



UNIVERSITY OF HELSINKI

<https://helda.helsinki.fi>

Applications of Bacterial Cellulose in Food, Cosmetics and Drug Delivery

Ullah, Hanif; Almeida Santos, Helder; Khan, Taous

2016-08

Springer Netherlands

<http://hdl.handle.net/10138/327360>

Ullah, H, Almeida Santos, H & Khan, T 2016, 'Applications of Bacterial Cellulose in Food, Cosmetics and Drug Delivery', *Cellulose*, vol. 23, pp. 2291-2314. <https://doi.org/10.1007/s10570-016-0986-y>

Downloaded from Helda, University of Helsinki institutional repository. <https://helda.helsinki.fi>
This is an electronic reprint of the original article.
This reprint may differ from the original in pagination and typographic detail.
Please cite the original version.

Applications of bacterial cellulose in food, cosmetics and drug delivery

Hanif Ullah^{1,2}Hélter A. Santos^{2,*}

Phone +358-294159661

Email helder.santos@helsinki.fi

Taus Khan^{1,*}

Phone +92-992383591-5

Email tausokhan@ciit.net.pk

¹ Department of Pharmacy, COMSATS Institute of Information Technology, Abbottabad, 22060 Pakistan² Division of Pharmaceutical Chemistry and Technology, Faculty of Pharmacy, University of Helsinki, 00014 Helsinki, Finland

Abstract

Bacterial cellulose (BC) is a versatile biopolymer with better material properties, such as purity, high degree of porosity, relative high permeability to liquid and gases, high water-uptake capacity, tensile strength and ultrafine network. This review explores the applications of BC and its hydrogels in the fields of food, cosmetics and drug delivery. Applications of BC in foods are ranging from traditional dessert, low cholesterol diet, vegetarian meat, and as food additive and dietary aid to novel applications, such as immobilization of enzymes and cells. Applications in cosmetics include facial mask, facial scrub, personal cleansing formulations and contact lenses. BC for controlled drug delivery, transdermal drug delivery, dental drug delivery, protein delivery, tissue engineering drug delivery, macromolecular prodng delivery and molecularly imprinted polymer based enantioselective drug delivery are also discussed in this review. The applications of BC in food and cosmetics provide the basis for BC-based functional foods, nutraceuticals, cosmeceuticals and medicated cosmetics. On the basis of current studies, the BC-based drug delivery could be further fine-tuned to get more sophisticated control on stimuli-responsive drug release. Along with the currently available literature, further experiments are required to obtain a blueprint of drug *in vivo* performance, bioavailability and *in vitro-in vivo* correlation.

Keywords

Bacterial cellulose
Cosmetics
Cosmeceuticals
Deracemization
Drug delivery
Food
Nutraceuticals
Protein delivery
Tissue engineering

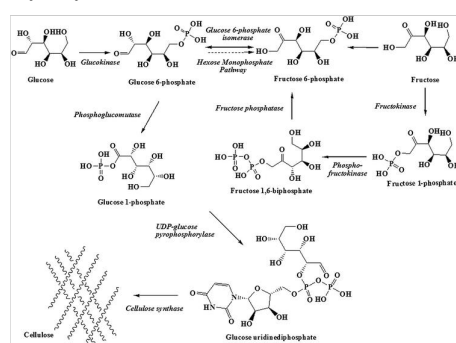
Introduction

Biomaterials play a vital role in the daily life of humans (Czaja et al. 2007; Hubbell 1995; Ratner and Bryant 2004; Shoicher 2009). The importance of biopolymers in our food applications, and personal and medical care cannot be ruled out (Ellis and Smith 2008; Murphy 2001; Jay et al. 2008). Cellulose is the most abundant biopolymer on the surface of earth with 1.5×10^{12} tons annual production (Czaja et al. 2004; Klemm et al. 2005), and most commonly it is obtained from plants (Siró and Plackett 2010).

In addition to plant cellulose (PC), cellulose is also obtained through *in vitro* synthesis with the help of enzymatic pathways, the chemical synthesis from glucose derivatives and the biosynthesis by various microorganisms, such as algae and fungi (Klemm et al. 2005), as well as various aerobic non-pathogenic bacteria of the genera *Agrobacterium*, *Sarcina*, *Rhizobium* and *Acetobacter* (Dufresne 2013; Khan et al. 2007; Shezad et al. 2010). While studying acetic fermentations in 1886, Brown reported the bacterial cellulose (BC) in the form of a strong white gelatinous pellicle on the surface of a liquid medium, which had a thickness up to 25 mm. The microbe responsible for this BC membrane (BCM) was called *Bacterium xylinum* that was later on renamed as *Acetobacter xylinum* (A. *xylinum*) and at the moment is recognized as *Gluconacetobacter xylinus* (G. *xylinus*) (Brown 1886a, b).

G. *xylinus* is the most extensively used microorganism in the basic and applied studies for BC production because of its higher productivity, and capability to consume different sugars and other compounds as sources of carbon (Ross et al. 1991; Saxena and Brown 2012). G. *xylinus* cultivated under controlled conditions with suitable nitrogen and carbon sources, produces highly porous BC network structures in the form of sheets or pellicles, subject to the culturing approach (Lin et al. 2013; Pircher et al. 2014). The culturing conditions may be agitated or static, and batch, semi-continuous or continuous cultivation (Lin et al. 2013, 2014; Pircher et al. 2014; Sulaeva et al. 2015). Typically, the synthesis of BC occurs in four enzymatically catalysed steps: (a) glucose is phosphorylated to glucose-6-phosphate; (b) glucose-6-phosphate is isomerized to glucose-1-phosphate; (c) glucose-1-phosphate is converted to uridine diphosphate glucose (UDP-glucose); and (d) finally glucan chains are synthesized from UDP-glucose (Ross et al. 1991). After this, the parallel glucan chains are aggregated and crystallized to form microfibrils followed by aggregation of the latter to discontinuous bundles of cellulose fibres (Iuchi et al. 2000; Lin et al. 2013). After removal of the culture medium and complete washing, colourless, odourless and tasteless BC is obtained in the form of a gel. This gel finds several applications in our life (Lin et al. 2013). The biosynthetic pathway of BC in G. *xylinus* is shown in Fig. 1.

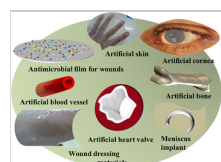
Fig. 1
Biosynthetic pathway of BC in G. *xylinus*.
Adapted with permission from Lin et al. (2013)



The auspicious properties associated with BC, such as exceptional mechanical characteristics, stress-strain behaviour, good light transmittance, *in situ* moldability, porosity, stability, biocompatibility, low immunogenic potential, and capability for cell adhesion, migration and proliferation (Helenius et al. 2006; Millon and Wan 2006; Qiu and Netravali 2014; Svensson et al. 2005) make it appropriate biomaterial for tissue engineering applications. These applications include, but are not limited to [artificial cartilage](#) (Nimeskern et al. 2013; Svensson et al. 2005); [artificial bone](#) (Zimmermann et al. 2011); [artificial cornea](#) (Hui et al. 2009); [heart valve prosthesis](#) (Millon and Wan 2006); [artificial blood vessels](#); [artificial cartilage](#) (Nimeskern et al. 2013; Svensson et al. 2005), [bone](#) (Zimmermann et al. 2011), [cornea and blood vessels](#) (Hui et al. 2009; Klemm et al. 2001; Wan et al. 2011), [heart valve prosthesis](#) (Millon and Wan 2006), [nerve surgery](#) (Klemm et al. 2001; Wan et al. 2011), [meniscus implant](#) (Bodin et al. 2007), [artificial skin and skin tissue repair](#) (Fu et al. 2012; Qiu and Netravali 2014).

Some of these BC-based applications of BC are shown in Fig. 2.

Fig. 2
Applications of BC in biomedicine



However, in the current review, we have focused mainly on the applications of BC in various fields, ranging from conventional food to modern functional foods, cosmetics, nutraceuticals, cosmeceuticals and drug delivery. This review will provide the readers an understanding of the role of BC as a functional additive, formulation stabilizer, biocatalysts platform, and ingredient for food, cosmetics and drug delivery systems. Furthermore, the review will be helpful for academic researchers and formulation scientists in food, cosmetics and pharmaceutical industries to give new insights to BC in terms of designing some novel BC-based functional foods, nutraceuticals, cosmeceuticals and drug delivery systems.

Better material properties of BC for food cosmetics and drug delivery applications

Although similar in chemical structure, BC has different and superior physical, mechanical and biological features to PC. Due to these superior properties, it finds applications in food, cosmetics, biomedicine and drug delivery (Lin et al. 2013). BC is biosynthesized in its purest form, which is entirely devoid of pectin, hemicelluloses and lignin (Chawla et al. 2009). Hence, BC is capable to be easily refined in comparison to PC (Shi et al. 2014b). As-synthesized, innate or pristine BC is highly porous in nature with high permeability to liquid and gases, and possesses high water-uptake capacity (more than 90 % of its weight) (Klemm et al. 2001). These characteristics of BC are due to the ultrafine network of the ribbon-shaped micro- and nanofibrils (Chawla et al. 2009), which are about 100-fold more thinner than the PC fibres. Such properties of BC make it suitable for application as formulation stabilizer, thickener, and for scrubbing and exfoliation without damaging the skin due to its soft texture of small fibres (Lin et al. 2015; Shi et al. 2014b). Moreover, the biocompatibility, low immunogenic potential and *in situ* foldability further enhance the applicability of BC in the field of biomedicine (Lin et al. 2016). These properties together with tensile strength (Campano et al. 2015) make BC a suitable candidate for dermal applications in cosmetics, and topical and transdermal drug delivery. Due to high water holding and ion exchange capacities, BC is an appropriate material for laxative effects and low cholesterol diet, respectively. Similarly, the transparency (Campano et al. 2015), along with high permeability to liquid and gases, makes BC an appropriate agent for facial mask, contact lenses and drug delivery to wound with ease of wound inspection (Czaja et al. 2006). The individual fibre strength of BC is almost comparable to that of Kevlar and steel (Yano et al. 2005), making it suitable candidate for applications, where high tensile strength is required, e.g., drug loaded bone cement. Several material characteristics of BC for applications under the scope of the current review are depicted in Fig. 3.

