressure causes 7.5 million deaths globally, which is about 12.8% of all deaths (World Health Organization 2011).
Table 4. Definitions for elevated blood pressure according to the European Society of Hypertension and the European Society of Cardiology (Mancia et al. 2007) and the Finnish Current Care Guideline for Hypertension (Jula et al. 2009).

<table>
<thead>
<tr>
<th>SBP (mmHg)</th>
<th>DBP (mmHg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Optimal</td>
<td>&lt; 120</td>
</tr>
<tr>
<td>Normal</td>
<td>120–129</td>
</tr>
<tr>
<td>High-normal</td>
<td>130–139</td>
</tr>
<tr>
<td>Mild hypertension, grade 1</td>
<td>140–159</td>
</tr>
<tr>
<td>Moderate hypertension, grade 2</td>
<td>160–179</td>
</tr>
<tr>
<td>Severe hypertension, grade 3</td>
<td>≥ 180</td>
</tr>
</tbody>
</table>

SBP, systolic blood pressure; DBP, diastolic blood pressure.

Increased salt intake is one risk factor for elevated blood pressure (He and MacGregor 2002, Strazzullo et al. 2009). Other major lifestyle-related risk factors of elevated blood pressure include overweight, smoking, excessive alcohol intake and physical inactivity (Mancia et al. 2007). In addition, increased intake of saturated fatty acids (SFAs) and decreased intake of potassium, fruits and vegetables elevate blood pressure (Mancia et al. 2007).

Blood pressure reduction in response to decreasing salt intake varies among individuals. Both genetic factors and acquired factors affect blood pressure reactivity to salt, which is also known as salt sensitivity. For example, metabolic syndrome (Hoffmann and Cubeddu 2007), older age and African-American race (Luft et al. 1991, Richardson et al. 2013) have been linked with increased salt sensitivity. The aetiology of salt sensitivity is still unknown; however, it has been proposed that differences in blood pressure reactivity to salt may be the result of alterations in renal handling of salt and the renin-angiotensin-aldosterone system, as well as impaired microvascular functioning (Ando and Fujita 2012, Richardson et al. 2013, Weinberger 1996).

2.3.2 Early growth and blood pressure in later life

Several epidemiological studies have shown an inverse relationship between birth size and adult hypertension. A few systematic reviews and meta-analyses have also been published (Gamborg et al. 2007, Huxley et al. 2000, Law and Shiell 1996, Mu et al. 2012), summarizing data for over 80 studies and with over 400 000 participants. These systematic reviews and meta-analyses have demonstrated that each 1-kg higher birth weight is associated with 2–3-mmHg lower SBP. Similar results regarding birth weight and adult-life DBP have also been observed, although
the association is not as strong as it is with SBP (Mu et al. 2012). Studies have shown that the relationship between birth weight and blood pressure is stronger in older populations than in younger populations and thus the association is age-dependent (Gamborg et al. 2007).

Alterations of kidney structure and functioning may be the underlining mechanism by which prenatal growth is linked with blood pressure in later life. Growth retardation during prenatal life leads to permanently reduced nephron number (Hinchliffe et al. 1992, Hughson et al. 2008, Manalich et al. 2000, Zidar et al. 1998) and impaired kidney functioning (Hallan et al. 2008) in humans. This association also exists across normal birth weight ranges (Hughson et al. 2003). It has been suggested that reduction in the number of nephrons leads to increased glomerular filtration rates, thereby increasing the risk of glomerulosclerosis and further nephron deaths, decreased sodium excretion and subsequently elevated blood pressure (Brenner and Chertow 1994, Zandi-Nejad et al. 2006). Animal models also support the hypothesis that prenatal growth affects blood pressure by showing that maternal dietary protein restriction during pregnancy causes reduced nephron number and leads to hypertension in adult offspring (Langley and Jackson 1994, Manning and Vehaskari 2001, Woods et al. 2001, Woods et al. 2004).

2.3.3 Early growth and salt sensitivity

It has been proposed that individuals who were born with low birth weight could be particularly sensitive to the blood pressure-raising effect of salt, because they have alterations in kidney functioning. This hypothesis was tested in two studies of rodents in which pregnant rats consumed a low-protein diet. The offspring of these rats suffered from salt-sensitive hypertension during adult life (Augustyniak et al. 2010, Woods et al. 2004). Two independent animal studies also demonstrated that prenatal growth retardation caused alterations in both protein and gene expression of sodium transporters in the kidney and thus affected salt sensitivity (Alwasel and Ashton 2009, Manning et al. 2002).

There are only two small intervention studies in which the effect of birth weight on salt sensitivity has been investigated in humans. A study of young children demonstrated that renal mass is reduced in children born with low birth weight and is dependent on the degree of growth retardation (Simonetti et al. 2008). Growth retardation also lowered the glomerular filtration rate, increased salt sensitivity and elevated blood pressure. In addition, they found that the highest prevalence of salt sensitivity was observed among those children who had experienced the most severe growth retardation during gestation. Another study of adults confirmed this finding by showing that birth weight affects the salt sensitivity of blood pressure (de Boer et al. 2008). In this study, healthy adults consumed a high-salt diet for one week and
thereafter a low-salt diet for one week. The blood pressure was measured before and after both test periods. The salt sensitivity of blood pressure was defined as the difference in mean arterial blood pressure between the high- and low-salt periods. They showed that birth weight was inversely related to salt sensitivity of blood pressure; a 1-kg lower birth weight increased the salt sensitivity of blood pressure by 2.0 mmHg. Based on these studies, birth weight seems to modify the blood pressure-raising effect of salt. However, it remains to be determined whether these findings are also seen in larger study populations.

2.4 Early growth and postprandial responses

2.4.1 Importance of postprandial responses

The associations between metabolic abnormalities and CVD risk factors have been studied mostly in the fasting state. However, with the exception of the first few hours in the morning, individuals spend most of their daytime hours in a postprandial state. The important contribution of the postprandial state to the differing risk of diseases is increasingly being recognized, and to date there is strong evidence that elevated postprandial levels of glucose and lipids are independent risk factors for CVD and type 2 diabetes. In fact, they may even be better predictors of cardiovascular morbidity and mortality than fasting levels (Bansal et al. 2007, Mah and Bruno 2012, Nordestegaard et al. 2007, Rendell and Jovanovic 2006).

There are several possible mechanisms by which elevated postprandial responses of glucose, insulin and lipids may cause adverse health effects and be directly atherogenic. For example, triglyceride (TG) rich chylomicrons during the postprandial period are converted to remnants that could penetrate the arterial wall and deposit cholesterol. The remnants could also affect the atherosclerotic process by converting macrophages into foam cells (Goldberg et al. 2011, Jackson et al. 2012, Kolovou et al. 2011). In addition, postprandial hyperglycaemia may promote atherogenesis by several mechanisms, e.g. by increasing formation of free radicals and causing nonreversible glycosylation of proteins (Mah and Bruno 2012, O'Keefe and Bell 2007). Furthermore, postprandial hyperinsulinaemia leads to sodium retention and sympathetic nervous system activation, both of which have adverse health effects (Kopp 2006). Indeed, elevated postprandial levels of both glucose and TG even within the physiological range in healthy individuals, lead to increased production of proinflammatory cytokines and oxidative stress causing postprandial inflammation, which may further contribute to endothelial dysfunctioning (Klop et al. 2012). These postprandial changes, when repeated multiple times each day, can predispose to the development of CVD and type 2 diabetes.
2.4.2 Early growth and findings in postprandial studies

There is an increased interest in the field of early growth and metabolic disease risk factors, such as glucose and lipids, later in life. However, surprisingly little information is available on whether prenatal or postnatal growth affects postprandial metabolism. In one of the first postprandial studies within the DOHaD field, Byrne et al. (1997) tested postprandial TG and free fatty acid (FFA) responses in 57 men of known birth weight. The subjects consumed a test meal that contained 65 g of fat per m² of body surface area (14% was derived from carbohydrates, 3% from protein and 83% from fat). Blood samples were drawn in the fasting state and four times during an 8-h test period. The effect of birth weight on the postprandial responses of TG and FFA was examined by stratification of the group by birth weight (< 3.1 kg, 3.1–4.1 kg, > 4.1 kg). Participants born with low birth weight had higher postprandial TG and FFA responses than subjects who were born with medium birth size. In addition, participants who belonged to the highest birth weight group also had higher postprandial lipid responses than did the medium birth weight group. However, the birth weight groups did not differ significantly. The study included both nonsmokers and smokers. Several studies have reported significantly greater postprandial TG levels in smokers than in nonsmokers, without differences in fasting levels (Kabagambe et al. 2009, Lopez-Miranda et al. 2007). Therefore, excluding smokers is desirable when postprandial lipid responses are examined.

In another postprandial study, the effect of birth weight on macronutrient oxidative and nonoxidative metabolism was investigated in elderly healthy men (Kensara et al. 2006). Participants were born either below the 25th percentile of birth weight (under 3.23 kg, n = 16) or above the 75th percentile (over 3.89 kg, n = 13). In this study, participants consumed a meal containing 3720 kJ of energy of which 40% was derived from carbohydrates, 15% from protein and 45% from fat. Kensara et al. observed that the 6-h postprandial responses of glucose, TGs and FFAs did not differ significantly between groups. However, it is nevertheless challenging to investigate the effect of low birth weight if the controls are not born at medium birth weight, because high birth weight can also cause adverse effects on glucose and lipid metabolism (Dabelea et al 1999, Eriksson et al 2003, Harder et al 2007).

In the third study of this field, Schou and coworkers (2005) measured glucose and insulin responses in young normal weight men who were born either with low birth weight (< 3010 g, n = 24) or were matched normal birth weight controls (3390 g, n = 25). The participants consumed a standardized breakfast containing 2370 kJ of energy; the carbohydrate content was 47%, protein 19% and fat 34%. In this study, participants with low birth weight had higher fasting glucose but not insulin levels. Indeed, postprandial 195-min responses of both glucose and insulin were significantly increased in the group born with low birth weight. In this study,
Incretins, glucose-dependent insulinotropic polypeptide (GIP) and GLP-1 responses were also measured. However, although Schou et al. (2005) observed higher insulin levels after a meal, they did not demonstrate that low birth weight affects the secretion of incretins or \( \beta \)-cell stimulation by incretins.

Taken together, these results of postprandial studies demonstrate that early growth may affect insulin secretory responses. However, only a handful of postprandial studies have been published. In addition, the study protocols used have varied between studies, as have the exclusion criteria and characteristics of the controls. Therefore, far-going conclusions cannot be made based on the studies published.

### 2.4.3 Early growth and findings in fasting studies

In contrast to postprandial studies, several other studies have investigated the associations between body size at birth and fasting glucose, insulin and lipid concentrations in later life. In addition to fasting values, these results are also based on oral glucose tolerance tests (OGTTs). To date, there is strong epidemiological evidence that low birth weight is related to elevated fasting and 2-h glucose levels in adulthood, which indicate impaired glucose regulation (Newsome et al. 2003). Impaired glucose tolerance is usually the result of impaired insulin secretion and insulin resistance. There have been assumptions that early growth may affect \( \beta \)-cell functioning and thus alter insulin secretion. Animal models support this by showing decreased \( \beta \)-cell mass among offspring whose mothers were fed a low-protein diet during pregnancy (Dumortier et al. 2007). However, when insulin secretion has been measured in humans, using intravenous glucose tolerance tests or OGTTs, variable results have been observed. In a meta-analysis, Newsome and colleagues (2003) showed that size at birth was inversely associated with the measure of insulin secretion in 15 of 27 studies, not related in 5 and positively related only in 7 studies. These findings suggest that individuals who were born with small body size have no clear primary defect in insulin secretion. In contrast to insulin secretion, several studies have shown that low birth weight is linked with reduced insulin sensitivity, as reported in the same meta-analysis (Newsome et al. 2003). Insulin resistance in low birth weight individuals could be the cause of multiple abnormalities in their insulin-sensitive tissues, including muscles and adipose tissues (Boiko et al. 2005, Eriksson et al. 2002, Jensen et al. 2007, Jensen et al. 2008, Maiorana et al. 2007, Taylor et al. 1995).