Fig. 3
Better material properties of BC

Table 1
BC applications in food

| Food, related item or process | Form of BC | Purpose of BC | References |
|-------------------------------|---------------------------------|--|--|
| Nata de coco | BC slices | Main structure | Iguchi et al. (2000) |
| Low cholesterol diet | Powdered BC | Fat adsorbent | Chau et al. (2008), Lin and Lin (2004), Stephens et al. (1990) |
| Vegetarian meat | BC sheets | Structural component, fat adsorbent | Júzová et al. (1996), Purwadaria et al. (2010), Wonganu and Kongruang (2010) |
| Pasty food and jams | Aqueous paste | Heat-stable suspending and bulk forming agent | Okizama et al. (1992) |
| Tofu | Aqueous paste | Gelling agent | Okizama et al. (1993) |
| Kamboko | Aqueous paste | Hardening agent, texture modifier | Okizama et al. (1993) |
| Chocolate drink | Aqueous paste | Stability against heat | Okizama et al. (1993) |
| Ice-cream | Aqueous paste | Hardening agent, stability against freeze-thaw process | Okizama et al. (1993) |
| Glucosylase | Beads | Solid support to increase enzymatic activity | Wu and Lia (2008), Wu et al. (2013) |
| Wine | BC pieces | Solid support to increase activity of yeast | Montelegre et al. (2012), Nguyen et al. (2009), Ton and Le (2011) |
| Fungal laccase | BC sponge | Solid support to increase activity of laccase | Chen et al. (2015) |
| L-lysine | BC cubes | Solid support to increase activity and cell viability | |
| Food packaging | BC sheets, film and powdered BC | Hydrophobic and antimicrobial packaging | Dobre et al. (2012), Jpa et al. (2012), Tomé et al. (2010) |

Cosmetics applications

Cosmetics are substances that are used to improve some of the organoleptic properties of the human body (Hasan et al. 2012). Cosmetics include products that are applied to the human body for altering the appearance, enhancing the attraction, and cleansing or beautifying the body parts without affecting the normal body functions or structure (Hasan et al. 2012). Currently, the majority of the cosmetics are used by customers to boost their beauty without bearing in mind the ill-effects on body, for example, toxicity concerns associated with parabens (Nagel et al. 1977; Darbre and Harvey 2008). In order to avoid harmful effects to the consumers, natural skin-care products are recommended, which utilize herbal or natural ingredients (Hasan et al. 2012).

In this context, cellulose fibrils are applied in cosmetics to stabilize oil-in-water (O/W) emulsion without the addition of any of surfactant. Such formulations may not be irritant to sensitive skin due to the absence of any surfactant (Hasan et al. 2012). BC has also been reported to be an exceptional non-allergenic biopolymer for use in the cosmetics. The various application of BC in cosmetics are discussed in the following sections.

Facial mask

BC facial masks are of great interest as cosmetic devices to treat dry skin due to its biodegradability, low toxicity and ability to hydrate the skin (Annuakitt et al. 2011). In a study, one group of volunteers was asked to apply moist towels on the face for 25 min, while the second group was asked to apply the translucent BC facial masks for the same period (Annuakitt et al. 2011). During the subsequent week, the groups were interchanged to the alternative treatment. Skin dullness, texture, elasticity, sebum content, moisture content and desquamation levels were evaluated using a system used for routine skin counselling before applying and after 5 min of removing the towels and trial product (Annuakitt et al. 2011). The user satisfaction about the BC mask was also investigated. The BC masks augmented the moisture contents of the skin significantly than moist towels upon a single treatment. No obvious effects on other characteristics of skin were observed. The cellulose-mask product BC facial masks rated around 4 out of 5 on the satisfaction rating scale (Annuakitt et al. 2011). The BC mask could be used as a natural cosmetic product for increasing moisture content of the skin can be used for increasing moisture content of the skin. The responses about user satisfaction in questionnaire-based study revealed that the BC facial mask was acceptable to consumer (Annuakitt et al. 2011).

Similarly, BC with and without glycerine was evaluated for skin irritation potential in human subjects (Almeida et al. 2014). There was no significant difference in terms of transepidermal water loss (i.e., absence of barrier disruption) and erythema with zero clinical score, except for few subjects with mild skin irritation. Moreover, addition of glycerine gave a significantly higher skin moisturizing effect, suggesting its potentials for moisturizing facial mask (Almeida et al. 2014).

In a patent, BC facial mask was fabricated with holes for eyes, mouth and nose (Zhong 2008). The author claimed that such mask may be suitable for repeated or prolonged use for skin beautifying purpose, skin nutrition, and moisturizing and cosmetic effects (Zhong 2008). Similarly, facial mask composed of BC membrane containing ginseng extracts has shown promising results in terms of moist feel, overall user satisfaction and skin elasticity in women over 30 years of age (Lee et al. 2011). In another study, BC facial mask with sodium bicarbonate (5 g), monohydrated citric acid (4 g), ascorbic acid (0.5 g) and salicylic acid (0.05 g) has been used for exfoliative and brightening purposes (Legendre 2008). In this study, the author has also claimed BC mask with thermal plasty as a constituent for its anti-wrinkle effects (Legendre 2008).

BC gel with controlled release of silk sericin were developed with improved moisture holding capability in comparison to commercially available paper mask (Aranwit and Bang 2014). Upon peel test using porcine skin, it was revealed that BC-based gel was biocompatible and less adhesive (peeled without pain) than paper mask (Aranwit and Bang 2014). The prepared gel may find potential applications in medicated cosmetics as anti-wrinkle, antiaging and moisturizing facial mask.

Keeping in view the above studies, it is worth mentioning that in addition to the aforementioned application, BC-based membranes could also be used for treating various skin conditions including xerosis, atopic dermatitis and psoriasis, whereby moisturizing effect is needed in addition to pharmacotherapy.

Facial scrub and medicated cosmetics

A facial scrub containing powdered BC and natural ingredients including olive oil, ascorbic acid (Vitamin C), *Aloe vera* extract and powdered glutinous rice was formulated (Hasan et al. 2012). Using plate-plate rheometer, both commercial and formulated facial scrubs showed shear thinning behaviour (non-Newtonian liquid). The formulated facial scrub possessed relatively higher viscosity at lower shear rates in comparison to the commercial one, but both possessed nearly comparable viscosities at higher shear rates. The tested samples were capable of drying out after 10 min at ~30 °C (room temperature). This novel formulated facial scrub containing BC as major ingredient engrosses the attention of cosmetics formulators for the development of facial scrub with natural ingredients, making it safe for skin. Moreover, Lin et al. (2015) claimed cosmetic containing fragments of BC film in the range of 0.05–1.0% by weight. By adding the fragments of BC in the cosmetic not only improved the dermal permeation of active ingredients present in the cosmetic, but also provided skin moisturizing function, sebum absorption and skin exfoliation (Lin et al. 2015). It has also been claimed that due to high water holding capacity and good gas permeability, BC is an appropriate carrier for cosmetically active ingredients including moisturizers, such as salicylic acid or hyaluronic acid whitening ingredients, such as kojic acid or ursolic acid, anti-wrinkling agents (e.g., polypeptides, and exfoliator), growth factors, enzymes, or a combination thereof (Lin et al. 2015). Moreover, according to authors (Lin et al. 2015; Tourmilhae and Lorant 2005), BC-based formulation can find extensive applications in designing the lip, skin and nail care products and long-lasting perfume.

Personal cleansing formulations

The purpose of personal cleansing formulations is remove dirt, reduce sebum and exogenous contaminants, and to control malodour and the skin microflora. In addition to hygienic benefits, surfactants in such formulations damage skin constituents and may entangle in the stratum corneum after washing (Kuehl et al. 2003; Walters et al. 2012). This can lead to allergic reactions and skin irritation, especially in case of sensitive skin (Draelos et al. 2013; Kuehl et al. 2003). In this regard, BC produced biosynthesized in agitated culture conditions (Ag-BC) showed exhibited the highest stabilizing effect for O/W emulsion among all the inspected cellulose-based materials (Ougiya et al. 1997). It was demonstrated that BC fine fibrils acted as a scaffolding structure and a mechanical barrier, interrupting the coalescence of oil droplets. Thus, the emulsion was stabilized without reducing the interfacial tension as occurs in the case of surfactants (e.g., sorbitan monolaurate). Due to its thinner fibrils, Ag-BC would protect a larger surface area of the droplets of oil in the form of mechanical barrier than any other cellulose-based material. Moreover, this emulsion was also stable against changes in temperature and pH, and against addition of salt in comparison with xanthan gum- and sorbitan monolaurate-based formulations. One of the potential applications of this O/W type emulsion could be the formulation of body parts cleansing products, especially for sensitive skin.

In a patent, a personal cleansing formulation consisting of liquid matrix, i.e., water, a lathering surfactant and an external structuring agent, comprising both BC network and a cationic polymer e.g., cationic starch derivatives and cationic cellulose derivatives or mixtures of these, was claimed to be formulated (Heath et al. 2012). The particles of these formulations were suspended in the liquid matrix with pH-values of less than ca. 4.0 or 7.0. Such compositions provided good lathering and easily rinse off properties without any unwanted filmy or slimy hand feel. The presence of particulate matters improve cleansing and exfoliation with conditioning benefits, and without any irritation or damage to the skin. A pH-value less than ca. 4.0 is especially preferred for salicylic acid formulations (Heath et al. 2012). Such formulation may be used for body cleansing for sensitive skin without any irritation, especially for body parts, where consumer's hand feel is important. Moreover, using compositions of such formulation at pH-value of less than ca. 4.0, salicylic acid formulation for personal cleansing can be formulated. Such formulations may be used to clear and prevent skin blemishes and pimples. These may also be used for the treatment of skin conditions with scaling or skin overgrowth (Heath et al. 2012).

Contact lenses

Other than optical indications, contact lenses find wide range of applications including cosmetic or decorative purpose (Rubinstein 2003; steinmann et al. 2005). BC is one of the potential candidates for fabrication of contact lenses due to its transparency, light transmittance, and permeability to liquid and gases. BC-based contact lens was fabricated by pouring high viscosity BC solution (in 1-butyl-3-methylimidazolium chloride) to a mould. Upon treating the solution with isopropyl alcohol followed by water, a clear BC membrane was precipitated, which spontaneously detached from the mould surface and the residual solvent diffused to the water. The hydrated BC contact lens retained its shape and transparency for a time of more than 8 weeks (Levinson and Glonek 2010). Similarly, transparent polymeric hydrogel was prepared by combining BC and 2-hydroxyethyl methacrylate polymer. The fabricated biomedical possessed ca. 40% (w/w) water content with good mechanical strength and integrity (Li et al. 2010) and had the ability to be used as contact lenses. Apart from optical and decorative purposes, these contact lenses can find potential applications for drug delivery to the cornea. Moreover, the ability of BC to take colour of the medium (Shi et al. 2014b) can be exploited for design of coloured and appealing contact lenses with transparent centre for the pupil, provided that the biocompatibility is not compromised by the colourant(s).

The applications of BC and BC-based products in cosmetics are summarized in Table 2.

Table 2
Applications of BC in cosmetics

| Cosmetic product | Form of BC | Purpose of BC | References |
|---|--|--|----------------------------|
| Facial mask | BC sheets | Moisturizer | Annuakitt et al. (2011) |
| Facial mask | BC-glycerine composites | Moisturizer | Almeida et al. (2014) |
| Facial mask | BC membrane | Moisturizer | Zhong (2008) |
| Facial mask | BC-ginseng | Moisturizer and carrier | Lee et al. (2011) |
| Facial mask | BC with cosmetically substances | Moisturizer and carrier for the actives for exfoliative, brightening and anti-wrinkle purposes | Legendre (2008) |
| Facial mask | BC-sericin composites | Moisturizer and carrier for silk sericin | Aranwit and Bang (2014) |
| Facial scrub | Powdered BC | Viscosity enhancer | Hasan et al. (2012) |
| Facial scrub | BC fragments | Moisturizer, sebum absorber and skin exfoliator | Lin et al. (2015) |
| Carrier for cosmetically active ingredients | BC fragments | Prolongs the contact time of the cosmetically active ingredient with the skin surface | Lin et al. (2015) |
| Foundation make-up | BC fragments | Stable make-up with less number of touch-ups and lesser amount required | Lin et al. (2015) |
| Personal cleansing product | BC fibres (synthesized in agitated conditions) | Surfactant free emulsion for sensitive skin | Ougiya et al. (1997) |
| Personal cleansing product | BC particles | Cleansing and exfoliation | Heath et al. (2012) |
| Contact lenses | Regenerated BC sheet | Film-forming agent | Levinson and Glonek (2010) |
| Contact lenses | BC-based hydrogel | Film-forming agent | Li et al. (2010) |

Drug delivery applications

Drug delivery to wounds

Studies have suggested that fluid, particularly exudates from chronic wounds may inhibit healing process (Wovden and Wovden 2003). An excessively wet environment may lead to wound and skin maceration resulting in prolonged wound healing, whereas a dry wound will also heal more gradually due to lack of moisture required for cell migration (Benbow and Stevens 2010). Hence, exudates reduction looks like a key parameter for normal healing process (Sulava et al. 2015). Being an excellent absorbent (Gayathri and Gopalaswamy 2014) and skin moisturizer (Annuakitt et al. 2011), BC can be an ideal candidate for lowering or removing the wound exudates, while at the same time maintaining a moist environment (Sulava et al. 2015). However, innate BC is devoid of antimicrobial activity against the wound deteriorating pathogens.

To achieve such goals, a BC film with antibacterial property was fabricated, whereby a lyophilized BC film was dipped in a benzalkonium chloride (BZK) solution followed by further lyophilization (Wei et al. 2011). Water uptake capacity, a feature important for wound dressing system, was also attained with a swelling ratio of 37.3 and 26.2 for saline solution and deionized water, respectively. A prolonged (at least 24 h) stable antibacterial activity was achieved against *Staphylococcus aureus* along with a higher water uptake capacity. Thus, BZK-loaded BC film may act as a potential functional wound dressing system for treatment of acute traumas.

Recently, Pavaloiu et al. (2014b) studied the release of the antibiotic amoxicillin (AMX) from BCM at nearly neutral (7.4) pH conditions. The concentration of AMX significantly influenced the drug release (Pavaloiu et al. 2014b). Among the other factors, there was a significant contribution of glycerol as plasticizer to *in vitro* drug release. The common topical drug delivery enhancer cetyl trimethyl ammonium bromide did not show any positive impact on the *in vitro* release of the drug. This system might provide a suitable way for antibiotic delivery to the wound.

The antimicrobial activity of antibiotics with prolonged drug release behaviour from BC was assessed *in vitro* using ampicillin (AMP) and gentamycin (GM) (Kaplan et al. 2014). For the assessment of exudate retention, the water uptake capacity of the BCM was found to be $65.6 \pm 1.6\%$ in phosphate buffer saline (PBS). The drug loading was 99 and 48 mg/cm² for AMP and GM, respectively. The BCM released only trace amount (0.107% of AMP and 0.113% of GM) within 24 h. Thus, with no burst release, the amount of drug released within 7 days was 28 and 17% for AMP and GM, respectively. Furthermore, due to sufficient amount of the drug in prolonged release manner, the antibacterial activity against *Pseudomonas aeruginosa*, *Staphylococcus epidermidis*, *Staphylococcus aureus*, *Enterococcus faecalis* and *Escherichia coli* prevailed for as long as 3 days of incubation period (Kaplan et al. 2014). Furthermore, Rouabhi et al. (2014) covalently attached the GM to the surface of chemically modified BC, i.e., BC-RGDC-GM, where RGDC is a peptide composed of arginine, glycine, aspartic acid and cysteine. In addition to its bactericidal effect, BC-RGDC-GM was devoid of any toxicity for human skin fibroblasts (Rouabhi et al. 2014). Likewise, the *in vitro* release of tetracycline from BC was assessed as function of electron beam irradiation, which considerably decreased the diffusion of tetracycline (Stoica-Guzun et al. 2007). Hence, BC-based drug delivery to wound is a feasible way with its ability to absorb exudates as well. Furthermore, silver sulfadiazine (SSD) particles impregnated into BC revealed *in vitro* antimicrobial activity, human epidermal cells biocompatibility, and good epithelialization and wound healing activity in rat models with burns in a rat model (Luan et al. 2012; Wen et al. 2015). A step further, a novel wound dressing system consisting of polyhexamethylene biguanide (PHMB) and the never dried BC (BC-PHMB) was evaluated for safety through *in vitro* cytotoxicity, haemolysis, sensitization in guinea pigs, irritation potential in rabbits and acute systemic toxicity (Serafica et al. 2010). In addition to good antimicrobial effects, wound healing in animal models showed that the BC-PHMB wound dressing rehabilitated 70% of the wounds in comparison to 0, 20 and 50% for the air-exposed, hydrogel treated and hydrocolloid treated wounds, respectively. Upon clinical effectiveness testing in humans, BC-PHMB showed promising results for wound healing of deep pressure wounds and venous leg ulcers (Serafica et al. 2010). Furthermore, in patient, this wound dressing system was more promising than the other commercial dressing in terms of bacterial load reduction and pain management (Haemmerle et al. 2012).

Though not a drug delivery system, BC has also been investigated as promising antimicrobial film for wound dressing with various agents, such as deacetylated chitosan (Butchosa et al. 2013), chitosan (viral protective membrane) (Wanling et al. 2012), montmorillonite (Ul-Islam et al. 2013), and nanoparticles of silver (Dobre and Stoica-Guzun 2013), copper (Pinto et al. 2013) and titanium dioxide (Khan et al. 2015). However, such metallic nanoparticles are infamous due to their possible concerns with human health, such as hepato-, neuro-, photo-, geno-, cyto- and dental toxicity, formation of oedema, and hyperplasia (Koochi et al. 2011; Lu et al. 2010; Prabhu et al. 2010; Ray et al. 2009; Sanberg et al. 2010; Wang and Wang 2014; Wang et al. 2014). The histopathological changes in the bones, hearts and kidneys of guinea pigs, and lack of studies about toxic effects with prolonged use of such nanoparticles further limit their practical applications (Korami et al. 2013).

Tissue engineering drug delivery

BC-based materials have also been demonstrated for drug delivery applications in the field of tissue engineering and regeneration. In this scenario, Mori et al. (2011) studied the release of antibiotics (GM and vancomycin) from a BC-based bone cement. It was demonstrated that **incorporating BC into the bone cement prevented compression and fracture fragility, improved fatigue life and increased antibiotic elution by incorporating BC into the bone cement, the compression and fracture fragility were prevented, while fatigue life and antibiotic elution were enhanced** (Mori et al. 2011). Such antibiotic containing BC-based cements may have clinical relevance, when high levels of antibiotic release are required, while the mechanical properties of the cement are not compromised. In addition, bone morphogenetic protein-2 (BMP) loaded BC was investigated for localized delivery system with osteogenic potentials in tissue engineering (Shi et al. 2012). The system was enough biocompatible and was capable (*in vitro*) to differentiate the mouse fibroblast-like C2C12 cells into osteoblasts. Upon *in vivo* studies on subcutaneous implants, the BMP₂ loaded BC scaffold was capable for bone formation with higher calcium concentration than the pristine BC scaffolds (Shi et al. 2012). Hence, it can be concluded that BC is a good carrier for localized delivery of therapeutic candidates, such as BMPs in tissue engineering.

Controlled drug delivery

Frequent dosing, fluctuation in plasma drug concentration and patient non-compliance associated with shorter half-lives of drugs necessitate such drugs to be formulated into controlled release dosage forms. Researchers have made some attempts in order to control the drug release from a BC-based delivery systems. For this purpose, Amin et al. (2012a-b) (2012a) reported the use of powdered BC to coat paracetamol tablets using a spray-coating technique (Amin et al. 2012a). The study demonstrated that BC formed high quality, foldable, flexible and uniform soft films without adding any plasticizer that was comparable to the film of ethyl cellulose aqueous dispersion (Aquacoat ECD). *In vitro* drug release rate was dependent on the BC film thickness and was slower (200 min for maximum release) for coated tablets with 200 µm thick film, than uncoated tablets (i.e., 100 min for maximum release) (Amin et al. 2012a).

In another study, Pávaiólu et al. (2015) described the swelling behaviour of mono- and multilayer hydrogels based on BC and gelatin (BC-G). The findings indicated that the swelling of BC-G hydrogels was higher in acidic pH as compared to the basic one due to the polyelectrolyte character of gelatin. Moreover, the concentration of gelatin had a direct relation with the swelling of BC-G hydrogel, while the coating of hydrogel with additional BC has inverse effects on the swelling rate (Pávaiólu et al. 2015). Due to its swelling in acidic condition of the stomach, the hydrogels may find potential applications in gastro-retentive drug delivery.

Amin et al. (2014) studied the potential of stimuli-responsive BC-g-poly(acrylic acid-co-acrylamide) hydrogels synthesized by graft copolymerization using the microwave irradiation technique for oral controlled drug delivery (Amin et al. 2014). The hydrogels were suitable for drug loading due to the highly porous morphology. Being pH-responsive, swelling of hydrogels was less in acidic media, reaching maximum swelling at neutral pH. Similarly, the hydrogels exhibited lesser drug (theophylline) release in SGF than SIF (Amin et al. 2014). Hence, it was suggested that such type of hydrogels may be suitable for drug delivery to the lower parts of the gastrointestinal tract, e.g., peptides, proteins, and acid-labile drugs, and targeted delivery in colonic diseases.

In another sustained drug release study, BC-based hydrogels in combination with carboxymethyl cellulose (BC-CMC) were investigated for controlled drug delivery using ibuprofen sodium (IbuNa) as a model drug (Pavaliou et al. 2014c). The results of this study showed that the CMC content and epichlorohydrin (cross-linker) concentration influenced the swelling and drug release properties of the hydrogels, which were governed by pseudo-Fickian diffusion. These preliminary findings suggested that BC-CMC hydrogels could be exploited as components in controlled drug delivery applications.

Likewise, mono- and multilayer films of BC, PVA and chitosan (BC-PVA-chitosan) have been reported for controlled release of IbuNa (Pavaliou et al. 2014a). The drug release was pH sensitive, which followed the Fickian model of diffusion. Moreover, the rate of drug release was inversely proportional to the concentration of BC in the film with pronounced effect in case of multilayer films. Shi et al. (2014a) fabricated hybrid hydrogels of BC and sodium alginate (SodAl) as a dual-stimuli-responsive system. The pH and electric field stimulus-responsive swelling and drug release behaviours of the BC-SodAl hydrogels were investigated *in vitro* using Ibu as a model drug. The swelling ratio was lower at acidic pH (less than 8-fold), while higher at alkaline pH (more than 13-fold). The electric field of 0.5 V increased the swelling ratio from 8-fold (at 0 V) to 14-fold. The release of Ibu was slower in acidic conditions and faster in alkaline conditions (Shi et al. 2014a). Furthermore, the drug release from the BC-SodAl hydrogels could be boosted with the application of an electric stimulus (Shi et al. 2014a). The BC-SodAl hybrid hydrogels with both pH- and electro-response are therefore new auspicious candidates for oral controlled drug delivery.