The association between low birth weight and unfavourable lipid profiles, such as elevated fasting total cholesterol and TG levels, has also been widely studied. Several systematic reviews and meta-analyses have been published. Three of these reviews or meta-analyses have shown that a 1-kg increase in birth weight was
associated with about a $0.03-0.05$-mmol/l decrease in total cholesterol in adult individuals (Huxley et al. 2004, Lawlor et al. 2006, Owen et al. 2003). These analyses comprised 28–58 studies and 20 000–70 000 individuals. However, one systematic review did not detect relationships between birth weight and total cholesterol. The authors did, however, find that there was a negative or U-shaped association between birth weight and fasting TG levels (Lauren et al. 2003). In addition to birth weight, some studies have also demonstrated that slow postnatal growth is associated with elevated fasting lipid levels (Kajantie et al. 2008, Skidmore et al. 2007, Tzoulaki et al. 2010). The associations between birth weight or postnatal growth and later-life fasting lipid concentrations are, however, surprisingly weak compared with the associations between birth weight and CVD risk.

The associations between low birth weight or slow postnatal growth and unfavourable lipid profiles could be due to altered liver functioning and metabolism (Burns et al. 1997, Lane et al. 2001, Winick and Noble 1966). In addition, alterations in lipid metabolism could be the result of excess lipid content in the liver (Kotronen and Yki-Järvinen 2008). Previous studies support this showing that low birth weight and small body size during infancy is associated with a higher likelihood of non-alcoholic fatty liver disease later in life (Fraser et al. 2008, Sandboge et al. 2013).
3 Aims of the study

The general aim of this study was to determine whether early growth is associated with dietary-related risk factors of metabolic diseases in later life. The specific aims were as follows:

1. To determine whether body size at birth is related to food consumption and nutrient intake in adulthood (I).

2. To investigate whether birth weight modifies the association between salt intake and blood pressure (II).

3. To examine the influences of early growth on postprandial glucose, insulin and lipid responses in later life (III, IV).

4. To explore whether early growth affects postprandial appetite regulatory hormone and incretin responses (IV).
4 Materials and methods

4.1 Helsinki Birth Cohort Study

The subjects in this thesis were all participants in the Helsinki Birth Cohort Study (HBCS), originally comprising 4630 men and 4130 women. They were born as singletons at the Helsinki University Central Hospital between 1934 and 1944, attended child welfare clinics in the city and lived in Finland in 1971, when a unique identification number was allocated to each member of the Finnish population (Figure 4).

Data on growth of the HBCS participants were obtained from hospital birth records and child welfare clinic records. Their birth records included date of birth, weight and length at birth and date of mother’s last menstrual period, based on which gestational age was calculated. Child welfare records included serial measurements of weight and height in infancy and childhood. Individuals belonging to the original study cohort were traced, using personal identification numbers.

Those HBCS participants who were still alive and residing in Finland were sent a questionnaire in the year 2000 (n = 7078). The questionnaires gave information on their general background including weight and height, health and diseases, medication, lifestyles and social circumstances. A total of 4515 individuals answered the questionnaires.

To obtain a sample size of over 2000 individuals for the clinical examination, a random-number table was used to invite 2902 subjects living in the greater Helsinki area. Of these, 2003 men (n = 928) and women (n = 1075) participated in the clinical examination between August 2001 and March 2004.
Figure 4. Flowchart of subgroups of the HBCS participants included in the studies in this thesis.
4.2 Clinical epidemiological studies (I, II)

4.2.1 Clinical examinations

The subjects attended the clinical examination after an overnight fast. Height was measured to the nearest 0.1 cm and weight to 0.1 kg in light clothing without shoes. The BMI was calculated as the weight in kilograms divided by the square of the height in metres. Blood pressure was measured from the right arm while the participant was in a sitting position and was recorded as the mean of two successive readings from a standard sphygmomanometer (Omron Matsusaka Europe, Hoofdorp, the Netherlands). A team of three trained research nurses performed all measurements. The participants were also asked about their medical history and current medication, using questionnaires at the clinic. In addition, educational attainment, smoking and exercise habits were obtained from a postal questionnaire before the clinical examination.

The Ethics Committee of Epidemiology and Public Health of the Hospital District of Helsinki and Uusimaa approved the clinical examination. Written informed consent was obtained from each subject.

4.2.2 Dietary assessments

The diet was assessed by a validated, self-administered, 128-item food-frequency questionnaire (FFQ) (Männistö et al. 1996, Paalanen et al. 2006). The FFQ was designed to assess the usual diet over the previous 12 months. The participants were asked to indicate the average intake frequency of each food-item and mixed dish, presented as 12 main food groups, e.g. dairy products and vegetables. The response options for nine possible frequency categories ranged from never or seldom to six or more times per day. The portion sizes for each food-item were specified in natural units (e.g. one banana, 190 g), common household measures (e.g. one glass of milk, 170 g) or portions (e.g. one portion of meat soup, 300 g). The portion sizes were based on the national FINDIET Survey and present the most commonly used portion sizes in Finland.

At the clinic, the participants completed the FFQ, which was then checked by a research nurse. The food intake data were entered and processed at the National Institute for Health and Welfare, Helsinki, Finland with the in-house calculation software Finessi, utilizing the National Food Composition Database FINELI! (National Institute for Health and Welfare 2005). The daily food intake was calculated by multiplying the frequency of food consumption by fixed portion sizes.
to obtain the weight of each listed food-item consumed as an average per day. The average daily intake of nutrients was calculated by multiplying the gram intake per day of each food by its nutrient content.

Salt (NaCl) intake included the natural sodium content of raw foods (sodium intake x 2.548), the salt added in cooking in average recipes and the salt used in manufactured foods. The dietary glycaemic index (GI) and glycaemic load (GL) were calculated, using the GI-database (Kaartinen et al. 2010). Dietary GI was calculated as the weighted mean of the GI values of the carbohydrate-containing foods in the diet, in which weighting was based on the proportion of the total carbohydrate content provided by each food. The dietary GL was calculated by multiplying the dietary GI value with the carbohydrate content of the diet and dividing by 100.

4.2.3 Statistical methods

The nutrients and foods were adjusted for energy intake by calculating the proportion of energy (E%) or by using the residual method (fibre, dietary GL, food groups, salt) (Willett and Stampfer 1986). Educational attainment was categorized into three groups, according to the number of years in school: basic (≤ 9 years of education); secondary (10–12 years) and higher (≥ 13 years of education). The participants were defined as current smokers if they smoked one or more cigarettes per day. Those who exercised at a level comparable to brisk walking three or more times per week were defined as physically active.

In Study I, the subjects were excluded if their FFQ had ≥ 10 blank food-items (n = 2) or if their calculated energy intake was under 2.7 MJ/d or over 25.5 MJ/d, corresponding to 0.5% at both ends of the self-reported daily energy intake distributions for men and women (n = 20). In all, 180 subjects were excluded, because their gestational age at birth was under 37 completed weeks, over 44 completed weeks or was not recorded. In addition, the BMI was not recorded for two subjects and for one it was considered too high to be included in the analysis (68.39 kg/m²). In addition, one participant was excluded because his fruit and berry intake was over 5 kg/d. The final analysis of Study I comprised 1797 subjects.

In Study II, the subjects were excluded if their FFQ was incomplete (n = 22), their gestational age was missing (n = 122) or their BMI was missing (n = 2) or considered to be too high (n = 1) as in Study I. In addition, underreporters were also excluded. Underreporters were defined as subjects whose energy intake was below 1.27 * the basal metabolic rate, which is the minimum survival requirements used when habitual diet is measured (Goldberg et al. 1991). The basal metabolic rate was estimated, using WHO equations that take into account weight, age and sex (World Health Organization 1985). The final analysis of Study II comprised 1512 subjects.
Associations were also recalculated for this thesis by excluding those whose gestational age at birth was under 37 completed weeks or over 44 completed weeks (n = 44).

The relationship between body size at birth and food and macronutrient intake was examined by linear regression analysis (Study I). The models were adjusted for potential confounding variables, which included sex and current age (Model 1), while Model 2 was further adjusted for current BMI, smoking, education and gestational age. Additional analyses were also adjusted for physical activity (Model 3). Statistical analyses were done using the PASW Statistics version 18 for Windows® (SPSS Inc., Chicago, IL, USA). For this thesis summary, the associations between body size at birth and food and macronutrient intake were recalculated by adjusting Model 2 further for total energy intake (Model 4). Linear regression analyses of Models 2, 3 and 4 rendered similar results. Therefore, only the results of Models 1 and 2 will be presented below.

The SBP and DBP were adjusted for sex, current age and BMI, use of antihypertensive medication, gestational age and total energy intake (Study II). The relationship between the adjusted SBP and DBP and the energy-adjusted salt intake was examined visually and a piecewise multivariate regression with unknown breakpoints (Muggeo 2003) was fitted. The best breakpoints with respect to both birth weight and salt intake were determined simultaneously by fitting the linear regression and choosing the model with the best (i.e. lowest) value of the Akaike information criterion. In addition, a likelihood-ratio test was used to assess the need for the breakpoint. The breakpoints themselves were considered to be a parameter, and the final P-values were adjusted as described by Hunsberger et al. (2002). The linear piecewise regression models for the best fitting breakpoints were then finally estimated. Statistical analyses were done, using R-software (version 2.8.1, The R Foundation for Statistical Computing) and PASW Statistics version 18 for Windows® (SPSS Inc., Chicago, IL, USA).

4.3 Postprandial studies (III, IV)

4.3.1 Subjects

A total of 24 overweight or obese (BMI 25–32 kg/m²) 65–75 year-old subjects, 12 with slow growth during infancy (SGI) and 12 age-, BMI- and sex-matched controls, participated in the postprandial studies between November 2009 and May 2010. The subjects belonged to the HBCS and had attended a clinical examination in 2001–2004. Growth during infancy was examined as gains in BMI between birth and 1 year of age. Each measurement of BMI for each individual was converted to a Z-
score, which represented the difference from the mean value for the entire study cohort (n = 2003) and was expressed as standard deviations (SDs). We examined how much the BMI at 1 year of age differed from that predicted by the BMI at birth, using the residual from linear regression; this measure was called ‘conditional growth’. Participants who were born preterm (before 37 weeks of gestation) or whose gestational age was over 44 completed weeks or was not reported were excluded. Subjects in the SGI-group had a conditional growth of < -0.9 SD, and the conditional growth of the control group was > -0.9 SD. The conditional growth breakpoints were based on the previous study of the HBCS, in which it was observed that participants whose conditional growth was < -1 SD had significantly higher fasting level of cholesterol and TG in adulthood (Kajantie et al. 2008).

Glucose tolerance was assessed by a 75-g 2-h OGTT, and diabetics (fasting glucose ≥ 7.0 mmol/l and/or 2-h glucose ≥ 11.1 mmol/l) were excluded. Other exclusion criteria included smoking, milk allergy, regular medication that could have affected postprandial glucose or lipid metabolism (e.g. antidiabetic drugs, fibrates and asthma medicine), gastrointestinal disease influencing absorption, a first-degree family history of diabetes or blood donation less than 3 months before the study. Participants’ diet, health and lifestyle data were assessed by questionnaires.

The postprandial study was conducted according to the guidelines laid down in the Declaration of Helsinki and all procedures involving human subjects were approved by the Ethics Committee of the Hospital District of Helsinki and Uusimaa. Written informed consent was obtained from all the subjects.

### 4.3.2 Study design and test meals

The study used a crossover design. The subjects were recruited to participate in six 1-day studies separated by approximately 1 week. One participant in the control group was able to attend for only the first two study visits (Study III). Therefore, another subject was recruited to attend the last four visits (Study IV). The subjects were requested to follow their usual diet during the study period. In addition, they were not allowed to drink alcohol and were asked to avoid strenuous exercise and sauna for 24 h before each study day. Moreover, the subjects were advised not to take lipid-lowering medication (statins) preceding the study visit. The day before the study day, they were asked to consume an evening meal, in accordance with the instructions they had been given, which would provide 15% of the calculated daily energy requirement. The mean energy intake of the subjects was calculated, based on their calculated basal metabolic rate, taking into account the questionnaire data on daily physical activity (World Health Organization 2004). The carbohydrate content of the evening meals was 55 E%. The subjects were also asked to fast for...
10–12 h after their standardized evening meal, to avoid exercise in the morning of the study and to arrive at the clinic by car or public transportation.

In the study clinic, body height and weight were measured and the BMI was calculated. Changes of up to 2 kg in weight were allowed during the study. The waist circumference was measured in a standing position, with the legs slightly apart, midway between the lowest ribs and the iliac crest. An intravenous cannula was inserted into an antecubital vein in the forearm and an intravenous blood sample was drawn. Thereafter, the subjects consumed the test meal within 10 min. After the start of the meal, venous blood samples were collected at 15, 30, 60, 90, 120, 180 and 240 min.

In Study III, all subjects consumed two different test meals, a fast-food meal (FF-meal) and a meal that followed the macronutrient composition of the dietary guidelines (REC-meal) in randomized order at 1-week intervals. Both test meals contained the same amount of energy. The foodstuffs and the nutrient composition of the test meals are shown in Table 5. In Study IV, all participants tested four test meals in random order: two high-protein meals and two high-fat meals (Table 5). Both high-protein meals contained the same amount of energy, carbohydrate, protein, and fat. Only the type of milk protein differed between meals: calcium caseinate (casein-meal) or whey protein isolate (whey-meal). Two high-fat meals also contained the same amount of energy, carbohydrates, protein and fat; only the type of fat (saturated [SFA-meal] or polyunsaturated [PUFA-meal] fat) differed between test meals.
### Materials and methods

#### Table 5. Foodstuffs and macronutrient content of the test meals.

<table>
<thead>
<tr>
<th>Components, g</th>
<th>FF-meal</th>
<th>REC-meal</th>
<th>Casein-meal</th>
<th>Whey-meal</th>
<th>SFA-meal</th>
<th>PUFA-meal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Barley porridge</td>
<td>-</td>
<td>215</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Raspberry jam</td>
<td>-</td>
<td>32</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Orange juice</td>
<td>-</td>
<td>260</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Rye bread</td>
<td>-</td>
<td>48</td>
<td>-</td>
<td>-</td>
<td>45</td>
<td>45</td>
</tr>
<tr>
<td>Margarine 70%</td>
<td>-</td>
<td>10</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>11</td>
</tr>
<tr>
<td>Cheese 5%</td>
<td>-</td>
<td>37</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Milk (0% fat)</td>
<td>-</td>
<td>150</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Cocoa powder</td>
<td>-</td>
<td>15</td>
<td>27</td>
<td>27</td>
<td>26</td>
<td>26</td>
</tr>
<tr>
<td>Cucumber</td>
<td>-</td>
<td>40</td>
<td>40</td>
<td>40</td>
<td>40</td>
<td>40</td>
</tr>
<tr>
<td>Rapeseed oil</td>
<td>-</td>
<td>15</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>BigMac-hamburger</td>
<td>210</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>French fries</td>
<td>99</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Light Cola</td>
<td>380</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Whey</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>44</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Casein</td>
<td>-</td>
<td>-</td>
<td>44</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Water</td>
<td>-</td>
<td>-</td>
<td>400</td>
<td>400</td>
<td>350</td>
<td>350</td>
</tr>
<tr>
<td>Milk powder</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>23</td>
<td>23</td>
</tr>
<tr>
<td>Vanilla sugar</td>
<td>-</td>
<td>-</td>
<td>5</td>
<td>5</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Sugar</td>
<td>-</td>
<td>-</td>
<td>15</td>
<td>15</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>White bread</td>
<td>-</td>
<td>-</td>
<td>37</td>
<td>37</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Butter</td>
<td>-</td>
<td>-</td>
<td>25</td>
<td>25</td>
<td>60</td>
<td>-</td>
</tr>
<tr>
<td>Solid margarine</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>51</td>
<td>-</td>
</tr>
<tr>
<td>Energy, kJ</td>
<td>3326</td>
<td>3359</td>
<td>2638</td>
<td>2638</td>
<td>2961</td>
<td>2961</td>
</tr>
<tr>
<td>Carbohydrate, g (E%)</td>
<td>76 (38)</td>
<td>102 (52)</td>
<td>59 (38)</td>
<td>59 (38)</td>
<td>51 (29)</td>
<td>51 (29)</td>
</tr>
<tr>
<td>Protein, g (E%)</td>
<td>31 (16)</td>
<td>29 (14)</td>
<td>45 (29)</td>
<td>45 (29)</td>
<td>12 (7)</td>
<td>12 (7)</td>
</tr>
<tr>
<td>Fat, g (E%)</td>
<td>40 (46)</td>
<td>30 (34)</td>
<td>23 (33)</td>
<td>23 (33)</td>
<td>50 (64)</td>
<td>50 (64)</td>
</tr>
</tbody>
</table>

E%, percentage of total energy; FF-meal, fast-food meal; PUFA-meal, polyunsaturated fatty acid meal; REC-meal, meal macronutrient composition followed dietary guidelines; SFA-meal, saturated fatty acid meal.

1. Barley porridge prepared with low-fat (1.5% fat) milk.
2. Raspberry jam, Menu Vadelmahillo; Roberts Ltd, Finland.
3. Orange juice, Vip Appelsiini täysmehu; Vip-Juicemaker Ltd, Finland.
4. Wholegrain rye bread, REAL-ruisleipä; Fazer Ltd, Finland.
5. Keiju margarine 70%; Raisio Ltd, Finland.
6. Aamu cheese 5%; Arla Ltd, Finland.
7. Dumle cocoa powder; Fazer Ltd, Finland.
8. McDonalds Ltd, Finland.
9. Whey protein isolate (protein content 89 g/100 g); Glanbia Nutritionals, Ireland.
10. Calcium caseinate I (protein content 90 g/100 g); DMW International, the Netherlands.
11. Milk powder (fat-free, lactose-free); Valio Ltd, Finland.
12. White bread, Vaasan Iso Paahto - vehnä; Vaasan&Vaasan Ltd, Finland.
13. Butter, lactose-free; Arla Ltd, Finland.
14. Solid margarine, Keiju Juokseva Rypsiöljyvalmiste; Raisio Ltd, Finland.
4.3.3 Laboratory analysis

Blood glucose was analysed by a glucose meter (HemoCue Glucose 201+ meter; HemoCue LtD, Espoo, Finland) which expresses concentrations as mmol/l of plasma glucose. The HemoCue Glucose system is based on a glucose dehydrogenase method. The mean interassay coefficient of variation (CV) was 0.8%. Blood for determination of plasma TGs, FFAs and insulin were collected in EDTA K2 tubes (Venosafe TM; Terumo Sweden AB, Västra Frölunda, Sweden). Blood for determination of plasma PYY, ghrelin, GLP-1 and GIP (Study IV) were collected in prechilled BD™ P800 tubes (Becton, Dickenson and Company, Franklin Lakes, NJ, USA), containing DPP-4 and other protease inhibitor cocktails. The samples were centrifuged for 15 min at 4000 rpm (Rotofix 32; Hettich Zentrifugen, Tuttlingen, Germany) immediately after sample collection and the separated plasma was stored at −70 °C until analysed.

The plasma concentrations of TGs were measured by an enzymatic glycerol-3-phosphate oxidase method (Abbott Laboratories, Abbott Park, IL, USA), FFAs were measured by an enzymatic colorimetric method (NEFA-HR(2); Wako Chemicals GmbH, Neuss, Germany) and insulin was determined by a chemiluminescent microparticle immunoassay with Abbott reagents. Fasting total cholesterol at baseline was analysed, using an enzymatic method (Abbott Laboratories, Abbott Park, IL, USA). The TG, FFA, insulin and cholesterol laboratory analyses were done, using an Architect ci8200 analyser (Abbott Laboratories, Abbott Park, IL, USA). The interassay CV of TGs at the levels of 1.35 mmol/l and 4.93 mmol/l were 1.7% and 1.3%, and the intra-assay CV was 0.6%. The interassay CV of FFAs at the levels of 0.59 mmol/l and 1.11 mmol/l were 1.8% and 1.7%, and the intra-assay CV varied from 0.3% (low-level control) to 0.8% (high-level control). The CVs of insulin at the levels of 187 pmol/l and 1191 pmol/l were 1.6% and 2.5%, and the intra-assay CV was 1.7%.

PYY (total), ghrelin (active) and GIP (total) were measured with a MILLIPLEX MAP Human Metabolic Hormone Panel kit (HMH-34K), using a Luminex 200 System (Luminex Corporation, Austin, TX, USA). The plasma concentrations of GLP-1 were measured against standards of synthetic GLP-1 7-36-amide, using antiserum code no. 89390 (Orskov et al. 1994). The assay measures the sum of the intact, active hormones and the metabolites generated by DPP-4. The results therefore reflect the secretion of the GLP-1. The interassay CV of PYY was 4% and the intra-assay CV was 12%. The interassay and intra-assay CVs of ghrelin were 7% and 7%, respectively, and the interassay and intra-assay CVs of GIP were 7% and 5%, respectively. The interassay CV of GLP-1 was 6%.
4.3.4 Subjective satiety profile

The casein-meal and whey-meal subjective satiety profiles were measured, using a seven-point visual analogue scale (VAS) (Holt and Miller 1995). The participants rated their satiety immediately before each blood sample was drawn. The categories were scored from 0 ‘I am extremely hungry’ through 3 ‘No particular feeling’ to 6 ‘I am extremely full’. The participants were requested to mark a vertical line on the horizontal axis corresponding to their sensations that were the most appropriate at the time.

4.3.5 Statistical methods

The GLP-1 and subjective satiety profiles were measured only in the casein-meal and whey-meal. The 4-h incremental area under the glucose, insulin, TG, PYY, GIP and GLP-1 response curve (IAUC) and the 4-h incremental area over the FFA response curve (IAOC) were calculated, using a trapezoidal method for each test meal (World Health Organization 1998). In addition, the 2-h glucose, insulin, PYY, GIP and GLP-1 IAUC and the 4-h total area under the curve (totAUC) of the satiety profile and ghrelin were also calculated.

One subject’s insulin IAUC results were excluded from the FF-meal analysis because his fasting insulin value was high and differed markedly from his previous study day fasting value as well as the fasting values of all other subjects.

The postprandial responses that were not normally distributed were log-transformed prior to the analyses. An independent sample t-test (Study III) and repeated-measures analysis of variance (ANOVA) (Study IV) were used for testing the differences in postprandial responses between the study groups. If the repeated-measures ANOVA was significant, an independent sample t-test was used to test whether the study groups differed in the different test meals (Study IV). An independent sample t-test was also used for testing the differences in baseline characteristics of the subjects between the study groups. All statistical analyses were carried out with the PASW Statistics version 18 for Windows® (SPSS Inc., Chicago, IL, USA); the level of significance was $P < 0.05$.

The sample size calculation for the postprandial studies was based on our previous study of elderly men and women who belonged to the HBCS (Perälä et al. 2011) and has been calculated for a sensitivity of 0.80 and a two-sided significance level of 0.05, indicating the 12 subjects were sufficient for both study groups. In addition, similar sample size has been used previous postprandial studies in the DOHaD field (Byrne et al. 1997, Kensara et al. 2006).
5 Results

5.1 Body size at birth and food intake (I)

There was no interaction between the effects of sex and PI at birth or birth weight or birth length on food and macronutrient intake, and therefore men and women were analysed together. Table 6 shows the participants’ characteristics at birth and adulthood.

Body size at birth was related to food intake in later life, such that each unit increase in PI at birth was associated with a 13.26-g higher intake of fruits and berries and a 1-kg increase in birth weight with an 83.16-g higher intake of fruits and berries (Table 7). Higher PI at birth also predicted higher intake of rye and rye products in adulthood. No other statistically significant associations were observed between PI at birth, birth weight or birth length (Tables 7 and 9) and adult-life food intake. Adding physical activity or total energy intake to Model 2 did not attenuate the results (data not shown).

A 1-kg/m³ increase in PI at birth was associated with a 0.14-E% lower intake of total fat and a 0.06-E% lower intake of monounsaturated fatty acids (MUFAs) (Table 8). One unit increase in PI at birth was also associated with a 0.18-E% higher intake of carbohydrates as well as a 0.08-E% higher intake of sucrose, 0.05-E% higher fructose and 0.18-g higher fibre intake in adulthood. Similar associations were observed between birth weight and macronutrient intake (Table 8). Birth length was not significantly associated with macronutrient intake or dietary GI or GL in adulthood (Table 9). Adding physical activity to Model 2 did not attenuate the results (data not shown).
### Table 6. Birth and adult data of the participants 1.