Proteinaceous therapeutic candidates have an extended role in several fields of medicine, such as diagnostics, vaccines, inflammatory diseases and cancer (Malik 2008). The increased use of pharmaceutical proteins could be justified by some beneficial properties in comparison to small-molecule drugs (Vermonden et al. 2012). However, the subtle 3D conformation of proteins is a limitation to the use of such therapeutic candidates due to chemical and proteolytic degradation, aggregation and physical unfolding (Bruno et al. 2013; Manning et al. 1989, 2010; Yang 2015). This kind of instability always results in loss of bioactivity and frequently provokes an immune response (De Groot and Martin 2009; Kalyaperumal and Jing 2009). Furthermore, oral administration of proteinaceous drugs is trickier due to acidic pH and high proteolytic activity of stomach that may lead to destabilization and degradation of the protein structure (Vermonden et al. 2012). In addition, first-pass effect of the liver, fast renal clearance and consequently the short half-lives of proteinaceous drugs require frequent intravenous administration that is associated with patients' discomfort, inconvenience and non-compliance (Harris and Chess 2003; Tang et al. 2004). Due to the abovementioned limitations, the delivery of proteins is an immense challenge in the field of modern medicine. Fabrication of hydrogels is one of the approaches for the improvement of pharmacodynamics and pharmacokinetics of proteinaceous drugs (in intact form) with improved patient's compliance (Peppas et al. 2004).

BC possesses abundant number of hydroxyl groups in addition to the hydrophilicity and biocompatibility (Pandey et al. 2014; Sulavea et al. 2015). Such properties enhance the chemical modification capacity with a range of chemical groups, which could modulate the loading and release of drugs from the delivery system. BC has been employed for the oral delivery of protein by fabricating BC-based hydrogels. In this perspective, Ahmad et al. (2014) investigated the stimuli-responsive BC-grafted polyacrylamide (BC-g-PAM) hydrogels for oral delivery of proteins (Ahmad et al. 2014). In this case, BC-g-PAM hydrogels were fabricated with the help of electron beam irradiation without any cross-linker, thus eradicating any potential toxic effects associated with it (Ahmad et al. 2014). The hydrogels showed potential for protection of bovine serum albumin (BSA) from gastric (acidic) environment with <10% BSA release in simulated gastric fluid (SGF). Moreover, the released BSA was stable and bioactive with enhanced penetration across the intestinal mucosal tissue that was evident from *ex vivo* penetration experiment. The fabricated hydrogels were biocompatible, non-toxic and safe for *in vivo* applications (Ahmad et al. 2014).

Mueller et al. (2013) studied BCM for loading and release of BSA as a model protein for delivery systems. It was demonstrated that the protein release was controlled by diffusion. In this study, the never-dried BC had more protein loading than freeze-dried BC, which might be related to the changes in the fibrous network during the process of freeze-drying (Müller et al. 2013). The study also demonstrated that the integrity and bioactivity of proteins could be maintained during the process of loading and release. In another study, BC and polyacrylic acid (BC-PAA) hydrogels were investigated *in vitro* for controlled delivery of BSA as model protein (Amin et al. 2012b). The study demonstrated that BC-PAA hydrogels were pH-dependent with lower swelling ratios (<1000%) below pH 5 and higher (>2000%, being maximum) at pH 7. Consequently, BSA was released in SGF much slower (15% at the end of 2 h) and faster in simulated intestinal fluid (SIF) (8 h to release maximum drug for lowest radiation dose and 13–14 h for the highest) (Amin et al. 2012b). The difference in release rates as function of pH were due to the different swelling rates of BC-PAA due to change in pH.

This clearly demonstrated the potential of BC for pH-responsive delivery system of proteinaceous and non-proteinaceous drugs. Such types of drug delivery systems have the capability for controlled oral delivery of peptides, proteins and acid-labile therapeutic candidates.

Enantioselective drug delivery

Approximately more than 50% of the drugs in practice exist as racemates and about 90% of these are marketed as racemic mixtures of an equimolar ratio of two enantiomers (Nguyen et al. 2006). Enantioselective drug delivery and deceleration are key processes in modern medicines and are predominantly significant in the field of pharmaceuticals, as the different diastereomers or enantiomers of a therapeutic candidate often have different bioactivities (Nguyen et al. 2006). Therefore, it is necessary to promote deceleration in pharmaceutical industry and clinical settings to eliminate the unwanted isomer from the product and deliver the desired isomer for optimal treatment, as well as a rational therapeutic control over the patient. In this domain, Bodhibukkana et al. (2006) fabricated BC-based molecularly imprinted polymeric (MIP) matrix system for the enantioselective delivery of **from the racemic mixture of propranolol-S-propranolol** (from its racemic mixture) through transdermal route. In this study, MIP matrix system with specific enantioselective binding sites for **drug-S-propranolol was obtained/fabricated by *in situ* copolymerization of methacrylic acid, using in the presence of ethylene glycol dimethacrylate as cross-linking agent** (Bodhibukkana et al. 2006). S-propranolol was used as a template molecules, which was removed later on. This MIP matrix system exhibited an enantioselective transport of S-propranolol. The enantioselectivity for S-propranolol was also revealed by the *in vitro* release of enantiomers employing the skin of rat (Bodhibukkana et al. 2006). Thus, the BC-based MIP membrane might have great potential for application in enantioselective drug delivery system through transdermal route in clinical settings, and deceleration of racemates of propranolol and other racemic drugs.

Dental drug delivery

Dental caries may promote to dental pulp infection, which needs a procedure, known as root canal treatment (RCT), of the affected tooth. The major aim of RCT is to thoroughly decontaminate the root canal system. The morphology of root canal is too complex to access in many humans. In addition, relapse of dental pulp infections is likewise common. In conventional RCT, a paper point made of PC or cotton pellet is employed in order to dry and sterilize the dental root canal. For such sterilization, high absorbency for residue, high biocompatibility and efficient intracanal medication delivery is desired. To achieve this, Yoshino et al. (2013) designed a pointed form of BC with its usability as a novel biomaterial for RCT (Yoshino et al. 2013). BC **in pointed form/point** exhibited outstanding **expansion and absorption/absorption and expansion than comparison to the conventional paper points**, with a higher tensile strength in wet form. Moreover, BC releases more drug than that from conventional paper points. Owing to the abovementioned finding, BC-drug composite has great potential for dental drug delivery and treatment of RCT.

Transdermal drug delivery

Transdermal drug delivery provides an attractive alternate route to both oral drug delivery and hypodermic injection (Prausnitz and Langer 2008; Prausnitz et al. 2004). Since remote times, folks apply different ingredients on the skin for therapeutic purposes, and in the current age, several transdermal formulations have been developed for delivery of drugs to systemic circulation (Prausnitz and Langer 2008). For the same purpose, BCM with and without plasticizer has the potential for transdermal delivery of therapeutic candidates due to absence of barrier disruption and erythema (Almeida et al. 2014). In addition, skin moisturizing effect and good skin tolerance further strengthens the reported interest of BCM as source for transdermal drug delivery (Almeida et al. 2014). There are few reports that describe the application of BCM for transdermal drug delivery. The rate of drug release can also be tailored by controlling the porosity of BC by physical or chemical means and also by changing the hydrophilicity of the environment. For example, a study was carried out by Olyveira et al. (2013), whereby gamma-irradiated and non-irradiated BCM was studied for *in vitro* drug release in a diffusion cell. It was shown that irradiated BCM has higher pores density than non-irradiated samples, and thus exhibited lower diffusion than the latter one (Olyveira et al. 2013). Likewise, Stoica-Guzun et al. (2007) assessed the effect of electron beam irradiation on the release of tetracycline from BCM as transdermal delivery system. This study showed that electron beam irradiation considerably decreased the *in vitro* diffusion of tetracycline. These findings suggest the potential of BCM in the form of transdermal patches (Stoica-Guzun et al. 2007). Hence, it is concluded that the drug release by diffusion can be tuned by treating BCM with ionizing radiations, giving a new way for physical control over drug release.

Likewise, for therapeutic feasibility in terms of transdermal delivery system, BCM was assessed for the *in vitro* permeation of lidocaine hydrochloride (LHC) and Ibu (model drugs) through human epidermis. The study showed that LHC loaded BCM gave lower permeation rate than that of conventional formulations (Trovati et al. 2012). In contrast, the permeation study for Ibu quite posed apart, as the *in vitro* permeation rate was almost threefold higher for Ibu-loaded BCM than that of Ibu gel or an Ibu solution in PEG400 (Trovati et al. 2012). Diclofenac sodium (DS), belonging to the class of NSAIDs, was loaded into BCM (Silva et al. 2014). Using glycerol as plasticizer, BCM was explored as novel nanostructured transdermal delivery systems for DS salt. The drug containing BCM was quite homogeneous and flexible having substantial swelling behaviours. Using human epidermal membranes, *in vitro* diffusion studies showed that DS loaded BCM had permeation rate comparable to marketed patches of DS and significantly lower than that of a commercial gel formulation (Silva et al. 2014). The simplistic preparation method having easy application and the comparable drug release profile clearly demonstrated the enormous potentialities of utilizing BCM in transdermal delivery of DS and other drugs. In a similar context, Pandey et al. (2013) utilized BC dispersion and solution for the preparation of superabsorbent BC-PAM cross-linked hydrogels by microwave irradiation. The hydrogels exhibited a swelling behaviour (maximum at pH 7). The swelling rate was much higher (ca. 2300–2500%) for hydrogels prepared from BC solution than that of BC dispersion (ca. 900%). Moreover, the hydrogels sustained the release of theophylline in buffers (pH 7.4) for 24 h (Pandey et al. 2013). The study demonstrates the application of this hydrogel for transdermal delivery of theophylline. However, there is still a need for the *in vitro* drug permeation studies to **further** make the feasibility clearer.

In a recent study, Huang et al. (2013) investigated the BCM for the *in vitro* controlled drug release of an alkaloid of isoquinoline group, i.e., berberine. In addition to the transdermal controlled drug release experiments, BCM was also tested in SGF, SIF, and acidic and alkaline solutions. The drug release rate was slower in low-pH fluids (such as SGF), intermediate in alkaline fluid and the highest in near-neutral conditions (such as SIF). The drug release was controlled by diffusion. This type of pH-dependent drug release can be correlated to the swelling of BCM at different pH values (Huang et al. 2013). From these findings, it is evident that BC and BC-based hydrogels are feasible for successful application in transdermal drug delivery, and to modulate the percutaneous drug bioavailability.