<table>
<thead>
<tr>
<th></th>
<th>Men</th>
<th>Women</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>836</td>
<td>961</td>
</tr>
<tr>
<td>Birth data</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight (g)</td>
<td>3514 (464)</td>
<td>3381 (438)</td>
</tr>
<tr>
<td>Gestational age (wk)</td>
<td>40.05 (1.42)</td>
<td>40.14 (1.44)</td>
</tr>
<tr>
<td>Length (cm)</td>
<td>50.8 (1.9)</td>
<td>50.1 (1.7)</td>
</tr>
<tr>
<td>Ponderal index (kg/m^3)</td>
<td>26.7 (2.3)</td>
<td>26.8 (2.2)</td>
</tr>
<tr>
<td>Adult data</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (y)</td>
<td>61.5 (2.8)</td>
<td>61.5 (3.1)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>85.9 (13.6)</td>
<td>74.0 (13.8)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>176.8 (6.0)</td>
<td>163.3 (5.7)</td>
</tr>
<tr>
<td>BMI (kg/m^2)</td>
<td>27.5 (4.0)</td>
<td>27.8 (5.0)</td>
</tr>
<tr>
<td>Educational attainment 2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Basic (%)</td>
<td>39.5</td>
<td>43.7</td>
</tr>
<tr>
<td>Secondary (%)</td>
<td>24.0</td>
<td>22.9</td>
</tr>
<tr>
<td>Higher (%)</td>
<td>36.4</td>
<td>33.4</td>
</tr>
<tr>
<td>Smoker (%) 3</td>
<td>21.9</td>
<td>15.9</td>
</tr>
<tr>
<td>Physically active (%) 4</td>
<td></td>
<td>46.1</td>
</tr>
<tr>
<td>Dietary intake</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Energy intake (MJ)</td>
<td>10.3 (3.6)</td>
<td>8.6 (3.0)</td>
</tr>
<tr>
<td>Carbohydrate (E%)</td>
<td>45.5 (6.7)</td>
<td>47.7 (6.6)</td>
</tr>
<tr>
<td>Sucrose (E%)</td>
<td>9.0 (3.4)</td>
<td>9.8 (3.6)</td>
</tr>
<tr>
<td>Fat (E%)</td>
<td>33.7 (5.7)</td>
<td>33.1 (5.3)</td>
</tr>
<tr>
<td>SFA (E%)</td>
<td>12.2 (2.6)</td>
<td>12.1 (2.6)</td>
</tr>
<tr>
<td>MUFA (E%)</td>
<td>11.3 (2.2)</td>
<td>10.9 (2.1)</td>
</tr>
<tr>
<td>PUFA (E%)</td>
<td>5.3 (1.2)</td>
<td>5.3 (1.2)</td>
</tr>
<tr>
<td>Protein (E%)</td>
<td>16.6 (2.5)</td>
<td>17.4 (2.5)</td>
</tr>
<tr>
<td>Alcohol (E%)</td>
<td>4.4 (5.2)</td>
<td>1.9 (2.7)</td>
</tr>
</tbody>
</table>

BMI, body mass index; E%, percentage of total energy intake; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; SFA, saturated fatty acids.

1 Results are expressed as mean (SD) or proportions.
2 Educational attainment; three categories by approximate years studied (0–9 = basic, 10–12 = secondary, 13 or more = higher).
3 Smoking one or more cigarettes per day.
4 Proportion of people who exercised three or more times per week.
Table 7. Association between ponderal index at birth and birth weight and energy-adjusted food intake in adulthood. Mean difference (95% CI) in daily food intake is given to the increase of 1 kg/m³ in ponderal index at birth or 1 kg in birth weight (n = 1797).

<table>
<thead>
<tr>
<th>Food group (g)</th>
<th>Ponderal index (kg/m³)</th>
<th>Birth weight (kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Model 1</td>
<td>Model 2</td>
</tr>
<tr>
<td></td>
<td>Regression coefficients</td>
<td>(95% CI)</td>
</tr>
<tr>
<td>Cereals</td>
<td>1.44 (-1.10, 3.97)</td>
<td>0.27</td>
</tr>
<tr>
<td>Rye products</td>
<td>1.41 (0.06, 2.76)</td>
<td>0.041</td>
</tr>
<tr>
<td>Wheat products</td>
<td>-0.09 (-1.80, 1.62)</td>
<td>0.92</td>
</tr>
<tr>
<td>Fruits and berries</td>
<td>10.90 (-1.93, 23.74)</td>
<td>0.096</td>
</tr>
<tr>
<td>Vegetables and roots</td>
<td>2.75 (-4.86, 10.36)</td>
<td>0.48</td>
</tr>
<tr>
<td>Potato products</td>
<td>-3.02 (-6.73, 0.69)</td>
<td>0.11</td>
</tr>
<tr>
<td>Fish products</td>
<td>-1.27 (-3.19, 0.65)</td>
<td>0.19</td>
</tr>
<tr>
<td>Total meat</td>
<td>0.09 (-3.81, 3.99)</td>
<td>0.96</td>
</tr>
<tr>
<td>Red meat</td>
<td>0.69 (-2.82, 4.19)</td>
<td>0.70</td>
</tr>
<tr>
<td>Processed meat</td>
<td>-0.07 (-2.16, 2.02)</td>
<td>0.95</td>
</tr>
<tr>
<td>Milk products</td>
<td>5.57 (-7.80, 18.95)</td>
<td>0.41</td>
</tr>
<tr>
<td>Fats</td>
<td>0.24 (-0.41, 0.90)</td>
<td>0.47</td>
</tr>
<tr>
<td>Butter</td>
<td>0.08 (-0.46, 0.61)</td>
<td>0.78</td>
</tr>
<tr>
<td>Margarine</td>
<td>0.21 (-0.15, 0.56)</td>
<td>0.25</td>
</tr>
<tr>
<td>Sugar and confectionery</td>
<td>0.12 (-0.97, 1.21)</td>
<td>0.83</td>
</tr>
</tbody>
</table>

95% CI; 95% confidence interval.
Model 1: Adjusted for sex and current age, tested by linear regression model.
Model 2: Adjusted for sex, current age and body mass index, education, smoking and gestational age, tested by linear regression model.
Table 8. Association between ponderal index at birth and birth weight and nutrient intake in adulthood. Mean difference (95% CI) in daily nutrient intake is given to the increase of 1 kg/m^3 in ponderal index at birth or 1 kg in birth weight (n = 1797).

<table>
<thead>
<tr>
<th>Nutrient intake</th>
<th>Model 1</th>
<th>Model 2</th>
<th>Model 1</th>
<th>Model 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ponderal index (kg/m^3)</td>
<td>Regression coefficients (95% CI)</td>
<td>P</td>
<td>Regression coefficients (95% CI)</td>
<td>P</td>
</tr>
<tr>
<td>Energy (kJ)</td>
<td>27.9 (-41.1, 97.0)</td>
<td>0.43</td>
<td>22.0 (-48.2, 92.2)</td>
<td>0.54</td>
</tr>
<tr>
<td>Carbohydrates (E%)</td>
<td>0.14 (-0.01, 0.28)</td>
<td>0.055</td>
<td>0.18 (0.04, 0.32)</td>
<td>0.010</td>
</tr>
<tr>
<td>Fructose (E%)</td>
<td>0.04 (0.00, 0.09)</td>
<td>0.051</td>
<td>0.05 (0.01, 0.09)</td>
<td>0.019</td>
</tr>
<tr>
<td>Sucrose (E%)</td>
<td>0.05 (-0.02, 0.13)</td>
<td>0.18</td>
<td>0.08 (0.00, 0.15)</td>
<td>0.050</td>
</tr>
<tr>
<td>Fibre (g)</td>
<td>0.15 (-0.02, 0.32)</td>
<td>0.074</td>
<td>0.18 (0.02, 0.34)</td>
<td>0.031</td>
</tr>
<tr>
<td>Dietary GI</td>
<td>-0.08 (-0.18, 0.01)</td>
<td>0.093</td>
<td>-0.08 (-0.18, 0.02)</td>
<td>0.11</td>
</tr>
<tr>
<td>Dietary GL</td>
<td>0.21 (-0.25, 0.67)</td>
<td>0.37</td>
<td>0.35 (-0.11, 0.81)</td>
<td>0.13</td>
</tr>
<tr>
<td>Protein (E%)</td>
<td>-0.01 (-0.06, 0.04)</td>
<td>0.67</td>
<td>-0.03 (-0.08, 0.03)</td>
<td>0.30</td>
</tr>
<tr>
<td>Fat (E%)</td>
<td>-0.11 (-0.22, 0.01)</td>
<td>0.061</td>
<td>-0.14 (-0.26, -0.03)</td>
<td>0.013</td>
</tr>
<tr>
<td>SFA (E%)</td>
<td>-0.03 (-0.08, 0.03)</td>
<td>0.35</td>
<td>-0.04 (-0.09, 0.02)</td>
<td>0.20</td>
</tr>
<tr>
<td>MUFA (E%)</td>
<td>-0.04 (-0.09, 0.01)</td>
<td>0.076</td>
<td>-0.06 (-0.10, -0.01)</td>
<td>0.015</td>
</tr>
<tr>
<td>PUFA (E%)</td>
<td>-0.02 (-0.04, 0.01)</td>
<td>0.22</td>
<td>-0.02 (-0.05, 0.01)</td>
<td>0.11</td>
</tr>
<tr>
<td>Alcohol (E%)</td>
<td>-0.01 (-0.10, 0.07)</td>
<td>0.76</td>
<td>-0.01 (-0.09, 0.08)</td>
<td>0.85</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Birth weight (kg)</th>
<th>Model 1</th>
<th>Model 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Regression coefficients (95% CI)</td>
<td>P</td>
<td></td>
</tr>
<tr>
<td>Energy (kJ)</td>
<td>247.7 (-90.7, 586.2)</td>
<td>0.15</td>
</tr>
<tr>
<td>Carbohydrates (E%)</td>
<td>0.36 (-0.32, 1.04)</td>
<td>0.30</td>
</tr>
<tr>
<td>Fructose (E%)</td>
<td>0.24 (0.03, 0.45)</td>
<td>0.029</td>
</tr>
<tr>
<td>Sucrose (E%)</td>
<td>0.29 (-0.09, 0.66)</td>
<td>0.13</td>
</tr>
<tr>
<td>Fibre (g)</td>
<td>0.65 (-0.16, 1.46)</td>
<td>0.12</td>
</tr>
<tr>
<td>Dietary GI</td>
<td>-0.51 (-0.99, -0.03)</td>
<td>0.036</td>
</tr>
<tr>
<td>Dietary GL</td>
<td>-0.22 (-2.45, 2.02)</td>
<td>0.85</td>
</tr>
<tr>
<td>Protein (E%)</td>
<td>-0.01 (-0.27, 0.25)</td>
<td>0.96</td>
</tr>
<tr>
<td>Fat (E%)</td>
<td>-0.39 (-0.95, 0.17)</td>
<td>0.18</td>
</tr>
<tr>
<td>SFA (E%)</td>
<td>-0.15 (-0.42, 0.12)</td>
<td>0.27</td>
</tr>
<tr>
<td>MUFA (E%)</td>
<td>-0.11 (-0.33, 0.12)</td>
<td>0.35</td>
</tr>
<tr>
<td>PUFA (E%)</td>
<td>-0.03 (-0.16, 0.10)</td>
<td>0.63</td>
</tr>
<tr>
<td>Alcohol (E%)</td>
<td>0.02 (-0.39, 0.44)</td>
<td>0.91</td>
</tr>
</tbody>
</table>

95% CI; 95% confidence interval; E%, percentage of total energy intake; GI, glycaemic index; GL, glycaemic load; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; SFA, saturated fatty acids.

Model 1: Adjusted for sex and current age, tested by linear regression model.
Model 2: Adjusted for sex, current age and body mass index, education, smoking and gestational age, tested by linear regression model.

1 Adjusted for energy by residual method.
Table 9. Association between birth length and food and nutrient intake in adulthood. Mean difference (95% CI) in daily food and nutrient intake is given to the increase of 1 cm in birth length (n = 1797).

<table>
<thead>
<tr>
<th>Foods and nutrients</th>
<th>Model 1 Regression coefficients (95% CI)</th>
<th>P</th>
<th>Model 2 Regression coefficients (95% CI)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Birth length (cm)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fruits and berries (g)</td>
<td>-0.47 (-4.81, 3.88)</td>
<td>0.83</td>
<td>-2.13 (-6.60, 2.35)</td>
<td>0.35</td>
</tr>
<tr>
<td>Processed meat (g)</td>
<td>-0.51 (-3.11, 2.08)</td>
<td>0.70</td>
<td>-1.46 (-3.95, 1.04)</td>
<td>0.25</td>
</tr>
<tr>
<td>Milk products (g)</td>
<td>-0.80 (-17.4, 15.8)</td>
<td>0.93</td>
<td>1.19 (-16.6, 19.0)</td>
<td>0.90</td>
</tr>
<tr>
<td>Fats (g)</td>
<td>0.05 (-0.77, 0.86)</td>
<td>0.92</td>
<td>-0.11 (-0.97, 0.76)</td>
<td>0.81</td>
</tr>
<tr>
<td>Butter (g)</td>
<td>0.09 (-0.57, 0.75)</td>
<td>0.78</td>
<td>-0.01 (-0.66, 0.65)</td>
<td>0.99</td>
</tr>
<tr>
<td>Margarine (g)</td>
<td>-0.26 (-0.76, 0.24)</td>
<td>0.31</td>
<td>-0.16 (-0.64, 0.31)</td>
<td>0.50</td>
</tr>
<tr>
<td></td>
<td><strong>Nutrients</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Energy (kJ)</td>
<td>40.3 (-45.4, 126.0)</td>
<td>0.36</td>
<td>37.6 (-54.0, 129.1)</td>
<td>0.42</td>
</tr>
<tr>
<td>Carbohydrates (E%)</td>
<td>-0.02 (-0.20, 0.15)</td>
<td>0.81</td>
<td>-0.02 (-0.20, 0.16)</td>
<td>0.81</td>
</tr>
<tr>
<td>Fructose (E%)</td>
<td>0.03 (-0.02, 0.08)</td>
<td>0.28</td>
<td>0.03 (-0.03, 0.08)</td>
<td>0.35</td>
</tr>
<tr>
<td>Sucrose (E%)</td>
<td>0.04 (-0.06, 0.13)</td>
<td>0.45</td>
<td>0.05 (-0.05, 0.15)</td>
<td>0.36</td>
</tr>
<tr>
<td>Fibre (g)</td>
<td>0.05 (-0.16, 0.25)</td>
<td>0.65</td>
<td>0.05 (-0.16, 0.27)</td>
<td>0.64</td>
</tr>
<tr>
<td>Dietary GI</td>
<td>-0.10 (-0.23, 0.02)</td>
<td>0.09</td>
<td>-0.09 (-0.21, 0.04)</td>
<td>0.19</td>
</tr>
<tr>
<td>Dietary GL</td>
<td>-0.33 (-0.90, 0.24)</td>
<td>0.25</td>
<td>-0.30 (-0.90, 0.29)</td>
<td>0.32</td>
</tr>
<tr>
<td>Protein (E%)</td>
<td>0.01 (-0.06, 0.07)</td>
<td>0.81</td>
<td>-0.01 (-0.08, 0.06)</td>
<td>0.76</td>
</tr>
<tr>
<td>Fat (E%)</td>
<td>-0.01 (-0.15, 0.14)</td>
<td>0.94</td>
<td>-0.03 (-0.18, 0.12)</td>
<td>0.69</td>
</tr>
<tr>
<td>SFA (E%)</td>
<td>-0.01 (-0.08, 0.05)</td>
<td>0.68</td>
<td>-0.01 (-0.08, 0.07)</td>
<td>0.91</td>
</tr>
<tr>
<td>MUFA (E%)</td>
<td>0.01 (-0.05, 0.07)</td>
<td>0.75</td>
<td>-0.01 (-0.07, 0.05)</td>
<td>0.65</td>
</tr>
<tr>
<td>PUFA (E%)</td>
<td>0.01 (-0.03, 0.04)</td>
<td>0.68</td>
<td>-0.01 (-0.04, 0.03)</td>
<td>0.81</td>
</tr>
<tr>
<td>Alcohol (E%)</td>
<td>0.01 (-0.09, 0.12)</td>
<td>0.80</td>
<td>0.06 (-0.06, 0.17)</td>
<td>0.32</td>
</tr>
</tbody>
</table>

95% CI; 95% confidence interval; E%, percentage of total energy intake; GI, glycaemic index; GL, glycaemic load; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; SFA, saturated fatty acids.

Model 1: Adjusted for sex and current age, tested by linear regression model.
Model 2: Adjusted for sex, current age and body mass index, education, smoking and gestational age, tested by linear regression model.

1 Adjusted for energy by residual method.
5.2 Early growth, salt intake and blood pressure (II)

Birth weight was inversely associated with unadjusted SBP throughout the study population ($\rho = -0.058; \ P = 0.02$) but not with unadjusted DBP ($\rho = 0.010; \ P = 0.71$). A positive association between unadjusted salt intake and unadjusted DBP was observed ($\rho = 0.070; \ P = 0.006$), but not between unadjusted salt intake and unadjusted SBP ($\rho = 0.004; \ P = 0.89$). The best-obtained breakpoint for birth weight was 3050 g and for salt intake 10.0 g (based on an energy intake of 10.5 MJ). The model with the breakpoints was significantly better than the one without breakpoints (adjusted $\ P = 0.003$).

Table 10. Birth measurements and adult clinical data 1.

<table>
<thead>
<tr>
<th></th>
<th>Men</th>
<th>Women</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$\leq 3050$ g</td>
<td>$&gt; 3050$ g</td>
</tr>
<tr>
<td>N</td>
<td>105</td>
<td>575</td>
</tr>
<tr>
<td>Birth data</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight (g)</td>
<td>2780 (247)</td>
<td>3619 (374)</td>
</tr>
<tr>
<td>Gestational age (wk)</td>
<td>38.6 (1.4)</td>
<td>40.2 (1.4)</td>
</tr>
<tr>
<td>Length (cm)</td>
<td>48.2 (1.7)</td>
<td>51.2 (1.6)</td>
</tr>
<tr>
<td>Adult data</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (y)</td>
<td>62.0 (2.4)</td>
<td>61.5 (2.9)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>81.8 (10.1)</td>
<td>84.9 (13.0)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>174.5 (5.1)</td>
<td>177.2 (5.9)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.9 (3.4)</td>
<td>27.0 (3.7)</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>91.1 (9.9)</td>
<td>90.1 (10.6)</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>148.9 (19.7)</td>
<td>144.8 (19.2)</td>
</tr>
<tr>
<td>Antihypertensive medication (%)</td>
<td>38.7</td>
<td>33.7</td>
</tr>
<tr>
<td>Salt intake (g/d)</td>
<td>10.7 (3.8)</td>
<td>10.7 (3.4)</td>
</tr>
</tbody>
</table>

BMI, body mass index; DBP, diastolic blood pressure; SBP, systolic blood pressure.

1 Differences are expressed as mean (SD) or proportions.

2 Differences between birth weight groups, tested by Student’s t-test, except in the case of antihypertensive medication, for which Pearson’s chi-square test for binomial proportions was applied.
Men and women who were born with low birth weight ($\leq 3050$ g) were lighter and smaller in adulthood; however, adulthood BMI was similar in low birth weight and higher birth weight groups (Table 10). Antihypertensive medication was more common in individuals with low birth weight than those with higher birth weight. Salt intake did not differ significantly between the birth weight groups.

The modifying effect of birth weight on the association between salt intake and SBP did not differ between men and women; therefore, men and women were analysed together. Among participants whose birth weight exceeded 3050 g, salt intake was not significantly related to SBP or DBP. However, among low birth weight subjects ($\leq 3050$ g), a 1-g higher daily salt intake was associated with a 2.48-mmHg (95% Confidence interval [CI]: 0.40, 4.52; $P = 0.025$) higher SBP until the saturation point of 10.0 g (Figure 5). After the saturation point (10.0 g), SBP was estimated to be 4.72 mmHg higher among the low birth weight group than among the higher birth weight group (95% CI: 1.96, 7.48; $P < 0.001$). After exclusion of participants taking antihypertensive medication from the analyses (data included 1000 participants), the modifying effect of birth weight on the association between salt intake and SBP became smaller and was not statistically significant (1.20 mmHg; $P = 0.42$). Among low birth weight subjects, salt intake was not significantly associated with DBP.

Excluding subjects whose gestational age at birth was under 37 completed weeks or over 44 completed weeks had only a minor effect on the results. In the low birth weight group, a 1-g increase in salt intake was associated with a 1.30-mmHg (95% CI: 0.01, 2.59; $P = 0.048$) higher SBP but not with DBP. Among the high birth weight group, salt intake was significantly associated with neither SBP nor DBP.
Figure 5. Association between energy-adjusted salt (NaCl) intake and systolic blood pressure among subjects with birth weight ≤ 3050 g (x = observed value, solid bold line = fitted model (linear regression with flexible breakpoint) and birth weight > 3050 g (o = observed value, dashed bold line = fitted model) (n = 1512). The thin dotted lines, obtained by locally weighted scatter plot smoothing (LOWESS method), are given for comparison. Reprinted from the American Journal of Clinical Nutrition 2011; 93:422–6, Perälä et al. “The association between salt intake and adult systolic blood pressure is modified by birth weight”, with permission from the American Society for Nutrition.
5.3 Early growth and postprandial responses (III, IV)

5.3.1 Characteristics

The basic characteristics of the study participants are shown in Table 11. Five subjects in the SGI and four subjects in the control group used lipid-lowering medications (statins). Seven participants in the SGI and five participants in the control group had impaired glucose tolerance (2-h glucose ≥ 7.8 mmol/l).

Table 11. Characteristics of the postprandial studies participants.

<table>
<thead>
<tr>
<th></th>
<th>SGI (III, IV)</th>
<th>Control group (III)</th>
<th>Control group (IV)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N (men, women)</td>
<td>12 (6,6)</td>
<td>12 (7,5)</td>
<td>12 (6,6)</td>
</tr>
<tr>
<td>Birth and childhood data</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>3129 (360)</td>
<td>3648 (399)</td>
<td>3597 (469)</td>
</tr>
<tr>
<td>Birth length (cm)</td>
<td>49.6 (2.4)</td>
<td>51.3 (0.9)</td>
<td>51.0 (1.5)</td>
</tr>
<tr>
<td>Birth BMI (kg/m²)</td>
<td>12.7 (1.0)</td>
<td>13.9 (1.3)</td>
<td>13.8 (1.3)</td>
</tr>
<tr>
<td>Gestational age (wk)</td>
<td>39.9 (1.3)</td>
<td>40.6 (1.0)</td>
<td>40.3 (1.9)</td>
</tr>
<tr>
<td>Z-score BMI at birth</td>
<td>-0.66 (0.85)</td>
<td>0.33 (1.0)</td>
<td>0.26 (1.1)</td>
</tr>
<tr>
<td>Weight at 1 y (kg)</td>
<td>8.9 (0.7)</td>
<td>10.9 (0.6)</td>
<td>10.8 (0.7)</td>
</tr>
<tr>
<td>Height at 1 y (cm)</td>
<td>74.7 (3.1)</td>
<td>77.1 (1.0)</td>
<td>76.9 (1.4)</td>
</tr>
<tr>
<td>BMI at 1 y (kg/m²)</td>
<td>15.9 (0.5)</td>
<td>18.3 (0.7)</td>
<td>18.3 (0.8)</td>
</tr>
<tr>
<td>Z-score BMI at 1 y</td>
<td>-1.34 (0.34)</td>
<td>0.46 (0.47)</td>
<td>0.44 (0.49)</td>
</tr>
<tr>
<td>Conditional growth</td>
<td>-1.23 (0.28)</td>
<td>0.40 (0.32)</td>
<td>0.40 (0.32)</td>
</tr>
<tr>
<td>Adult data</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (y)</td>
<td>68.0 (2.13)</td>
<td>68.1 (3.23)</td>
<td>68.2 (3.38)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>75.5 (9.5)</td>
<td>80.5 (9.0)</td>
<td>80.5 (9.8)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>167.1 (10.0)</td>
<td>173.8 (7.5)</td>
<td>172.9 (8.6)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>27.0 (2.78)</td>
<td>26.7 (2.94)</td>
<td>27.0 (3.08)</td>
</tr>
<tr>
<td>Waist circumference (cm)</td>
<td>90.1 (9.7)</td>
<td>92.2 (10.2)</td>
<td>91.6 (9.8)</td>
</tr>
<tr>
<td>OGGT fasting glucose (mmol/l)</td>
<td>5.39 (0.39)</td>
<td>5.68 (0.50)</td>
<td>5.63 (0.47)</td>
</tr>
<tr>
<td>OGGT glucose 2-h (mmol/l)</td>
<td>8.03 (1.43)</td>
<td>7.91 (1.20)</td>
<td>7.77 (1.03)</td>
</tr>
<tr>
<td>Fasting cholesterol (mmol/l)</td>
<td>5.06 (0.72)</td>
<td>4.70 (0.85)</td>
<td>4.72 (0.84)</td>
</tr>
</tbody>
</table>

BMI, body mass index; OGGT, oral glucose tolerance test; SGI, group with slow growth during infancy.