Briefly, in most of the studied systems with BC, the release of the therapeutic candidates was controlled by diffusion. The rates of drug release were temperature- and pH-dependent, where the latter affects the swelling of the nanofibers drastically and thus the porosity of the material is altered (Huang et al. 2013; Pandey et al. 2013).

Macromolecular prodrug delivery

Besides gastric irritation (Radi and Khan 2006), one of the major concerns associated with Ibu is the shorter half-life that needs its most frequent dosing with associated side effects (Wright 2002). To avoid these concerns, researchers have tried some novel pH-dependent conjugates of non-steroidal anti-inflammatory drugs (NSAIDs) with different macromolecules (Hussain et al. 2009; Peng et al. 2006). BC gives more opportunities for modification by different methods due to the presence of abundant surface hydroxyl groups (Stenstad et al. 2008). One of such attempts was made by Shi et al. (2013), who developed a novel BC-based conjugates of Ibu by esterification between -OH and -COOH groups of BC and Ibu, respectively (Shi et al. 2013). BC-Ibu as a macromolecular prodrug had the capability to control the drug release via the process of hydrolysis of the ester bond under different pH-conditions. The drug release profiles were dependent on the ester bond hydrolysis, faster in alkaline and acid solution, but relatively slower in neutral pH (Shi et al. 2013). Such pH-dependent drug release suggests a great potential of BC-Ibu as a more effective and stable prodrug candidate. However, this strategy can be further applied to other NSAIDs with carboxyl functional group for the preparation of prodrugs. For example, aspirin could be conjugated to BC to avoid gastric irritation and to target colonic cancer, if the ester bond is sufficiently stable in acidic pH.

All the studies discussed above regarding BC-based drug delivery are summarized in Table 3.

Table 3

Applications of BC in drug delivery

| Purpose | Therapeutic candidate(s) | Strategy | Finding | References |
|------------------------|---|---|---|--|
| Drug delivery to wound | BZK | Drug loaded BCM | Prolonged drug release and antimicrobial activity | Wei et al. (2011) |
| | AMX | Drug loaded BCM | AMX- and glycerol-dependent <i>in vitro</i> drug release | Pavaliou et al. (2014b) |
| | AMP, GM | Drug loaded BCM | Good water uptake capacity, no burst release and prolonged drug release with antibacterial effects | Kaplan et al. (2014) |
| | GM | Covalently attached to the surface of RGDC-modified BCM | Antibacterial effects without toxicity for human skin fibroblasts | Rouabhia et al. (2014) |
| | Tetracycline | Drug loaded BCM | Antibiotics release was sustained by electron beam-irradiation | Stoica-Guzun et al. (2007) |
| | SSD | BCM | <i>In vitro</i> antimicrobial activity, human epidermal cells biocompatibility, <i>in vitro</i> epithelialization and wound healing activity | Luan et al. (2012), Wen et al. (2015) |
| | PHMB | BCM | No <i>in vitro</i> cytotoxicity or haemolysis; no sensitivity, irritation potential or acute systemic toxicity in animals; good antimicrobial effects; promising for <i>in vivo</i> wound healing, and more pain reduction than the commercial dressing | Serafica et al. (2010), Haemmerle et al. (2012) |
| | Deacetylated chitosan, chitosan montmorillonite | BCM | Antimicrobial effects | Butchosa et al. (2013), Wanling et al. (2012), Ul-Islam et al. (2013) |
| | Metallic nanoparticles (silver, copper, titanium dioxide) | BCM | Antimicrobial effects | Dobre and Stoica-Guzun (2013), Pinto et al. (2013), Khan et al. (2015) |
| | Tissue engineering drug delivery | GM vancomycin | BC-based bone cement | Presence of BC in the bone cement prevented compression and fracture fragility, improved fatigue life and increased antibiotic elution The compression and fracture fragility were prevented, while the fatigue life and antibiotic elution were improved |

| Purpose | Therapeutic candidate(s) | Strategy | Finding | References |
|---------------------------------|--------------------------|---|---|----------------------------|
| | BMP ₂ | BC-based protein composite | Biocompatible, and capable of <i>in vitro</i> fibroblast differentiation and bone formation | Shi et al. (2012) |
| Controlled drug delivery | | | | |
| | Paracetamol | BC coated tablets | The flexible BC film sustained the drug release | Amin et al. (2012a) |
| | – | BC-based hydrogels with gelatine | Potentials for gastro-retentive drug delivery due to more swelling in acidic conditions | Pavaliou et al. (2015) |
| | Theophylline | BC-based hydrogels | Lesser drug release in SGF than SIP | Amin et al. (2014) |
| | IbuNa | BC-based hydrogels | pH-dependent sustained drug release | Pavaliou et al. (2014c) |
| | IbuNa | BC-based hydrogels | pH-dependent sustained drug release | Pavaliou et al. (2014a) |
| | Ibu | BC-based hydrogels | pH- and electro-dependent drug release | Shi et al. (2014a) |
| | Propranolol | MIP matrix | Selective transport and release of 5-propranolol | Bodhibukkana et al. (2006) |
| | BSA | BC-based hydrogels without cross-linker | Devoid of cross-linker associated toxicity, protection of BSA from gastric conditions, <i>in vitro</i> sustained release of BSA, which was stable after loading and release | Ahmad et al. (2014) |
| | BSA | BCM | More BSA loading in never-dried BCM, and integrity and bioactivity of BSA was maintained after loading and release | Müller et al. (2013) |
| | BSA | BC-based hydrogels | Lower swelling and lower drug release in acidic pH in comparison to alkaline pH | Amin et al. (2012b) |
| Dental drug delivery | | | | |
| | Tyran blue | BC point | Greater drug release in comparison to paper point | Yoshino et al. (2013) |
| Transdermal drug delivery | | | | |
| | Insulin | BCM | Gamma-irradiated BCM has slower insulin release than non-irradiated BCM | Olyveira et al. (2013) |
| | Tetracycline | Drug loaded BCM | Electron beam-irradiated BCM has slower tetracycline release than non-irradiated BCM | Stoica-Guzun et al. (2007) |
| | Ibu, LCH | Drug loaded BCM | Slower permeation of LCH than conventional formulations, faster permeation of Ibu than conventional formulations | Trovati et al. (2012) |
| | DS | Drug loaded BCM | Permeation rate was comparable to commercial patches and significantly lower than commercial gel | Silva et al. (2014) |
| | Theophylline | BC-based hydrogels | Sustained release of theophylline | Pandey et al. (2013) |
| | Berberine | BCM | Drug release was slower in acidic pH, intermediate in alkaline pH and highest in neutral pH | Huang et al. (2013) |
| Macromolecular prodrug delivery | | | | |
| | Ibu | pH-dependent ester conjugates | Sustained release of Ibu (faster in alkaline and acid pH, while slower at neutral pH) | Shi et al. (2013) |

Conclusion and future prospects

The current review demonstrates that BC is a natural biomaterial with 'GRAS' status, biosynthesized by non-pathogenic bacteria. BC has great potential for application in food, cosmetics and drug delivery systems, in addition to the aforementioned biomedical applications. Nevertheless, there are a limited number of available studies in the field of foods and cosmetics, and there still exists adequate scope for the advanced research in these areas in more detail. For example, cholesterol lowering studies on animals and humans need further attention. Studies on food and cosmetics pave the way for potential applications of BC in nutraceuticals and cosmeceuticals. In terms of nutraceuticals, BC can be further employed in the form of fortified food by adding certain nutritional entities, such as vitamins and minerals to the existing and the new BC-based food items. The traditional dessert may also act as a sweetened vehicle for oral drug delivery, particularly for children. Moreover, due to its bulk-forming and water retention capacity, the fibrous nature of the BC-based food products can be assessed for its laxative effect in the treatment of constipation. BC can be used in O/W emulsion without surfactant, as a substitute to PC and PC-derivatives. This emulsion could be used for cleansing, makeup, and care or treatment of the lips, skin and the eyelashes, as well as treating certain medical conditions of the skin. Furthermore, a comparison between BC produced under static and agitated conditions should be carried out for pure BC in terms of water uptake for sweat retention in cosmetics, exudate retention in wounds, drug loading, drug release, and *in vivo* physical and chemical modifications.

In the modern era, most of the people rely on cosmetics in one way or the other. By addition of antioxidants and therapeutic agents to BC-based cosmetics, a paradigm shift is expected from conventional cosmetics to cosmeceuticals and medicated cosmetics. It is noteworthy that due to its transparency, light transmittance and biocompatibility, BC and/or its hydrogels can be used for the fabrication of disposable and multiple use contact lenses for cosmetic purpose and optometry, and as *ocular* implants for *ocular* drug delivery.

BCM provides a good tool for delivery of antibiotics to the wound with potentials for exudates retention and moisturizing environment that are favourable for wound healing. These studies lack experiments on wound models to study the favourable properties of pristine and antibiotics loaded BCM for *in vivo* wound healing.

The fabrication of BC-based hydrogels has been tested with several polymeric matrices, such as PAA, PVA and PAM. However, many other polymeric biomaterials could be tested while considering specific interactions between carriers and drugs that might tailor the drug release. Despite of several studies regarding the oral delivery of proteins, only BSA as model protein has been studied. There still exists a space for research on *in vivo* loading, release and stability studies of other proteins with therapeutic value.

Regarding transdermal drug delivery, BC-based delivery systems have shown promising results in term of biocompatibility (pure BC) with skin, and *in vitro* drug diffusion studies. However, further *in vivo* studies for skin irritation potentials of drug loaded BC and *in vivo* bioavailability of drugs from the BC-based delivery systems with and without penetration enhancer(s) are needed.

As discussed in some of the abovementioned studies, drug release is controlled by the process of pH-dependent diffusion, which could be further tailored by additional physical treatments or chemical modifications. Such approaches would enable a sophisticated control over the drug release, particularly in a response to body stimuli, such as temperature above normal, i.e., fever condition and tumour micro-environment. Moreover, BC-based nanogels would be helpful for invasive targeted delivery of proteinaceous and non-proteinaceous drug, provided that such nanogels are biodegradable *in vivo*. Such nanogels could also be used in the form of biomolecule-sensitive hydrogels delivery systems, e.g., glucose-sensitive hydrogels for delivery of insulin. Moreover, such biodegradable hydrogels can also be used for targeted drug delivery to tumour (acidic pH), provided that suitable nanogels are formed that are more sensitive to slight changes for invasive drug delivery. In addition, due to the presence of several-OH groups on its surface, BC has also potential for the formation of macromolecular prodrug by covalent conjugation. The BC conjugates can be further tested *in vitro* in the presence of enzymes (microbial esterases and cellulases) and in *in vivo* animal models to predict the *in vivo* performance. Moreover, this approach can be further tailored for pH-dependent (more stable in acidic environment) sustained release of other therapeutic candidates associated with gastric irritation (e.g., NSAIDs). The abundant surface free-OH groups can also be used for surface functionalization of BC for targeted drug delivery, for example, colon-specific drug delivery.

BC alone or in composite form with biocompatible polymer(s) may also find interesting applications in subcutaneous implantable devices for the delivery of therapeutic candidates, where prolonged therapy is desired, such as hormonal replacement therapy and contraception.