1 Results are expressed as mean (SD).
2 Differed significantly from the SGI, \( P < 0.05 \).
3 Differed significantly from the SGI, \( P < 0.001 \).
5.3.2 Glucose and insulin responses

The 2-h and 4-h glucose responses did not differ significantly between the study groups. After ingestion of all the test meals, the concentration of plasma glucose increased for 30 min, declining thereafter and returning towards the fasting concentrations after 60–90 min (Table 12, Figure 6).

Growth during infancy affected the insulin postprandial responses and the groups differed significantly. The insulin 2-h and 4-h responses were significantly higher after the FF-meal (P = 0.082 and P = 0.036, respectively), the casein-meal (P = 0.031 and P = 0.023, respectively), the whey-meal (P = 0.066 and P = 0.037, respectively) and after the PUFA-meal (P = 0.008 and P = 0.002, respectively) for the SGI than for the control group. In addition, the insulin 2-h and 4-h responses were also higher after the REC-meal (P = 0.37 and P = 0.23) and the SFA-meal (P = 0.073 and P = 0.61) for the SGI than for the controls, although the differences were not statistically significant (Table 12, Figure 6). Excluding subjects who had impaired glucose tolerance from the analysis did not have major effect on the glucose or insulin results (data not shown).
Table 12. Glucose and insulin responses (IAUC) after test meals\(^1\).

<table>
<thead>
<tr>
<th></th>
<th>FF-meal</th>
<th>REC-meal</th>
<th>Casein-meal</th>
<th>Whey-meal</th>
<th>SFA-meal</th>
<th>PUFA-meal</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>SGI Glucose 2-h</strong></td>
<td>101 (18)</td>
<td>69 (16)</td>
<td>83 (22)</td>
<td>55 (16)</td>
<td>55 (13)</td>
<td>72 (17)</td>
</tr>
<tr>
<td><strong>Controls Glucose 2-h</strong></td>
<td>118 (26)</td>
<td>126 (31)</td>
<td>53 (13)</td>
<td>48 (11)</td>
<td>67 (13)</td>
<td>43 (9)</td>
</tr>
<tr>
<td><strong>SGI Glucose 4-h</strong></td>
<td>113 (17)</td>
<td>76 (12)</td>
<td>105 (24)</td>
<td>88 (23)</td>
<td>80 (20)</td>
<td>103 (16)</td>
</tr>
<tr>
<td><strong>Controls Glucose 4-h</strong></td>
<td>139 (31)</td>
<td>166 (41)</td>
<td>81 (19)</td>
<td>81 (14)</td>
<td>103 (24)</td>
<td>77 (12)</td>
</tr>
<tr>
<td><strong>SGI Insulin 2-h</strong></td>
<td>40 (319) (6290)</td>
<td>50 492 (7792)</td>
<td>49 192 (6817) (^2)</td>
<td>57 524 (9066)</td>
<td>27 083 (3644)</td>
<td>27 869 (5563) (^2)</td>
</tr>
<tr>
<td><strong>Controls Insulin 2-h</strong></td>
<td>24 925 (4897 (^3))</td>
<td>40 406 (7669)</td>
<td>30 366 (5721)</td>
<td>38 115 (8200)</td>
<td>17 314 (2359)</td>
<td>13 508 (1485)</td>
</tr>
<tr>
<td><strong>SGI Insulin 4-h</strong></td>
<td>53 458 (8763) (^2)</td>
<td>70 843 (10 699)</td>
<td>66 607 (10 168) (^2)</td>
<td>81 107 (13 418) (^2)</td>
<td>33 469 (4824)</td>
<td>35 227 (6885) (^2)</td>
</tr>
<tr>
<td><strong>Controls Insulin 4-h</strong></td>
<td>29 849 (5613 (^3))</td>
<td>54 206 (10 711)</td>
<td>38 636 (6946)</td>
<td>52 570 (11 881)</td>
<td>21 608 (3150)</td>
<td>19 203 (3203)</td>
</tr>
</tbody>
</table>

FF-meal, fast-food meal; IAUC, incremental area under the curve; PUFA-meal, polyunsaturated fatty acid meal; REC-meal, meal macronutrient composition followed dietary guidelines; SFA-meal, saturated fatty acid meal; SGI, group with slow growth during infancy.

\(^1\) Results are mean (standard error of the mean); Glucose responses are expressed as mmol·min/l and insulin responses are expressed as pmol·min/l; Glucose and insulin responses were log-transformed prior to the analyses.

\(^2\) Differed significantly from the controls, \(P < 0.05\), tested by an independent sample t-test.

\(^3\) \(n = 11\).
Results

Figure 6. Mean responses of plasma A) glucose and B) insulin to a fast-food-meal (Δ ▲) and a meal in which the macronutrient composition was according to dietary guidelines (○ ●), C) glucose and D) insulin to a casein-meal ( ▽ ▼) and whey-meal (◊ ●), and E) glucose and F) insulin to a saturated fatty acid meal (◊ ●) and a polyunsaturated fatty acid meal (□ ■) for subjects with slow growth during infancy (white marks) and for the control group (black marks).
5.3.3 Triglycerides and free fatty acids

After each test meal, the TG concentrations increased and were highest 3–4 hours after the start of the meal (Figure 7). Early growth affected the TG responses. The 4-h TG responses were significantly higher for the SGI than for the controls after the FF-meal (P = 0.047) and the PUFA-meal (P = 0.006) (Table 13). The TG responses of the REC-meal (P = 0.058), casein-meal (P = 0.158), whey-meal (P = 0.053) and SFA-meal (P = 0.074) were also higher for the SGI compared with the controls, although the differences were not statistically significant. Excluding subjects who used statins in the analysis did not have major effect on the results (data not shown).

Early growth did not significantly affect the postprandial FFA responses (Table 13). All the study meals suppressed the FFAs initially and reached nadir within 60–90 min.

Table 13. Postprandial triglyceride (TG) and free fatty acid (FFA) responses of the test meals.

<table>
<thead>
<tr>
<th></th>
<th>FF-meal</th>
<th>REC-meal</th>
<th>Casein-meal</th>
<th>Whey-meal</th>
<th>SFA-meal</th>
<th>PUFA-meal</th>
</tr>
</thead>
<tbody>
<tr>
<td>SGI TG 4-h IAUC</td>
<td>101 (15)</td>
<td>107 (12)</td>
<td>59 (9)</td>
<td>70 (11)</td>
<td>132 (18)</td>
<td>154 (21)</td>
</tr>
<tr>
<td>Controls TG 4-h IAUC</td>
<td>66 (8)</td>
<td>76 (10)</td>
<td>44 (8)</td>
<td>45 (10)</td>
<td>88 (12)</td>
<td>91 (10)</td>
</tr>
<tr>
<td>SGI FFA 4-h IAOC</td>
<td>67 (12)</td>
<td>64 (10)</td>
<td>71 (13)</td>
<td>59 (10)</td>
<td>31 (9)</td>
<td>36 (11)</td>
</tr>
<tr>
<td>Controls FFA 4-h IAOC</td>
<td>78 (14)</td>
<td>75 (9)</td>
<td>55 (9)</td>
<td>89 (12)</td>
<td>38 (7)</td>
<td>41 (9)</td>
</tr>
</tbody>
</table>

FF-meal, fast-food meal; IAOC, incremental area over the curve; IAUC, incremental area under the curve; PUFA-meal, polyunsaturated fatty acid meal; REC-meal, meal macronutrient composition followed dietary guidelines; SFA-meal, saturated fatty acid meal; SGI, group with slow growth during infancy.

1 Results are mean (standard error of the mean), expressed as mmol·min/l.
2 Differed significantly from the controls, P < 0.05, tested by an independent sample t-test.
3 Postprandial responses were log-transformed prior to the analyses.
Figure 7. Mean responses of plasma A) triglycerides (TG) and B) free fatty acids (FFA) to a fast-food-meal (Δ, ▲) and a meal in which the macronutrient composition was according to dietary guidelines (○, ●), C) TG and D) FFA to a casein-meal (▽, ▼) and whey-meal (♦, ◆), and E) TG and F) FFA to a saturated fatty acid meal (◦, ⋄) and a polyunsaturated fatty acid meal (○, □) for subjects with slow growth during infancy (white marks) and for the control group (black marks).
5.3.4 Appetite-related outcomes (PYY, ghrelin, satiety profiles)

The postprandial concentration of PYY increased and ghrelin decreased after consumption of the test meals (Figure 8, Table 14). The PYY responses were slightly higher for the SGI than for the controls; however, only the 2-h responses differed significantly between the study groups after the casein-meal ($P = 0.025$) and the whey-meal ($P = 0.046$). Early growth had no effect on the postprandial ghrelin responses. Measures of subjective satiety (VAS) increased after ingestion of the high-protein test meals, reaching peak values by 30–60 min and declining thereafter. The 4-h VAS response curves were slightly, but not significantly, higher for the SGI than for the control group (Table 14, Figure 9).

![Figure 8](image-url)

**Figure 8.** Mean responses of plasma A) peptide YY (PYY) and B) ghrelin to a casein-meal (△, ▼) and whey-meal (⊙, ●) and C) PYY and D) ghrelin to a saturated fatty acid meal (△, ▼) and a polyunsaturated fatty acid meal (⊙, ●) for subjects with slow growth during infancy (white marks) and for the control group (black marks).
5.3.5 GLP-1 and GIP responses

Neither the fasting concentrations nor the postprandial responses of GLP-1 or GIP differed significantly between the study groups. The postprandial concentration of GLP-1 peaked at 120 min and GIP at 60–90 min after the study meals (Figure 9, Table 14).

Figure 9. Mean responses of plasma glucose-dependent insulinotropic peptide (GIP) to A) a casein-meal (▽, ▼) and whey-meal (○, ▪) and B) a saturated fatty acid meal (●, ●) and a polyunsaturated fatty acid meal (□, ■), and C) plasma glucagon-like peptide 1 (GLP-1) and D) satiety profile to a casein-meal (▽, ▼) and whey-meal (○, ▪) for subjects with slow growth during infancy (white marks) and for the control group (black marks).
Table 14. Appetite regulatory hormone and incretin 2-h and 4-h postprandial responses after the test meals.  

|                      | Casein-meal | Whey-meal | SFA-meal | PUFA-meal | SGI Controls | SGI Controls | SGI Controls | SGI Controls | P  
|----------------------|-------------|-----------|----------|-----------|--------------|--------------|--------------|--------------|---  
| **PYY 2-h IAUC (pg·min/l)** | 6876 (1549) | 6880 (994) | 4772 (830) | 6218 (1428) | 7097 (1222) | 8849 (1682) | 0.005  
| **PYY 4-h IAUC (pg·min/l)** | 11 300 (2895) | 11 010 (1646) | 8042 (1536) | 15 537 (5589) | 15 955 (3373) | 20 944 (4086) | 0.15  
| **Ghrelin 4-h totAUC (pg·min/l)** | 6410 (1469) | 5213 (1033) | 4390 (746) | 6673 (1104) | 6156 (1056) | 7347 (576) | 0.95  
| **Satiety profile 4-h totAUC** | 803 (56) | 777 (57) | 882 (68) | 781 (62) | - | - | - | - | 0.35  
| **GIP 2-h IAUC (pg·min/l)** | 26 990 (3218) | 35 222 (4653) | 36 445 (4519) | 31 376 (7063) | 34 776 (6324) | 35 848 (5421) | 0.78  
| **GIP 4-h IAUC (pg·min/l)** | 63 734 (6139) | 71 880 (9387) | 77 132 (7896) | 72 576 (12 001) | 81 344 (13 974) | 56 530 (11 113) | 0.70  
| **GLP-1 2-h IAUC (pmol·min/l)** | 2283 (925) | 1810 (353) | 2058 (400) | - | - | - | - | 0.86  
| **GLP-1 4-h IAUC (pmol·min/l)** | 4312 (1369) | 3587 (619) | 3765 (676) | - | - | - | - | 0.29  

GIP, glucose-dependent insulinotropic peptide; GLP-1, glucagon-like peptide 1; IAUC, incremental area under the curve; PUFA-meal, polyunsaturated fatty acid meal; PYY, peptide YY; SFA-meal, saturated fatty acid meal; SGI, group with slow growth during infancy; totAUC, total area under the curve.

1 Results are expressed as mean (standard error of the mean).
2 Differences between the groups tested by repeated-measures ANOVA.
3 Postprandial responses were log-transformed prior to the analyses.
4 Differed significantly from the controls, $P < 0.05$. Tested an independent sample t-test if repeated-measures ANOVA were significant ($P < 0.05$).
6 Discussion

6.1 Birth size as a modifying factor

6.1.1 Food and macronutrient intakes

Diet plays an important and independent role in the development of several metabolic diseases. Although prenatal growth is strongly linked with these diseases, few studies have investigated whether prenatal growth is related to dietary habits.

We observed that body size at birth was positively related to consumption of fruits and berries, and rye and rye products in Finnish men and women aged 56 to 70. In addition, body size at birth was positively related to intake of carbohydrates, sugars, and fibre and inversely associated with intake of fat. Our findings indicate that prenatal growth may modify food intake in adult life, which may subsequently affect health outcomes in later life. The relationship between prenatal growth and food and nutrient intake later in life may be due to alterations of several physiologic and sensory factors (Ayres et al. 2012, Bayol et al. 2007, Beauchamp and Moran 1982, Breton et al. 2009, Erhuma et al. 2007, Mennella et al. 2001, Nakamura et al. 2008, Shigemura et al. 2004, Stein et al. 2006, Umabiki et al. 2010) that affect food choices.

Our findings are in agreement with previous studies in which low birth weight or famine during prenatal life was associated with higher intake of fat in children (Shultis et al. 2005, Stafford and Lucas 1998) and adults (Lussana et al. 2008, Stein et al. 2009). In contrast findings was observed Barbieri et al. (2009) who demonstrated that in Brazilian women severe intrauterine growth retardation was related to higher intake of carbohydrates. Differences in diet between study populations may, however, at least partly explain the inconsistent findings between the studies. Moreover, one study showed that Finnish adults who were born with very low birth weight consumed less fruits and vegetables than the controls (Kaseva et al. 2013). In that study, the difference between preterm and controls in the consumption of fruits and vegetables was about 60 g/d.

In our study population, carbohydrate intake was on average 2 E% lower than has been reported in Finnish adults in the FINDIET 2007 study (Paturi et al. 2008, Pietinen et al. 2010). In addition, women’s fat intake and men’s alcohol intake were higher in our study cohort than in the FINDIET 2007 study. However, the macronutrient intakes in our study population do meet the current Nordic Nutrition recommendations (Nordic Council of Ministers 2013), except for the SFA intake, which exceeded the recommended level.
In our study, a 1-kg increase in birth weight was associated with an about 80-g increase in daily intake of fruits and berries. Low consumption of fruits and berries reflects an unhealthy diet, which may increase the risk of CVD (Dauchet et al. 2009). Indeed, the importance of low fruit and berry intake as well as vegetable intake is highlighted by the WHO (Lopez et al. 2006), and is estimated to be the sixth major risk factor for death worldwide, thus causing a higher proportion of attributable deaths than overweight or physical inactivity. We suggest that low consumption of fruits and berries may be one explanation behind the association between small body size at birth and increased risk for metabolic diseases.

In addition to fruit and berries, low consumption of rye products has also been linked to an increased risk of CVD (Leinonen et al. 2000, Pietinen et al. 1996, Söderholm et al. 2012). In Finland, rye products are the major source of fibre. Based on the national FINDIET 2007 study, about half of the fibre intake in men and over a third in women were derived from rye bread (Paturi et al. 2008). Therefore, we propose that the positive association between body size at birth and fibre intake was mostly due to higher consumption of rye products.

### 6.1.2 Salt intake and blood pressure

The association between low birth weight and elevated blood pressure in later life has been well established (Gamborg et al. 2007). The association between birth weight and SBP, but not with DBP, was observed in our study as well. A previous report from the HBCS also demonstrated that those participants who had a diagnosis of hypertension in adulthood had a lower birth weight, were shorter in length and had a lower BMI at birth than did those with no diagnosis of hypertension (Eriksson et al. 2007).

We observed a relationship between salt intake and SBP in adult subjects with a birth weight of 3050 g or less, but not in subjects with higher birth weight. This finding supports two previously published intervention studies, in which low birth weight individuals had higher salt sensitivity to blood pressure in adulthood (de Boer et al. 2008) as well as in childhood (Simonetti et al. 2008). Increased salt sensitivity to blood pressure may well be explained by several alterations in kidney functioning that have been demonstrated in low birth weight individuals (Hallan et al. 2008, Hinchliffe et al. 1992, Hughson et al. 2003, Hughson et al. 2008, Manalich et al. 2000, Zidar et al. 1998). These results suggest that due to increased salt sensitivity, individuals with low birth weight may especially benefit from a reduction in dietary salt intake.

One previous study of young infants showed that birth weight was inversely related to salty taste preference, which could lead to increased salt intake (Stein et al. 2006). We did not observe differences in salt intake between birth weight groups.
Therefore, we do not believe that, at least in our study, elevated blood pressure of low birth weight individuals was due to their preference for salty foods. We found that in those who were born with low birth weight, a 1-g higher daily salt intake resulted in a 2.48-mmHg increase in SBP. In any population, even small differences in blood pressure have a high impact. For example, it was shown that even a 2-mmHg reduction in SBP among adults could decrease stroke mortality by about 10% and ischaemic heart disease or other vascular causes of mortality by about 7% (Lewington et al. 2002). In addition, it has been estimated that decreasing daily salt intake by 1 g and replacing 1 E% of SFA by 1 E% PUFA could lead to 8000–13 000 preventable CVD cases in Finnish adults during the next 20 years (Martikainen et al. 2011). Therefore, even a small persistent reduction in salt intake and thus in blood pressure could have major implications for public health. In our study population, salt intake was 2–4 g higher than the daily recommended intake (National Nutrition Council 2005, Nordic Council of Ministers 2013). Therefore, if participants who were born with low birth weight were to decrease their salt intake to the recommended level, their SBP could, in theory, decrease by at least 5 mmHg, leading to a major impact in cardiovascular health.

6.2 Early growth and postprandial responses

6.2.1 Glucose, insulin and incretin responses

Strong epidemiological evidence is available that low birth weight or small body size in infancy is associated with elevated fasting glucose and lipid levels. Less has been reported on whether early growth affects postprandial responses as well. We showed that birth size and growth during infancy have a major impact on adult postprandial insulin responses. In particular, both high-protein meals produced 1.5-fold greater insulin responses for the SGI-group than for the controls, despite similar adult anthropometric characteristics. Although insulin responses were significantly greater within the SGI-group, neither fasting levels nor postprandial responses of glucose differed between groups. It is, however, known that glucose homeostasis remains relatively normal for long periods of time, despite the development of hyperinsulinaemia or insulin resistance. Our finding is in line with a previous study in which higher postprandial insulin responses were observed among low birth weight participants compared with the age- and sex-matched controls after a standardized breakfast (Schou et al. 2005). Elevated insulin responses indicate insulin resistance. Previous studies have also reported increased insulin responses and impaired insulin sensitivity among low birth weight participants after intravenous or oral glucose tolerance testing.
(Newsome et al. 2003). Impaired insulin sensitivity could be the cause of multiple abnormalities in their insulin-sensitive tissues, including muscles and adipose tissues (Boiko et al. 2005, Eriksson et al. 2002, Jensen et al. 2007, Jensen et al. 2008, Maiorana et al. 2007, Taylor et al. 1995). We did not, however, find evidence to support the recent hypothesis that prenatal or postnatal growth reduces insulin secretion. However, the postprandial design used is not the optimal way for testing insulin secretions.

We detected no effect of early growth on fasting or postprandial incretin levels, although insulin responses were significantly greater in SGI. This finding is in line with a previous study by Schou et al. (2005), who found that subjects with low birth weight had higher insulin responses than did matched normal birth weight participants, whereas the GLP-1 and GIP responses did not differ between the groups. It has been observed that GLP-1 concentrations after a meal are reduced in type 2 diabetics, whereas individuals with impaired glucose tolerance have only slightly reduced incretin secretion (Nauck et al. 2011). In addition, recently published meta-analysis showed that type 2 diabetics do not display reduced GLP-1 secretion; only type 2 diabetics who have poorer glycaemic control may have reduced GLP-1 secretion (Calanna et al. 2013). Therefore, it is unlikely that growth retardation during early life alone affects incretin levels. Thus, even though individuals who grow slowly during prenatal life have an increased risk of developing type 2 diabetes (Whincup et al. 2008), incretins do not play a key role in disease risk.

Insulin resistance (Bansilal et al. 2007) as well as elevated fasting and nonfasting insulin levels (Kopp 2006, Sarwar et al. 2007) are associated with an increased risk of CVD. Studies have shown that elevated insulin levels can be directly atherogenic (Kopp 2006, Yu et al. 2011a). Insulin resistance may also cause elevated postprandial TG levels and thus cause atherosclerosis through dyslipidemia (Pansuria et al. 2012). We, therefore, propose that our findings of elevated postprandial insulin levels could be one mechanism underlying the increased risk of CVD in individuals who were small at birth and who grew slowly during infancy.

6.2.2 Triglycerides and free fatty acids responses

We observed that the postprandial TG responses were significantly greater, particularly, after high-fat meals, for the SGI-group than for the controls. No differences were seen between groups in either fasting TG concentrations or fasting FFA or FFA postprandial responses. Thus, elevated TG responses were not the result of increased adipose tissue lipolysis in the late postprandial state. Elevated TG responses indicate that individuals who grow slowly during early life have altered lipid metabolism. Restricted liver growth during early life may permanently alter
liver functioning and lipid metabolism, as detected and reported in animal studies (Burns et al. 1997, Lane et al. 2001, Winick and Noble 1966). Excess lipid content in the liver may also cause elevated postprandial lipid responses and insulin resistance. Studies have shown that non-alcoholic fatty liver disease is more common in adults who were born with small body size or were small at infancy (Fraser et al. 2008, Sandboge et al. 2013). Therefore, elevated TG responses could be the result of non-alcoholic fatty liver disease as well.

Our results differ from those of Byrne et al. (1997) and Kensara et al. (2006), who found no effect of birth weight on postprandial TG and FFA responses in elderly men and women. Unlike these two studies in which the effect of birth weight was investigated, we investigated the effect of early growth on postprandial responses and our participants were small at birth as well as at infancy. It was previously reported that prenatal growth has only a moderate effect on fasting TG levels (Huxley et al. 2004), whereas postnatal growth has a much greater effect (Kajantie et al. 2008). Therefore, we suggest that our observed elevated TG responses were mostly due to growth retardation during infancy.

It is well known that elevated postprandial lipid levels are closely related to a number of metabolic diseases, including CVD (Bansal et al. 2007, Nordestegaard et al. 2007). Small body size at birth as well as thinness during infancy are also associated with these diseases (Barker 1995, Barker et al. 2005, Eriksson et al. 2007, Huxley et al. 2007, Mu et al. 2012, Osmond et al. 2007, van Abeelen et al. 2011). The associations of birth weight with fasting lipid concentrations are quite modest, compared with the associations between birth weight and disease risk in later life. Since postprandial lipaemia occurs over 4–6 h in healthy individuals, most individuals spend the majority of their waking hours in a postprandial state. In addition, elevated postprandial TG levels are directly atherogenic (Goldberg et al. 2011, Jackson et al. 2012, Kolovou et al. 2011) and cause adverse health effects, such as increase oxidative stress and inflammation (Klop et al. 2012). Therefore, our findings of elevated postprandial TG levels are of significance for the understanding of increased metabolic disease risk among individuals who were born with small body size and who grew slowly during the first year of life.