In case of MIP, the technique could further be tailored by designing specific MIP to obtain for enantioselective drug delivery for better patient's outcomes, and for enantiomer differentiation and deracemization. Moreover, MIP-based nanoparticle columns with improved surface area can be designed for efficient enantiomer separation, analysis and deracemization.

In case of all drug delivery systems, discussed in this review, there is a need for further *in vivo* studies using various animal models and/or human volunteers for getting a clearer idea about the *in vivo* performance, bioavailability and *in vitro-in vivo* correlation (IVIVC) of the prepared delivery systems.

Acknowledgments

Hanif Ullah is grateful to Higher Education of Pakistan for fully funded indigenous scholarship (213-62780-2BM2-148) and foreign research sponsorship (IRSP 30 PS 20) at University of Helsinki, Finland. Dr. H.A. Santos acknowledges financial support from the Academy of Finland (Grants Nos. 252215 and 281300), the University of Helsinki Research Funds, Biocentrum Helsinki, and the European Research Council under the European Union's Seventh Framework Programme (FP/2007-2013) (Grant No. 310892).

References

- Ahmad N, Anin MCIM, Mahali SM, Ismail I, Chuang VTG (2014) Biocompatible and mucoadhesive bacterial cellulose-g-poly (acrylic acid) hydrogels for oral protein delivery. *Mol Pharm* 11:4130–4142
- Almeida IF, Pereira T, Silva NH, Gomes FP, Silvestre AJ, Freire CS, Sousa Lobo JM, Costa PC (2014) Bacterial cellulose membranes as drug delivery systems: an *in vivo* skin compatibility study. *Eur J Pharm Biopharm* 86:332–336
- Anin MCIM, Abali AG, Ahmad N, Katas H, Jamal JA (2012a) Bacterial cellulose film coating as drug delivery system: physicochemical, thermal and drug release properties. *Sains Malays* 41:561–568
- Anin MCIM, Ahmad N, Halib N, Ahmad I (2012b) Synthesis and characterization of thermo- and pH-responsive bacterial cellulose/acrylic acid hydrogels for drug delivery. *Carbohydr Polym* 88:465–473
- Anin MCIM, Ahmad N, Pandey M, Xin CJ (2014) Stimuli-responsive bacterial cellulose-g-poly (acrylic acid-co-acrylamide) hydrogels for oral controlled release drug delivery. *Drug Dev Ind Pharm* 40:1340–1349
- Annuaikit T, Chusuit T, Rakanan P, Boonme P (2011) Effects of a cellulose based synthesized by a bacterium on facial skin characteristics and user satisfaction. *Med Devices* 4:77–81
- Aramwit P, Bang N (2014) The characteristics of bacterial nanocellulose gel releasing silk sericin for facial treatment. *BMC Biotechnol* 14:104
- Benbow M, Stevens J (2010) Exudate, infection and patient quality of life. *Br J Nur* 19:S32–S36
- Bodhibukkana C, Srichana T, Kaewnopparat S, Tangthong N, Bouking P, Martin GP, Suedee R (2006) Composite delivery of bacterially-derived cellulose and molecularly imprinted polymer for use as a transdermal enantioselective controlled-release system of racemic propranolol. *J Control Rel* 113:43–56
- Bodin A, Concaro S, Britberg M, Gatenholm P (2007) Bacterial cellulose as a potential meniscus implant. *J Tissue Eng Regen Med* 1:406–408
- Brown AJ (1886a) XIX.—The chemical action of pure cultivations of *bacterium aceti*. *J Chem Soc Trans* 49:172–187
- Brown AJ (1886b) XLIII.—On an acetic ferment which forms cellulose. *J Chem Soc Trans* 49:432–439
- Bruno BJ, Miller GD, Lim CS (2013) Basics and recent advances in peptide and protein drug delivery. *Ther Deliv* 4:1443–1467
- Butchosa N, Brown C, Larsson PT, Berglund LA, Bulone V, Zhou Q (2013) Nanocomposites of bacterial cellulose nanofibers and chitin nanocrystals: fabrication, characterization and bactericidal activity. *Green Chem* 15:3404–3413
- Campano C, Balea A, Blanco A, Negro C (2015) Enhancement of the fermentation process and properties of bacterial cellulose: a review. *Cellulose* 23:1–35
- Chau C-F, Yang P, Yu C-M, Yen G-C (2008) Investigation on the lipid-and cholesterol-lowering abilities of biocellulose. *J Agric Food Chem* 56:2291–2295
- Chawla PR, Bajaj JB, Survasse SA, Singhal RS (2009) Microbial cellulose: fermentative production and applications. *Food Technol Biotechnol* 47:107–124
- Chen X, Chen Z, Zhu J, Xu C, Yan W, Yao C (2011) A novel H₂O₂ amperometric biosensor based on gold nanoparticles/self-doped polyaniline nanofibers. *Bioelectrochemistry* 82:87–94
- Chen L, Zou M, Hong FF (2015) Evaluation of fungal laccase immobilized on natural nanostructured bacterial cellulose. *Front Microbiol* 6:1245
- Czaja W, Romanovicz D, Brown RM (2004) Structural investigations of microbial cellulose produced in stationary and agitated culture. *Cellulose* 11:403–411
- Czaja W, Krystynowicz A, Bielecki S, Brown RM (2006) Microbial cellulose—the natural power to heal wounds. *Biomaterials* 27:145–151
- Czaja WK, Young DJ, Kawecki M, Brown RM (2007) The future prospects of microbial cellulose in biomedical applications. *Biomacromolecules* 8:1–12
- Darbre PD, Harvey PW (2008) Paraben esters: review of recent studies of endocrine toxicity, absorption, esterase and human exposure, and discussion of potential human health risks. *J Appl Toxicol* 28:561–578
- De Groot AS, Martin W (2009) Reducing risk, improving outcomes: bioengineering less immunogenic protein therapeutics. *Clin Immunol* 131:189–201
- Dobre ML, Stoica-Guzun A (2013) Antimicrobial Ag-poly(vinyl alcohol)-bacterial cellulose composite films. *J Biobased Mater Bioenergy* 7:157–162
- Dobre L-M, Stoica-Guzun A, Stroescu M, Jipa I, Dobre T, Ferdeş M, Ciompilea Ş (2012) Modelling of sorbic acid diffusion through bacterial cellulose-based antimicrobial films. *Chem Pap* 66:144–151
- Draeolz Z, Hornby S, Walters RM, Appa Y (2013) Hydrophobically modified polymers can minimize skin irritation potential caused by surfactant-based cleansers. *J Cosmet Dermatol* 12:314–321
- Dufresne A (2013) Nanocellulose: from nature to high performance tailored materials. *Walter de Gruyter, Berlin*
- Ellis B, Smith R (2008) *Polymers: a property database*. CRC Press, Boca Raton
- Fernandes P (2010) Enzymes in food processing: a condensed overview on strategies for better biocatalysts. *Enzyme Res*. [862537](https://doi.org/10.1080/00141801.2010.500000) Article ID 862537