6.2.3 Appetite regulatory hormones

Epidemiologic evidence is available that subjects who were born with small body size and grew slowly during infancy are less likely to become obese, whereas rapid growth during infancy has been associated with obesity later in life (Baird et al. 2005, Monasta et al. 2010, Monteiro and Victora 2005, Ong and Loos 2006, Schellong et al. 2012, Yu et al. 2011b, Zhao et al. 2012). It has been proposed that the relationship between early growth and obesity is due to alterations in the appetite

We showed that the postprandial responses of the appetite regulatory hormone PYY were significantly higher for the SGI-group than for the age-, sex- and BMI-matched controls. This finding may indicate greater satiety responses after a meal among subjects who grow slowly during early life. Greater satiety responses could potentially be one explanation why subjects who born with small body size and grow slowly during infancy are less likely to become obese.

Our results are in accordance with previous studies in young children that showed that infants who were born with low birth weight (Chen et al. 2012, Siahanidou et al. 2005) or born preterm (Berseth et al. 1992, Chen et al. 2012, Siahanidou et al. 2005, Siahanidou et al. 2007) had elevated levels of fasting PYY, reflecting greater satiety. Although early growth influenced the PYY responses in our study, we did not detect differences in ghrelin levels between the groups. Previous results from studies, in which the association between birth weight or weight gain during infancy and fasting levels of ghrelin have been investigated, have also been conflicting (Chen et al. 2012, Darendeliler et al. 2009, Iniguez et al. 2002, Kyriakakou et al. 2009, Larnkjaer et al. 2010, Park 2010, Sahin et al. 2012, Siahanidou et al. 2005).

To date, only one study has investigated the effect of birth weight on postprandial appetite regulatory hormone responses in adulthood (Schou et al. 2005). In that study, birth weight did not influence appetite regulation. However, in contrast to our study, they measured only GLP-1 levels, not other hormones that are involved in appetite regulation. We also saw that early growth did not influence the GLP-1 levels, even though it affected the PYY levels.

In addition to these gut-released appetite regulatory hormones, insulin is also an important satiety signal (Flint et al. 2007). Elevated postprandial insulin levels have found to reflect higher satiety feelings (Flint et al. 2007). Therefore, our observed elevated postprandial insulin responses in the SGI-group may thus also reflect greater satiety feelings in the postprandial state. However, it remains to be seen whether these alterations in PYY levels, as well as insulin levels, also reduce meal sizes in normal life and thus decrease the risk for developing obesity.
6.3 Strengths and limitations

6.3.1 Birth cohort and clinical epidemiological studies

The main strength of the clinical epidemiological studies of this thesis was the use of a large, well-characterized study cohort consisting of both men and women. In addition, birth data were obtained from reliable medical records and were not based on merely recalled values.

Our study cohort was restricted to people who were born in Helsinki University Central Hospital and who had attended child welfare clinics in Helsinki. The majority of children attended these clinics at that time. In addition, the participation rate of the clinical examination was 69% of those invited. However, the participants may not have represented the entire population living in Helsinki at that time. The social class distribution of the children in our study cohort, as defined by fathers’ occupations, was similar to that in Helsinki as a whole. In addition, participants in the clinical examinations did not differ significantly from nonparticipants throughout the cohort regarding birth measures, socioeconomic status and educational attainment. Selection bias would be expected to affect the results only if the relationships were different in participants compared with nonparticipants. This is unlikely, but cannot be excluded.

In this study cohort, most individuals were born or grew up during World War II. Families might have suffered from food shortages in Finland at that time. In addition, intergenerational effects may also have impacted the results. Factors causing low birth weight may have differed from factors we have today; e.g. maternal smoking was rare in Finland in that time. Therefore, this must be considered in generalizing these results to current settings. On the other hand, small size at birth predicts premature death (Risnes et al. 2011) and several chronic diseases (Barker 1995, Barker et al. 2005, Eriksson et al. 2007, Huxley et al. 2007, Mu et al. 2012, Osmond et al. 2007, van Abeelen et al. 2011, Whincup et al. 2008, Xiao et al. 2010), which could cause individuals to drop out from the follow-ups of studies more frequently. Therefore, survivor bias might have caused some underestimation in terms of the results, because individuals who participated in the clinical examinations were about 60 years of age.

The cross-sectional design of the epidemiological part allows no conclusions regarding causality to be made. Nevertheless, epidemiological observations are invaluable for generating hypotheses on which associations can be confirmed in experimental studies. Finally, gestational age was based on the date of last menstrual period; thus it is prone to error.
6.3.2 Dietary assessments

**FFQ**
In the clinical epidemiological part of this thesis, dietary intake was measured by a validated semiquantitative FFQ. FFQ measures individuals’ usual frequency of each food-item from a list of foods for a specific period (e.g. the past 12 months). Therefore, retrospective dietary assessment methods, such as diet history or FFQ, are widely used to assess the subjects’ past long-term diet (Thompson and Byers 1994). Food records are, however, often regarded as the gold standards against other dietary assessment methods. Food records have been developed to assess the frequency of consumption and usual portion size of foods and mixed dishes. However, it is time-consuming and costly and, therefore, not suitable for large cohorts (Thompson and Byers 1994). Instead of food records, FFQ is widely used to estimate the usual dietary intake in large cohort studies, because the costs of data collection and processing as well as the respondent burden are much lower for FFQ methods than for food records. In epidemiological studies, FFQ is used to rank subjects according to food consumption or nutrient intakes for the purpose of assessing the relationships between dietary intake and disease risk (Thompson and Byers 1994).

When the diet is assessed by an FFQ, participants may, however, overestimate the consumption of food considered healthy and underreport the intake of food considered unhealthy (Paalanen et al. 2006). This is especially true for women who may overestimate their fruit and vegetable intake and for obese subjects who may underreport intakes of energy-dense foods (Paalanen et al. 2006). However, the FFQ used measured the entire diet and ranks subjects reasonably well according to their dietary intakes (Männistö et al. 1996, Paalanen et al. 2006).

It is challenging to measure sodium or salt intake from food consumption data. However, the continually updated Finnish food composition database has included sodium content values since the 1980s (Pietinen 1982). In addition, our database was validated against 24-h urinary sodium excretion, and there is a correlation between dietary and urinary sodium measurements (Reinivuo et al. 2006). When salt intake is estimated by an FFQ, it is not possible to accurately measure the salt used in cooking and at the table. However, a validation study showed that sodium intake, as measured with our FFQ, was similar to that observed by diet records (Männistö et al. 1996). Furthermore, the average salt intake in our study was at the same level than in the national FINDIET 2007 study (Paturi et al. 2008, Pietinen et al. 2010). Therefore, we believe that the limitations of the FFQ in estimating salt intake did not affect the results.
Energy adjustments

Adjusting for nutrient or food intakes by applying the residual method (Willett and Stampfer 1986, Willett et al. 1997) is commonly used to control for confounding. Confounding can result if the total energy intake is related to disease risk, e.g. due to variation in body size or physical activity. The residual method may, however, introduce attenuation bias in the association, unless the total energy intake is also entered into the model (Palmgren and Kushi 1991). However, in Study I, the associations between body size at birth and food and nutrient intake in later life did not change, even though the total energy intake was included in the models.

6.3.3 Postprandial studies

Participants

The postprandial studies of this thesis included subjects who had either normal or impaired glucose tolerance. This may have increased the variation in the glucose responses and thus have affected the results. We, however, previously showed that impaired glucose tolerance affects only glucose responses, but not insulin, TG or FFA responses (Perälä et al. 2011).

The limitations of the postprandial studies may also have included the relatively small number of participants included in the studies. Therefore, the power of the postprandial studies may not have been sufficient to disclose all true differences between the study groups. Postprandial studies are extremely labour intensive and therefore the number of participants is limited. The sample size was based on power calculations and postprandial studies traditionally include 10–30 participants (Lairon et al. 2007). In addition, earlier postprandial studies in the DOHaD field have also contained similar amount of participants (Byrne et al. 1997, Kensara et al. 2006).

Another possible limitation related to the postprandial study populations was that participants who grew slowly during the first year of life were also born small. Therefore, we cannot distinguish between the effect of size at birth, growth during infancy or a combination of the two on postprandial responses. In addition, since all the subjects were overweight, our findings of early growth and postprandial responses may be limited only to overweight populations.

Meal size

It has been proposed that meal size in postprandial studies should be based on the participant’s body weight or body surface area (Lairon et al. 2007). This approach takes into account the greater blood volume in larger subjects. However, fat mass also influences postprandial fat clearance (Lairon et al. 2007). Differences in fat mass cause the main variation in body weight in adults. Thus, meal sizes that have been estimated by body size may introduce bias into the results. In addition, most of
the studies in the field of postprandial research have used a fixed meal size (Lairon et al. 2007). Therefore, we do believe that meal size used in our studies was suitable.

**Measurements**

In the postprandial studies, PYY was measured using an assay that did not differentiate between PYY (1-36) and PYY (3-36); therefore, the total hormone concentrations may not have reflected the active hormone levels. The differing forms of PYY may also play different roles in food intake regulations (Sloth et al. 2007). Therefore, we do not know whether the active PYY responses would have varied among the study groups.

Participants’ satiety profiles were assessed, using questionnaire that contained a scale of satiety feelings. These types of self-reported VAS questionnaires are widely used to measure several appetite-related subjective sensations (Blundell et al. 2010). In our study, each participant was instructed to use the satiety scale to increase the reliability of the method. The satiety feelings were also assessed immediately before blood drawing. Therefore, the experience of blood sampling did not affect the subjective evaluation of the satiety feelings. However, the most commonly used questionnaires typically contain multiple appetite sensation scales, such as hunger, satiety, fullness and desire to eat (Blundell et al. 2010). It is recommended that multiple appetite sensation scales should be used because the various sensations may sometimes be confused, e.g. satiety is often confused with fullness. Therefore, in future studies it would be desirable to use a VAS that measures different appetite feelings.

**6.4 Implications for further research**

To date, few studies have been performed within the DOHaD field that have focused on nutritional factors. Particularly, although numerous studies have investigated the role of prenatal or postnatal growth on glucose and insulin responses during an OGTT, only a handful of postprandial studies have been published. Postprandial assessment is, however, much more physiologic and describes better the normal life situation. Therefore, further studies that elucidate the effect of early growth on postprandial metabolism in large study groups are required. Upcoming postprandial studies should also put special emphasis on elucidating whether the size at birth, growth during infancy or the combination of the two is behind the elevated postprandial responses.

The findings of this thesis demonstrate that size at birth affects and modifies dietary related risk factors for metabolic disease, including unhealthy dietary habits and salt sensitivity. To see whether early growth modifies other associations
between diet and health conditions, for example physical functioning and body composition, further studies are needed. The results also suggest that PYY levels are elevated for participants who born with small body size and grow slowly during infancy. Although PYY and other physiologic factors regulate appetite, meals are mostly initiated by factors that are not based on energy needs, such as time of day. Therefore, more knowledge is needed whether elevated PYY responses could explain decreased risk of developing obesity among subjects with slow growth during early life.

The results of this thesis demonstrate that the responses to adult diet are conditioned by early growth. Therefore, longitudinal intervention studies, which include dietary counselling, are essential. A target group should be low birth weight individuals who have increased risk for developing chronic disease in later life and preferably, follow-up should cover the whole life span. These studies would shed important light within the DOHaD field.
7 Conclusions

The results presented in this thesis provide convincing evidence that diet plays a role in the association between early growth and disease risk in later life. These findings support the DOHaD hypothesis which suggests that early growth has long-term effects on health.

The main conclusions of this thesis can be summarized as follows:

1. Small body size at birth was associated with lower consumption of fruits and berries, decreased intake of carbohydrates and sugars and increased intake of fats, which suggest that adult dietary habits may be, in part, programmed during prenatal life. Therefore, dietary counselling would be particularly beneficial for those born with small body size, because they have increased risk of developing chronic disease in later life and may have unhealthy dietary habits.

2. Individuals who were born with low birth weight were especially sensitive to the blood pressure-raising effect of salt. Therefore, they would benefit from restricted salt intake.

3. Growth retardation during early life had adverse effects on postprandial insulin and TG responses. These elevated responses may partly explain the association between slow pre- and postnatal growth and increased risk for metabolic diseases in adulthood.

4. Those who grew slowly during early life had elevated appetite regulatory hormone PYY postprandial responses. This could potentially be one explanation why individuals born with small body size and who grow slowly during infancy are less likely to become obese later in life.
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