<http://dx.doi.org/10.4061/2010/862537>

AQ3

- Fu L, Zhang Y, Li C, Wu Z, Zhuo Q, Huang X, Qiu G, Zhou P, Yang G (2012) Skin tissue repair materials from bacterial cellulose by a multilayer fermentation method. *J Mater Chem* 22:12349–12357
- Gayathry G, Gopalaswamy G (2014) Production and characterisation of microbial cellulosic fibre from *Acetobacter xylinum*. *Indian J Fibre Text Res* 39:93–96
- Haemmerle G, Signer M, Mittleboeck M (2012) Comparison of PHMB-containing dressing and silver dressings in patients with critically colonised or locally infected wounds. *J Wound Care* 21:13–19
- Harris JM, Chess RB (2003) Effect of pegylation on pharmaceuticals. *Nat Rev Drug Discov* 2:214–221
- Hasan N, Biak DRA, Kamrudin S (2012) Application of bacterial cellulose (BC) in natural facial scrub. *IJASEIT* 2:1–4
- Heath BP, Coffindaffer TW, Kyte III KE, Smith III ED, McConaughy SD (2012) Personal cleansing compositions comprising a bacterial cellulose network and cationic polymer. US patent, US 8097574 B2
- Helenius G, Bäckdahl H, Bodin A, Nannmark U, Gatenholm P, Risberg B (2006) *In vivo* biocompatibility of bacterial cellulose. *J Biomed Mater Res A* 76:431–438
- Huang L, Chen X, Nguyen TX, Tang H, Zhang L, Yang G (2013) Nano-cellulose 3D-networks as controlled-release drug carriers. *J Mater Chem B* 1:2976–2984
- Hubbell JA (1995) Biomaterials in tissue engineering. *Nat Biotechnol* 13:565–576
- Hui J, Yuanjuan J, Jiao W, Yuan H, Yuan Z, Shiru J (2009) Potentiality of bacterial cellulose as the scaffold of tissue engineering of cornea. In: Biomedical engineering and informatics, 2009. 2nd international conference, IEEE, Tianjin, China, pp 1–5
- Hussain MA, Badshah M, Iqbal MS, Tahir MN, Tremel W, Bhosale SV, Sher M, Haseeb MT (2009) HPMC-salicylate conjugates as macromolecular prodrgs: design, characterization, and nano-rods formation. *J Polym Sci Part A Polym Chem* 47:4202–4208
- Iguchi M, Yamataka S, Budhiono A (2000) Bacterial cellulose—a masterpiece of nature's arts. *J Mater Sci* 35:261–270
- Jay JM, Loessner MJ, Golden DA (2008) Modern food microbiology. Springer, New York
- Jipa IM, Stoica-Guzun A, Stroescu M (2012) Controlled release of sorbic acid from bacterial cellulose based mono and multilayer antimicrobial films. *LWT Food Sci Technol* 47:400–406
- Júzová P, Martinková L, Křen V (1996) Secondary metabolites of the fungus *Monascus*: a review. *J Ind Microbiol* 16:163–170
- Kaliyaperumal A, Jing S (2009) Immunogenicity assessment of therapeutic proteins and peptides. *Curr Pharm Biotechnol* 10:352–358
- Kaplan E, Ince T, Yorulmaz E, Yener F, Harputlu E, Laçin NT (2014) Controlled delivery of ampicillin and gentamycin from cellulose hydrogels and their antibacterial efficiency. *J Biomater Tissue Eng* 4:543–549
- Khan T, Park JK, Kwon J-H (2007) Functional biopolymers produced by biochemical technology considering applications in food engineering. *Korean J Chem Eng* 24:816–826
- Khan S, Ul-Islam M, Khatkhat WA, Ullah MW, Park JK (2015) Bacterial cellulose-titanium dioxide nanocomposites: nanostructural characteristics, antibacterial mechanism, and biocompatibility. *Cellulose* 22:565–579
- Kilara A, Shahani KM, Shukla TP (1979) The use of immobilized enzymes in the food industry: a review. *Crit Rev Food Sci Nutr* 12:161–198
- Kirdponattara S, Phisalaphong M (2013) Bacterial cellulose-alginate composite sponge as a yeast cell carrier for ethanol production. *Biochem Eng J* 77:103–109
- Klemm D, Schumann D, Uthardt U, Marsch S (2001) Bacterial synthesized cellulose—artificial blood vessels for microsurgery. *Prog Polym Sci* 26:1561–1603
- Klemm D, Heublein B, Fink HP, Bohn A (2005) Cellulose: fascinating biopolymer and sustainable raw material. *Angew Chem Int Ed* 44:3358–3393
- Klemm D, Schumann D, Kramer F, Hebler N, Hornung M, Schmauder H-P, Marsch S (2006) Nanocelluloses as innovative polymers in research and application. *Adv Polym Sci* 295:49–96
- Koochi MK, Hejaz M, Asadi F, Asadian P (2011) Assessment of dermal exposure and histopathologic changes of different sized nano-silver in healthy adult rabbits. *J Phys Conf Ser* 304:012028
- Korami M, Rezaayari SM, Bidgoli SA (2013) Sub-chronic dermal toxicity of silver nanoparticles in guinea pig: special emphasis to heart, bone and kidney toxicities. *Iran J Pharm Res* 12:511–519
- Koutinas AA, Sypsas V, Kandyli F, Michelis A, Bekatorou A, Kourkoutas Y, Korudis C, Lycourgiotis A, Banat IM, Nigam P, Marchant R (2012) Nano-tubular cellulose for bioprocess technology development. *PLoS ONE* 7:e34350
- Krystynowicz A, Czaja W, Wiktorowska-Jeziarska A, Goncalves-Miskiewicz M, Turkiewicz M, Bielecki S (2002) Factors affecting the yield and properties of bacterial cellulose. *J Ind Microbiol Biotechnol* 29:189–195
- Kuehl BL, Fyfe KS, Shear NH (2003) Cutaneous cleansers. *Skin Ther Lett* 8(3):1–4
- Lee CK, Hsu KC, Cho JC, Kim YJ, Han SH (2011) Cosmetic bio-cellulose mask pack sheet and method for manufacturing same. US patent, US 20130244977 A1
- Legendre JY (2008) Assembly comprising a substrate comprising biocellulose, and a powdered cosmetic composition to be brought into contact with the substrate. US patent, US 20090041815 A1
- Levinson DJ, Glonek T (2010) Microbial cellulose contact lens. US patents, US 7832857 B2
- Li X, Wan W, Panchal CJ (2010) Transparent bacterial cellulose nanocomposite hydrogels. US patent, US 8940337 B2
- Lin K, Lin H (2004) Quality characteristics of chinese-style meatball containing bacterial cellulose (nata). *J Food Sci* 69:SNQ107–SNQ111
- Lin SP, Calvar IL, Catchmark JM, Liu JR, Demirci A, Cheng KC (2013) Biosynthesis, production and applications of bacterial cellulose. *Cellulose* 20:2191–2219
- Lin SP, Hsieh SC, Chen KL, Demirci A, Cheng KC (2014) Semi-continuous bacterial cellulose production in a rotating disk bioreactor and its materials properties analysis. *Cellulose* 21:835–844
- Lin Y-C, Wey Y-C, Lee M-L, Lin P-C (2015) Cosmetic composition containing fragments of bacterial cellulose film and method for manufacturing thereof. US patent, US 20150216784 A1
- Lin SP, Liu CT, Hsu KD, Hung YT, Shih TY, Cheng KC (2016) Production of bacterial cellulose with various additives in a PCS rotating disk bioreactor and its material property analysis. *Cellulose* 3:367–377
- Lu W, Senapati D, Wang S, Tovmachenko O, Singh AK, Yu H, Ray PC (2010) Effect of surface coating on the toxicity of silver nanomaterials on human skin keratinocytes. *Chem Phys Lett* 487:92–96
- Luan J, Wu J, Zheng Y, Song W, Wang G, Gao J, Ding X (2012) Impregnation of silver sulfadiazine into bacterial cellulose for antimicrobial and biocompatible wound dressing. *Biomed Mater* 7:065006
- Lu P, Feng Q, Wang Q, Li G, Li D, Wei Q (2016) Biosynthesis of bacterial cellulose/carboxylic multi-walled carbon nanotubes for enzymatic biofuel cell application. *Materials* 9:183
- Malik NN (2008) Drug discovery: past, present and future. *Drug Discov Today* 13:909–912
- Manning MC, Patel K, Borchardt RT (1989) Stability of protein pharmaceuticals. *Pharm Res* 6:903–918
- Manning MC, Chou DK, Murphy BM, Payne RW, Katayama DS (2010) Stability of protein pharmaceuticals: an update. *Pharm Res* 27:544–575
- Millon L, Wan W (2006) The polyvinyl alcohol–bacterial cellulose system as a new nanocomposite for biomedical applications. *J Biomed Mater Res B Appl Biomater* 79:245–253
- Monteleagre CM, Dionisio ER, Sumera LV, Adolacion JR, De Leon RL (2012) A comparison between the performance of *S. cerevisiae* cells immobilized in nata de coco biocellulose and calcium alginate during continuous bioethanol production. *Int J Chem Eng Appl* 3:237–242
- Mori R, Nakai T, Enomoto K, Uchio Y, Yoshino K (2011) Increased antibiotic release from a bone cement containing bacterial cellulose. *Clin Orthop Relat Res* 469:600–606
- Müller A, Ni Z, Hessler N, Wesarg F, Müller FA, Kralisch D, Fischer D (2013) The biopolymer bacterial nanocellulose as drug delivery system: investigation of drug loading and release using the model protein albumin. *J Pharm Sci* 102:579–592
- Murphy O (2001) Non-polyol low-digestible carbohydrates: food applications and functional benefits. *Br J Nutr* 85:547–553
- Nagel JE, Fuscaldo JT, Fireman P (1977) Paraben allergy. *JAMA* 237:1594–1595
- Ng C-C, Shyu Y-T (2004) Development and production of cholesterol-lowering *Monascus*-nata complex. *World J Microbiol Biotechnol* 20:875–879
- Nguyen LA, He H, Pham-Huy C (2006) Chiral drugs: an overview. *Int J Biomed Sci* 2:85–100
- Nguyen DN, Ton NMN, Le VVM (2009) Optimization of *Saccharomyces cerevisiae* immobilization in bacterial cellulose by 'adsorption-incubation' method. *Int Food Res J* 16:59–64
- Nimeskern L, Avila HM, Sundberg J, Gatenholm P, Müller R, Stok KS (2013) Mechanical evaluation of bacterial nanocellulose as an implant material for ear cartilage replacement. *J Mech Behav Biomed Mater* 22:12–21
- Okiyama A, Motoki M, Yamataka S (1992) Bacterial cellulose II. Processing of the gelatinous cellulose for food materials. *Food Hydrocol* 6:479–487
- Okiyama A, Motoki M, Yamataka S (1993) Bacterial cellulose IV. Application to processed foods. *Food Hydrocol* 6:503–511
- Olyveira GM, Costa LMM, Basmuji P (2013) Physically modified bacterial cellulose as alternative routes for transdermal drug delivery. *J Biomater Tissue Eng* 3:227–232
- Osmi JF, Toca-Herrera JL, Rodríguez-Couto S (2010) Uses of laccases in the food industry. *Enzyme Res*. [9:487-491](https://doi.org/10.4061/2010/918761) Article ID 918761
- <http://dx.doi.org/10.4061/2010/918761>
- Ougiya H, Watanabe K, Morinaga Y, Yoshinaga F (1997) Emulsion-stabilizing effect of bacterial cellulose. *Biosci Biotechnol Biochem* 61:1541–1545
- Pandey M, Mohd Amin MCIM, Ahmad N, Aber MM (2013) Rapid synthesis of superabsorbent smart-swelling bacterial cellulose/acrylamide-based hydrogels for drug delivery. *Int J Polym Sci*. [905471](https://doi.org/10.1155/2013/905471) Article ID 905471
- <http://dx.doi.org/10.1155/2013/905471>
- Pandey M, Mohamad N, Amin MCIM (2014) Bacterial cellulose/acrylamide pH-sensitive smart hydrogel: development, characterization, and toxicity studies in ICR mice model. *Mol Pharm* 11:3596–3608
- Park JK, Khan T, Jung JY (2009) Bacterial Cellulose. In: Phillips GO, Williams PA (eds) Handbook of hydrocolloids. Woodhead Publishing Ltd., Abington, pp 724–739
- Pavaloiu R-D, Stoica-Guzun A, Stroescu M, Jinga SI, Dobre T (2014a) Composite films of poly(vinyl alcohol)–chitosan–bacterial cellulose for drug controlled release. *Int J Biol Macromol* 68:117–124
- Pavaloiu R-D, Stoica A, Stroescu M, Dobre T (2014b) Controlled release of amoxicillin from bacterial cellulose membranes. *Cent Eur J Chem* 12:962–967
- Pavaloiu R-D, Stroescu M, Parvulescu O, Dobre T (2014c) Composite hydrogels of bacterial cellulose-carboxymethyl cellulose for drug release. *Rev Chim Buchar* 65:948–951
- Păvăloiu R-D, Stoica-Guzun A, Dobre T (2015) Swelling studies of composite hydrogels based on bacterial cellulose and gelatin. *UPB Sci Bull Ser B* 77:54–62

- Peng Y-S, Lin S-C, Huang S-J, Wang Y-M, Lin Y-J, Wang L-F, Chen J-S (2006) Chondroitin sulfate-based anti-inflammatory macromolecular prodrugs. *Eur J Pharm Sci* 29:60–69
- Peppas NA, Wood KM, Blanchette JO (2004) Hydrogels for oral delivery of therapeutic proteins. *Expert Opin Biol Ther* 4:881–887
- Petersen N, Gatenholm P (2011) Bacterial cellulose-based materials and medical devices: current state and perspectives. *Appl Microbiol Biotechnol* 91:1277–1286
- Phisalaphong M, Chiaoapakobkij N (2012) Applications and products—nata de coco. In: Gama M, Gatenholm P, Klemm D (eds) *Bacterial nanocellulose: a sophisticated multifunctional material*. CRC Press, Boca Raton, pp 143–156
- Pinto RJ, Daina S, Sadocco P, Pascoal Neto C, Trindade T (2013) Antibacterial activity of nanocomposites of copper and cellulose. *Biomed Res Int*. [280512](https://doi.org/10.1155/2013/280512) Article ID 280512 <http://dx.doi.org/10.1155/2013/280512>
- Pircher N, Veigel S, Aigner N, Nedelez JM, Rosenau T, Liebner F (2014) Reinforcement of bacterial cellulose aerogels with biocompatible polymers. *Carbohydr Polym* 111:505–513
- Prabhu BM, Ali SF, Murdock RC, Hussain SM, Srivatsan M (2010) Copper nanoparticles exert size and concentration dependent toxicity on somatosensory neurons of rat. *Nanotoxicology* 2010:150–160
- Prausnitz MR, Langer R (2008) Transdermal drug delivery. *Nat Biotechnol* 26:1261–1268
- Prausnitz MR, Mitragotri S, Langer R (2004) Current status and future potential of transdermal drug delivery. *Nat Rev Drug Discov* 3:115–124
- Purwadaria T, Ganawan L, Agustini GW (2010) The production of nata colored by *Monascus purpureus* J1 pigments as functional foods. *Microbiol Indones* 4:6–10
- Qiu K, Netravali AN (2014) A review of fabrication and applications of bacterial cellulose based nanocomposites. *Polym Rev* 54:598–626
- Radi ZA, Khan NK (2006) Effects of cyclooxygenase inhibition on the gastrointestinal tract. *Exp Toxicol Pathol* 58:163–173
- Ratner BD, Bryant SJ (2004) Biomaterials: where we have been and where we are going. *Annu Rev Biomed Eng* 6:41–75
- Ray PC, Yu H, Fu PP (2009) Toxicity and environmental risks of nanomaterials: challenges and future needs. *J Environ Sci Health A* 27:1–35
- Ross P, Mayer R, Benziman M (1991) Cellulose biosynthesis and function in bacteria. *Microbiol Rev* 55:35–58
- Rouahbia M, Asselin J, Tazi N, Messaddeq Y, Levinson D, Zhang Z (2014) Production of biocompatible and antimicrobial bacterial cellulose polymers functionalized by RGDC grafting groups and gentamicin. *ACS Appl Mater Interfaces* 6:1439–1446
- Rubinstein MP (2003) Applications of contact lens devices in the management of corneal disease. *Eye* 17:872–876
- Sanberg ME, Oldenburg SJ, Monteiro-Riviere NA (2010) Evaluation of silver nanoparticle toxicity in skin *in vivo* and keratinocytes *in vitro*. *Environ Health Perspect* 118:407–413
- Saxena IM, Brown RM Jr (2012) Biosynthesis of bacterial cellulose. In: Gama M, Gatenholm P, Klemm D (eds) *Bacterial nanocellulose: a sophisticated multifunctional material*. CRC Press, Boca Raton, pp 1–18
- Serafica G, Mormino R, Oster GA, Lentz KE, Koehler KP (2010) Microbial cellulose wound dressing for treating chronic wounds. US patent, US 7704523 B2
- Shezad O, Khan S, Khan T, Park JK (2010) Physicochemical and mechanical characterization of bacterial cellulose produced with an excellent productivity in static conditions using a simple fed-batch cultivation strategy. *Carbohydr Polym* 82:173–180
- Shi Q, Li Y, Sun J, Zhang H, Chen L, Chen B, Yang H, Wang Z (2012) The osteogenesis of bacterial cellulose scaffold loaded with bone morphogenetic protein-2. *Biomaterials* 33:6644–6649
- Shi X, Zheng Y, Zhang W, Zhang Z, Peng Y (2013) A novel drug carrier based on functional modified nanofiber cellulose and the control release behavior. In: Fourth international conference on smart materials and nanotechnology in engineering. International society for optics and photonics, Gold Coast, Australia, pp 879304–879306
- Shi X, Zheng Y, Wang G, Lin Q, Fan J (2014a) pH-and electro-response characteristics of bacterial cellulose nanofiber/sodium alginate hybrid hydrogels for dual controlled drug delivery. *RSC Adv* 4:47056–47065
- Shi Z, Zhang Y, Phillips GO, Yang G (2014b) Utilization of bacterial cellulose in food. *Food Hydrocoll* 35:539–545
- Shoichet MS (2009) Polymer scaffolds for biomaterials applications. *Macromolecules* 43:581–591
- Silva NH, Rodrigues AF, Almeida IF, Costa PC, Rosado C, Neto CP, Silvestre AJ, Freire CS (2014) Bacterial cellulose membranes as transdermal delivery systems for diclofenac: in vitro dissolution and permeation studies. *Carbohydr Polym* 106:264–269
- Siró I, Plackett D (2010) Microfibrillated cellulose and new nanocomposite materials: a review. *Cellulose* 17:459–494
- Steinemann TL, Fletcher M, Bonny AE, Harvey RA, Hamlin D, Zloty P, Besson M, Walter K, Gagnon M (2005) Over-the-counter decorative contact lenses: cosmetic or medical devices? A case series. *Eye Contact Lens* 31:194–200
- Stenstad P, Andresen M, Tanem BS, Stenius P (2008) Chemical surface modifications of microfibrillated cellulose. *Cellulose* 15:35–45
- Stephens RS, Westland JA, Neogi AN (1990) Method of using bacterial cellulose as a dietary fiber component. US patent, US 4960763 A
- Stoica-Gazun A, Stroescu M, Tache F, Zaharescu T, Grosu E (2007) Effect of electron beam irradiation on bacterial cellulose membranes used as transdermal drug delivery systems. *Nucl Instrum Methods Phys Res B* 265:434–438
- Sulueva I, Henniges U, Rosenau T, Potthast A (2015) Bacterial cellulose as a material for wound treatment: properties and modifications. A review. *Biotechnol Adv* 33:1547–1571
- Svensson A, Nicklasson E, Hamah T, Panilaitis B, Kaplan D, Brittberg M, Gatenholm P (2005) Bacterial cellulose as a potential scaffold for tissue engineering of cartilage. *Biomaterials* 26:419–431
- Tam TTM, Huang NT (2014) Optimization of *Corynebacterium glutamicum* immobilization process on bacterial cellulose carrier and its application for lysine fermentation. *IOSR JEN* 4:33–38
- Tang L, Persky AM, Hochhaus G, Meibohm B (2004) Pharmacokinetic aspects of biotechnology products. *J Pharm Sci* 93:2184–2204
- Tomé LC, Brandão L, Mendes AM, Silvestre AJ, Neto CP, Gandini A, Freire CS, Marrucho IM (2010) Preparation and characterization of bacterial cellulose membranes with tailored surface and barrier properties. *Cellulose* 17:1203–1211
- Ton N, Le V (2011) Application of immobilized yeast in bacterial cellulose to the repeated batch fermentation in wine-making. *Int Food Res J* 18:983–987
- Tournilhac F, Lorant R (2003) Composition in the form of an oil-in-water emulsion containing cellulose fibrils, and its uses, especially cosmetic uses. US patent, US 6534071 B1
- Trovatti E, Freire CS, Pinto PC, Almeida IF, Costa P, Silvestre AJ, Neto CP, Rosado C (2012) Bacterial cellulose membranes applied in topical and transdermal delivery of lidocaine hydrochloride and ibuprofen: in vitro diffusion studies. *Int J Pharm* 435:83–87
- Ul-blam M, Khan T, Khattak WA, Park JK (2013) Bacterial cellulose-MMTs nanoreinforced composite films: novel wound dressing material with antibacterial properties. *Cellulose* 20:589–596
- Vermondten T, Censi R, Hennink WE (2012) Hydrogels for protein delivery. *Chem Rev* 112:2853–2888
- Vowden K, Vowden P (2003) Understanding exudate management and the role of exudate in the healing process. *Br J Community Nurs* 8:S4–S13
- Walters RM, Mao G, Gunn ET, Hornby S (2012) Cleansing formulations that respect skin barrier integrity. *Dermatol Res Pract*. [495917](https://doi.org/10.1155/2012/495917) Article ID 495917 <http://dx.doi.org/10.1155/2012/495917>
- Wan Y, Gao C, Han M, Liang H, Ren K, Wang Y, Luo H (2011) Preparation and characterization of bacterial cellulose/heparin hybrid nanofiber for potential vascular tissue engineering scaffolds. *Polym Adv Technol* 22:2643–2648
- Wang LP, Wang JY (2014) Skin penetration of inorganic and metallic nanoparticles. *J Shanghai Jiaotong Univ (Sci)* 19:691–697
- Wang W, Li HY, Zhang DW, Jiang J, Cui YR, Qiu S, Zhou YL, Zhang XX (2010) Fabrication of biozymatic glucose biosensor based on novel gold nanoparticles-bacteria cellulose nanofibers nanocomposite. *Electroanalysis* 22:2543–2550
- Wang T, Long X, Cheng Y, Liu Z, Yan S (2014) The potential toxicity of copper nanoparticles and copper sulphate on juvenile *Epinephelus coioides*. *Aquat Toxicol* 152:96–104
- Wanling Z, Zhe L, Zerui Z, Bihui Z, Shiyuan C, Huaping W, Wen Z (2012) Preparation method for anti-virus bacteria cellulose protective. CN patent, 102321261 A
- Wei B, Yang G, Hong F (2011) Preparation and evaluation of a kind of bacterial cellulose dry films with antibacterial properties. *Carbohydr Polym* 84:533–538
- Wen X, Zheng Y, Wu J, Yue L, Wang C, Luan J, Wu Z, Wang K (2015) *In vitro* and *in vivo* investigation of bacterial cellulose dressing containing uniform silver sulfadiazine nanoparticles for burn wound healing. *Prog Nat Sci* 25:197–203
- Wonganu B, Kongruang S. (2010) Red bacterial cellulose production by fermentation of *Monascus purpureus*. In: Chemistry and chemical engineering (CCCE), international conference, IEEE, Kyoto, Japan, pp 137–141
- Wright JM (2002) The double-edged sword of COX-2 selective NSAIDs. *CMAJ* 167:1131–1137
- Wu S-C, Lia Y-K (2008) Application of bacterial cellulose pellets in enzyme immobilization. *J Mol Catal B Enzym* 54:103–108
- Wu S-C, Lia Y-K, Ho C-Y (2013) Glucoamylase immobilization on bacterial cellulose using periodate oxidation method. *IJSE* 3:1–4
- Yadav V, Panilaitis BJ, Shi H, Lee K, Cebe P, Kaplan DL (2010) Novel *in vitro*-degradable cellulose-chitin copolymer from metabolically engineered *Gluconacetobacter xylius*. *Appl Environ Microbiol* 76:6257–6265
- Yang M (2015) Stress and protein instability during formulation and fill/finish processes. *BioPharm Int* 28:46–49
- Yano H, Sugiyama J, Nakagaito AN, Nogi M, Matsuura T, Híkita M, Handa K (2005) Optically transparent composites reinforced with networks of bacterial nanofibers. *Adv Mater* 17:153–155
- Yoshino A, Tabuchi M, Uo M, Tatsumi H, Hideshima K, Kondo S, Sekine J (2013) Applicability of bacterial cellulose as an alternative to paper points in endodontic treatment. *Acta Biomater* 9:6116–6122
- Zhang T, Wang W, Zhang D, Zhang X, Ma Y, Zhou Y, Qi L (2010) Biotemplated synthesis of gold nanoparticle-bacteria cellulose nanofiber nanocomposites and their application in biosensing. *Adv Funct Mater* 20:1152–1160
- Zhong CY (2008) Bacterial cellulose gel face mask. CN patent, 200610075040.8
- Zimmermann KA, LeBlanc JM, Sheets KT, Fox RW, Gatenholm P (2011) Biomimetic design of a bacterial cellulose/hydroxyapatite nanocomposite for bone healing applications. *Mater Sci Eng C* 31:43–49